

# Comparison of Biotage® Extrahera™ vs. Manual Sample Processing Using a Vacuum Manifold

## Extraction of 25-OH Vitamin D from Plasma Using ISOLUTE® SLE+

Automated sample preparation using the Biotage® Extrahera™ was compared to an equivalent manual method utilizing a vacuum manifold. Analytes were extracted from pooled stripped plasma using a supported liquid extraction procedure. ISOLUTE® SLE+ 400 µL sample volume plates, part number 820-0400-P01 were used for extraction.

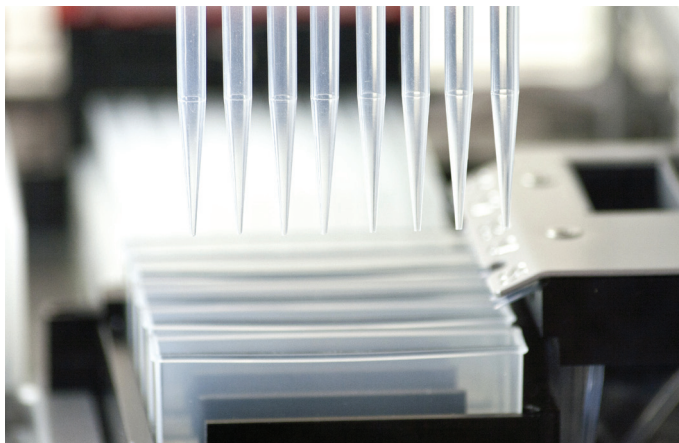
Resulting extracts from both sample preparation methods were subsequently analyzed by LC-MS/MS.



## Procedure

A pooled plasma sample was prepared in a sufficient quantity to run a full 96-well plate for each processing method. This pooled plasma sample was fortified with 25-OH Vitamin D2 and D3 at a concentration of 30 ng/mL respectively. 25-OH Vitamin D3-d6 was added as an internal standard at a concentration of 30 ng/mL.

From this pooled plasma sample 200 µL was transferred to all wells of two 96-well plates.



**All subsequent aspects of sample preparation were performed in duplicate on two separate plates utilizing either Extrahera or manual preparation using a calibrated air-displacement pipette.**

The pooled plasma sample was then pre-treated 1:1 (v/v) with Water:Propan-2-ol 1:1 (v/v) (200 µL).

After pre-wetting the pipette tips via aspirate/dispense cycling and to mix the samples, 300 µL of the pre-treated sample was loaded to each well of the ISOLUTE® SLE+ plates. Flows were initiated using a pulse of positive pressure (Extrahera) or vacuum (manual method).

After leaving for 5 minutes to allow the sample to completely absorb into the plates, elution was performed by the application of 2 x 750 µL of Heptane to the ISOLUTE® SLE+ plates.

The extracts were collected in 2 mL 96-well collection plates under gravity elution, and as a final step to recover all available solvent from the media, by applying a pulse of positive pressure (Extrahera) or vacuum (manual method).

The extracts were evaporated to dryness in a TurboVap® 96 at 37 °C or a SPE Dry at 40 °C and reconstituted in 100 µL of 30:70 (v/v) water/methanol solution.

The plates were mixed on an orbital shaker for 10 minutes.

## HPLC Conditions

<b>Instrument:</b>	Waters Aquity UHPLC
<b>Column:</b>	Restek Pinnacle DB BiPhenyl, 50 mm x 2.1 mm 1.9 µm
<b>Mobile Phase:</b>	80:20 (v/v) 2mM ammonium formate with 0.1 % formic acid (aq.)/ Methanol with 0.1 % formic acid at 0.4 mL/min
<b>Injection Volume:</b>	15 µL

## Mass Spectrometry

<b>Instrument:</b>	Waters Quattro Premier XE
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### MRM Conditions

Analyte	Transition	RT (min)	Dwell (sec)	Cone (V)	Col Energy (V)
25-OH Vitamin D2	395.5 to 119.2	1.7	0.1	30	26
25-OH Vitamin D3	383.5 to 107.2	1.6	0.1	30	25
25-OH Vitamin D3-d6	389.6 to 263.5	1.5	0.1	30	16

## Results

Average peak area data was calculated for all three compounds to compare any improvements in analyte recovery between Biotage® Extrahera™ and manually processed samples.

