

# Determination of Fatty Acids in Foods Using Gas Chromatography with Positive-ion Chemical Ionization Tandem Mass Spectrometry

ASMS 2013 ThP-201

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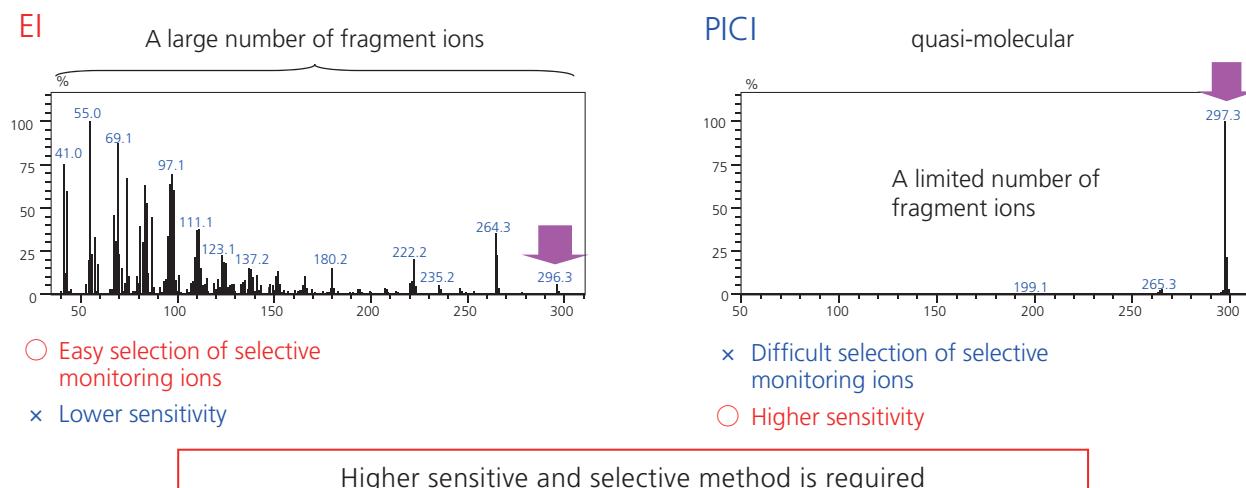
## Determination of Fatty Acids in Foods Using Gas Chromatography with Positive-ion Chemical Ionization Tandem Mass Spectrometry

# Introduction

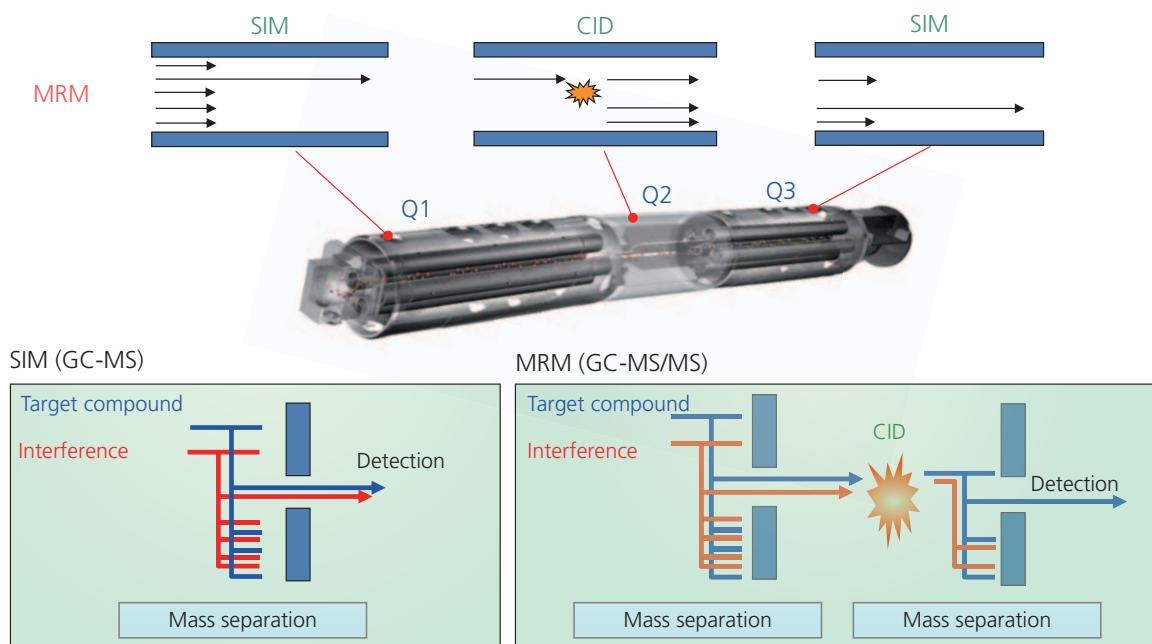
A GC-MS with electron ionization mode (EI) is widely used for qualitative and quantitative determination of multi-component fatty acids in foods. However, selection of an appropriate monitoring ion is not always easy in EI mode because of the low abundance of fragment ions and the presence of co-eluting interferences. The more abundant quasi-molecular ion of positive ion chemical ionization