Peak area ratio data was also generated for all samples by referencing the analyte vs. 25-OH Vitamin D3-d6. This provides standardized data to allow a comparison of the % RSD of the Extrahera vs. manual data sets.

	Extrahera Peak Area Ratio Summary	Manual Peak Area Ratio Summary
Average 25-OH D2 Peak Area	1697	1577
Improvement (%) vs. Manual Method	7.6	-
Average 25-OH D3-d6 Peak Area	1441	1384
Improvement (%) vs. Manual Method	4.1	-
Average Peak Area Ratio	1.1771	1.1421
% RSD of Extrahera Extraction	7.7	7.5
Improvement (%) vs. Manual Method	-3.6	-

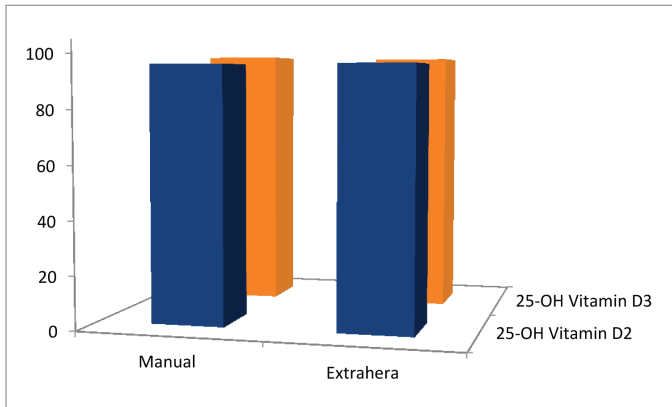
	Extrahera Peak Area Ratio Summary	Manual Peak Area Ratio Summary
Average 25-OH D3 Peak Area	4758	4528
Improvement (%) vs. Manual Method	5.1	-
Average 25-OH D3-d6 Peak Area	1441	1384
Improvement (%) vs. Manual Method	4.1	-
Average Peak Area Ratio	3.3051	3.2813
% RSD of Extrahera Extraction	5.9	6.7
Improvement (%) vs. Manual Method	11.8	-

## Experimental Precautions

The following precautions were performed to minimize differences between the manual and Extrahera extracted plates.

- » Both plates were evaporated side by side on the same evaporation instrument (TurboVap® 96).
- » During analysis on the LC-MS system samples were injected alternately from the two plates to reduce the effect of any sample stability issues.
- » The same batch/bottles of samples, reagents and solvents were used for both methods.

Additional experiments were also completed using serum as the sample matrix. The average recovery comparison data is presented in **Figure 1** below.



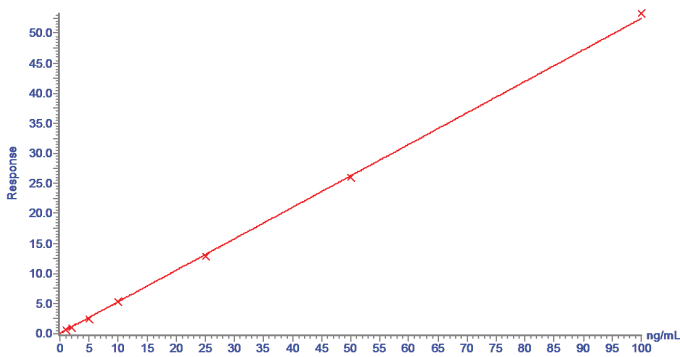
**Figure 1.** Average recovery (n=8) of 25-OH-Vitamin D2 and 25-OH-Vitamin D3 from serum

Calibration series samples prepared manually in Phosphate Buffered Saline with Bovine Serum Albumin (PBS-BSA) and those commercially available from Chromsystems were also processed and extracted using the procedure above with both Biotage® Extrahera™ and manual sample processing methods.

Calibration curves for both manual and Extrahera processed samples over concentration range 1–100 ng/mL for 25-OH Vitamin D2 and D3 in Phosphate Buffered Saline with Bovine Serum Albumin (PBS-BSA) are shown in **Figures 2 to 5**.

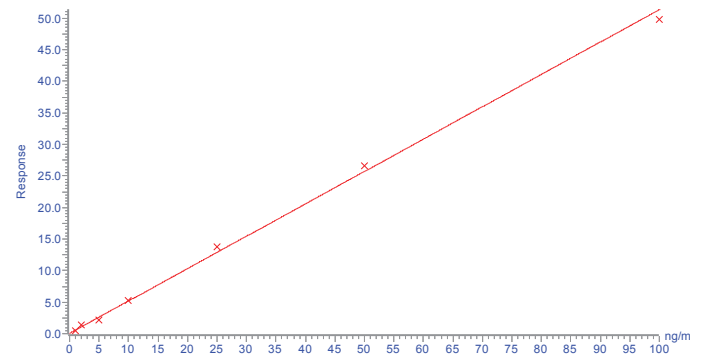
Chromsystems calibration curves for both manual and Extrahera processed samples for 25-OH Vitamin D2 (15.8–61.6 ng/mL) and D3 (4.5–66.7 ng/mL) are shown in **Figures 6 to 9**.

Compound name: 25 OH-Vitamin D2 (1)  
 Correlation coefficient:  $r = 0.999446$ ,  $r^2 = 0.998892$   
 Calibration curve:  $0.523968 * x + 0.0587074$   
 Response type: Internal Std ( Ref 3 ), Area \* ( IS Conc. / IS Area )  
 Curve type: Linear, Origin: Exclude, Weighting: 1/x, Axis trans: None



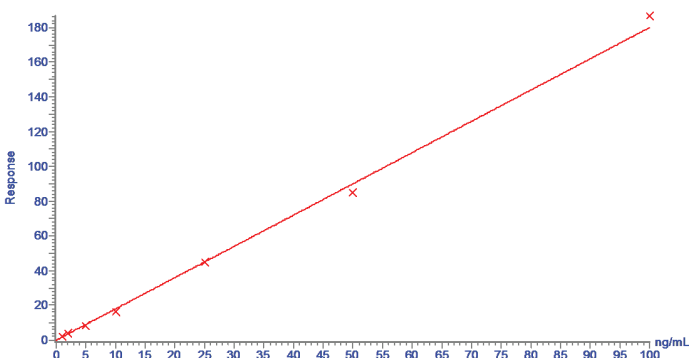
**Figure 2.** 25-OH Vitamin D2 in PBS-BSA – Biotage® Extrahera™

Compound name: 25 OH-Vitamin D2 (1)  
 Correlation coefficient:  $r = 0.998431$ ,  $r^2 = 0.996865$   
 Calibration curve:  $0.512922 * x + 0.0533979$   
 Response type: Internal Std ( Ref 3 ), Area \* ( IS Conc. / IS Area )  
 Curve type: Linear, Origin: Exclude, Weighting: 1/x, Axis trans: None



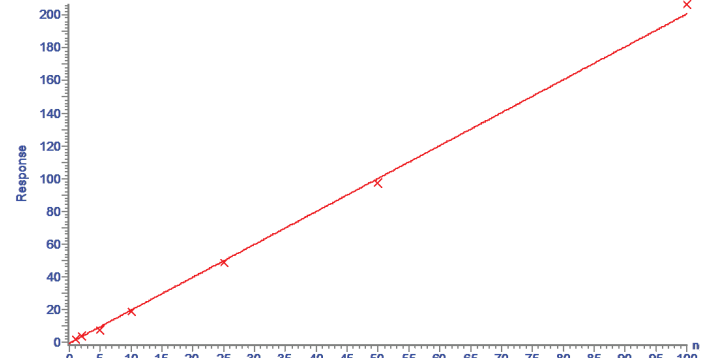
**Figure 3.** 25-OH Vitamin D2 in PBS-BSA – Manual