mode (PICI) can be used instead, but it is also subject to overlapping co-elutants, in which case neither EI nor PICI mode produces ideal results. The tandem mass spectrometry (MS/MS) is one of choices available to solve this problem. In this study, the following four methods, GC-EI-MS, GC-PICI-MS, GC-EI-MS/MS, and GC-PICI-MS/MS, are evaluated.

### EI and PICI      Oleic acid methyl ester (C<sub>18</sub>:1n-9) (M.W. 296.3)



### New approach of GC-MS/MS



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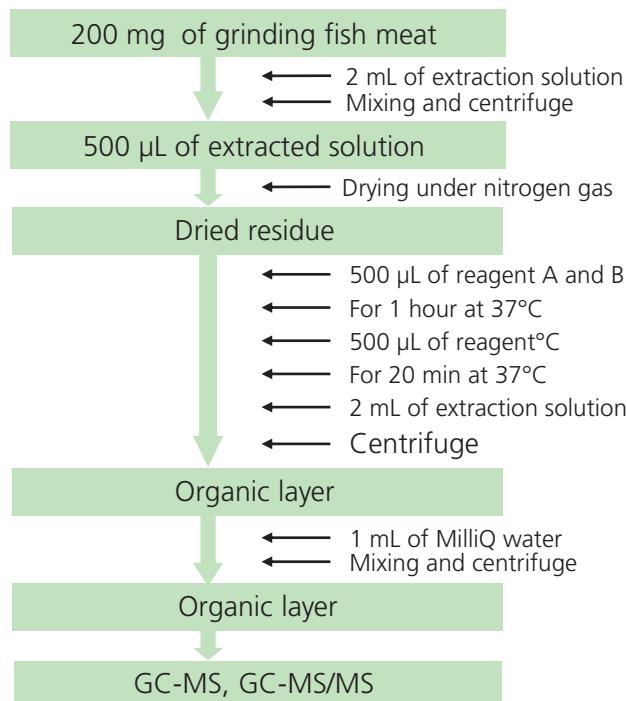
# Methods and Materials

## Sample Preparation

Brevoort



A fatty acid methylation kit and a fatty acid methyl ester purification kit.  
(Nacalai Tesque, Inc.)



## Analytical Conditions

GC-MS: GCMS-TQ8030

Column: SP-2560 (L=100 m, 0.20 mm I.D., df=0.25  $\mu$ m)

[GC]

Injection temp. :250°C  
Column oven temp. :40°C (2 min)→(4°C/min)→240°C(15 min)  
Injection mode :Split (1:10)  
Carrier gas flow control :Constant linear velocity (20.0 cm/sec)

[MS]

Interface temp. :250°C  
Ion source temp. :200°C  
Measuring mode :SIM, MRM  
Collision gas :Ar (200 kPa)  
Collision energy (CE) :An optimal value for each compound  
Ionization mode :  
**EI** :70eV  
**PICl** :Iso-butane (70 kPa)



Monitoring  $m/z$

EI-SIM :the most intense ion above  $m/z$  100 (below  $m/z$  100 for C4:0 and C6:0)

EI-MRM

Precursor ions :the same as the ions used in EI-SIM mode.

Product ions :the most intense ion above  $m/z$  100.