Compound name: 25 OH-Vitamin D3 (1)  
 Correlation coefficient:  $r = 0.998842$ ,  $r^2 = 0.997685$   
 Calibration curve:  $1.80001 * x + -0.0531757$   
 Response type: Internal Std ( Ref 3 ), Area \* ( IS Conc. / IS Area )  
 Curve type: Linear, Origin: Exclude, Weighting: 1/x, Axis trans: None



**Figure 4.** 25-OH Vitamin D3 in PBS-BSA – Biotage® Extrahera™

Compound name: 25 OH-Vitamin D3 (1)  
 Correlation coefficient:  $r = 0.999054$ ,  $r^2 = 0.998108$   
 Calibration curve:  $2.00995 * x + -0.403428$   
 Response type: Internal Std ( Ref 3 ), Area \* ( IS Conc. / IS Area )  
 Curve type: Linear, Origin: Exclude, Weighting: 1/x, Axis trans: None



**Figure 5.** 25-OH Vitamin D3 in PBS-BSA - Manual

Compound name: 25 OH-Vitamin D2 (1)  
 Correlation coefficient:  $r = 0.996517, r^2 = 0.993045$   
 Calibration curve:  $0.893101 * x + 2.43618$   
 Response type: Internal Std ( Ref 3 ), Area \* ( IS Conc. / IS Area )  
 Curve type: Linear, Origin: Exclude, Weighting: 1/x, Axis trans: None

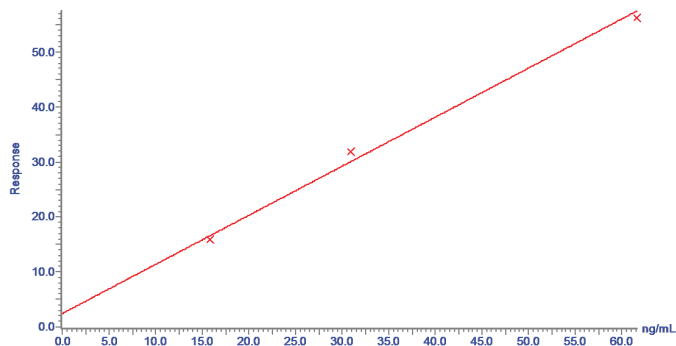


Figure 6. 25-OH Vitamin D2 Chromsystems – Biotage® Extrahera™

Compound name: 25 OH-Vitamin D2 (1)  
 Correlation coefficient:  $r = 0.996832, r^2 = 0.993675$   
 Calibration curve:  $0.757448 * x + 0.437403$   
 Response type: Internal Std ( Ref 3 ), Area \* ( IS Conc. / IS Area )  
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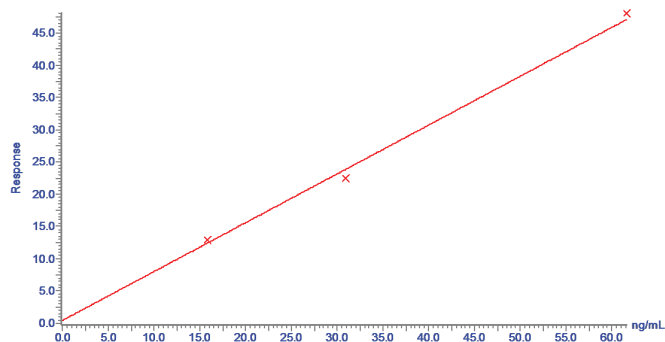


Figure 7. 25-OH Vitamin D2 Chromsystems – Manual

Compound name: 25 OH-Vitamin D3 (1)  
 Correlation coefficient:  $r = 0.999602, r^2 = 0.999205$   
 Calibration curve:  $2.42281 * x + -1.82497$   
 Response type: Internal Std ( Ref 3 ), Area \* ( IS Conc. / IS Area )  
 Curve type: Linear, Origin: Exclude, Weighting: 1/x, Axis trans: None

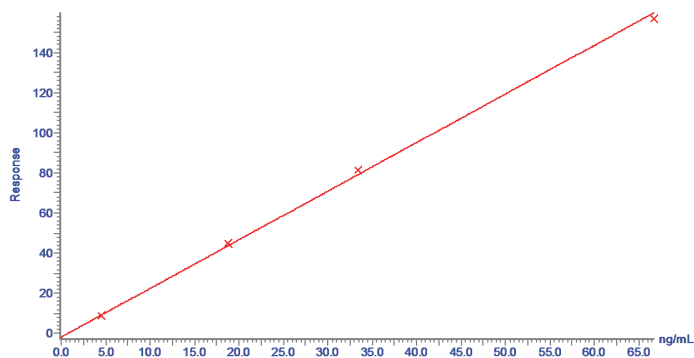


Figure 8. 25-OH Vitamin D3 Chromsystems – Biotage® Extrahera™

Compound name: 25 OH-Vitamin D3 (1)  
 Correlation coefficient:  $r = 0.998765, r^2 = 0.997533$   
 Calibration curve:  $2.04345 * x + 1.82945$   
 Response type: Internal Std ( Ref 3 ), Area \* ( IS Conc. / IS Area )  
 Curve type: Linear, Origin: Exclude, Weighting: 1/x, Axis trans: None

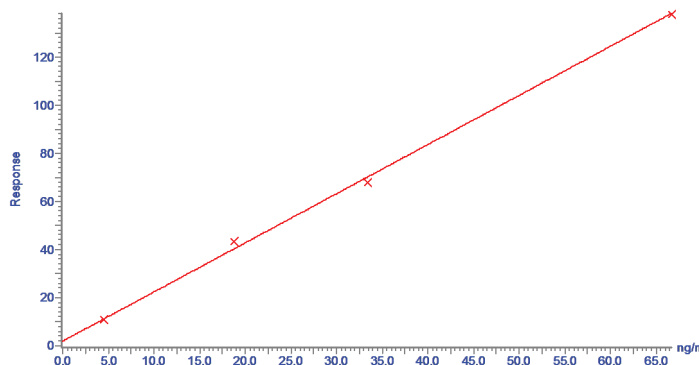


Figure 9. 25-OH Vitamin D3 Chromsystems - Manual

## Conclusion

A reduction to the % RSD was measured when using the Biotage® Extrahera™ for 25-OH Vitamin D3, the precision of the D2 data was slightly better when processed manually.

For 25-OH Vitamin D3 the % RSD improved by 11.8 %, for 25-OH Vitamin D2 there was a slight drop in the % RSD of 3.6 % to 7.47 vs. 7.47, this is not considered to be statistically significant.

The results suggest that methods performed on the Extrahera could give higher recoveries due to increases in the absolute average peak areas.

% RSD	Manual Procedure (n=93)	Extrahera (n=92)
25-OH Vitamin D2	7.47	7.74
25-OH Vitamin D3	6.70	5.91

Average Peak Area	Manual Procedure (n=93)	Extrahera (n=962)
25-OH Vitamin D2	1577	1697
25-OH Vitamin D3	4528	4758
25-OH VitaminD3-d6	1384	1441

All analytes returned average peak areas that were higher when sample extraction was performed using the Biotage® Extrahera™, than the manual method with an average peak area increase of 5 %. The greatest improvement was measured with 25-OH Vitamin D<sub>2</sub> where the average peak area was increased by 7 %.

The serum samples extracted and analyzed under the same conditions showed near identical recoveries between Extrahera and manual processing, 99 % versus 100 % for 25-OH Vitamin D<sub>3</sub> respectively and 97 % for 25-OH Vitamin D<sub>2</sub> for both methods.

The calibration series data for samples prepared in PBS-BSA and the Chromsystems calibrators both returned good correlation coefficients with  $r^2$  greater than 0.99 for samples prepared using Extrahera and processed manually.



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