PICl-SIM : $M+H^+$

PICl-MRM

Precursor ions :the same as the ions used in PICl-SIM mode.

Product ions :the most intense ion above  $m/z$  100.

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## Results

### 3-1. Limit of Quantitation (LOQs)

LOQs=  $s \times t (n-1, 0.01)$  where n=8, s: standard deviation, t (n-1, 0.01): Student's t value

Black: Saturated fatty acids

Red: Unsaturated fatty acids

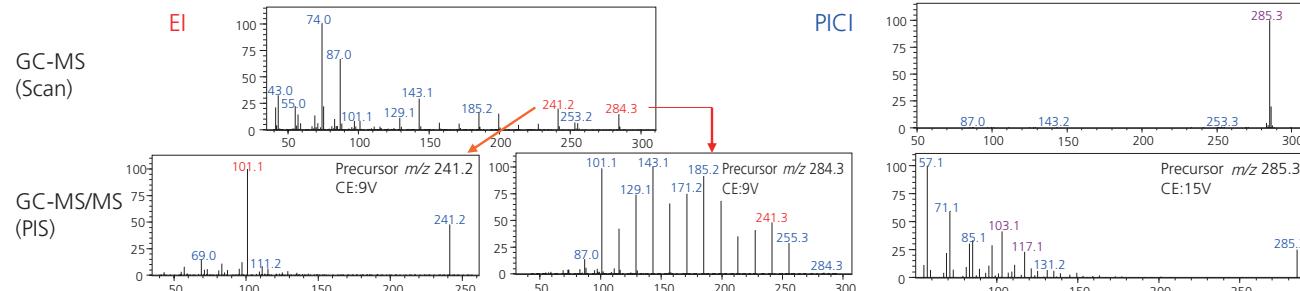
more than two times more sensitive

		CI MRM LOQ (pg)	CI SIM LOQ (pg)	EI MRM LOQ (pg)	EI SIM LOQ (pg)		CI MRM LOQ (pg)	CI SIM LOQ (pg)	EI MRM LOQ (pg)	EI SIM LOQ (pg)
1	Methyl butanoate;4:0	1.9	8.3	2.8	26.0					
2	Methyl caproate;6:0	3.6	5.9	3.5	39.2					
3	Methyl caprylate;8:0	4.8	4.8	10.5	53.9					
4	Methyl caprate;10:0	8.8	7.2	32.9	36.8					
5	Methyl undecanoate;11:0	20.6	30.1	22.0	44.0					
6	Methyl laurate;12:0	10.9	6.6	56.5	74.4					
7	Methyl tridecanoate;13:0	13.2	5.2	48.7	42.2					
8	Methyl myristate;14:0	10.1	5.5	5.6	69.7					
9	Methyl myristoleate;(Z)14:1n-5	2.7	4.7	178.3	134.0					
10	Methyl pentadecanoate;15:0	36.6	4.8	29.3	32.1					
11	Methyl cis-10-pentadecenoate;(Z)15:1n-5	4.6	4.0	225.3	33.7					
12	Methyl palmitate;16:0	15.2	7.3	12.6	74.5					
13	Methyl palmitoleate;(Z)16:1n-7	16.2	19.0	36.0	249.5					
14	Methyl margarate;17:0	29.2	20.0	5.6	22.0					
15	Methyl cis-10-heptadecenoate;(Z)17:1n-7	14.5	22.1	215.7	245.9					
16	Methyl stearate;18:0	35.3	8.9	10.2	11.6					
17	Methyl elaidate;(E)18:1n-9	19.8	5.5	180.7	173.9					
18	Methyl oleate;(Z)18:1n-9	9.8	6.4	353.2	58.1					
19	Methyl linolelaidate;(E)18:2n-6			23.2	28.8	253.9	52.0			
20	Methyl linoleate;(Z)18:2n-6			16.5	23.2	297.9	160.7			
21	Methyl arachisate;20:0			58.5	11.5	9.1	11.6			
22	Methyl gamma-linolenate;(Z)18:3n-6			81.2	17.6	167.8	349.8			
23	Methyl cis-11-icosenoate;(Z)20:1n-9			36.0	22.0	45.1	145.1			
24	Methyl linolenate;(Z)18:3n-3			135.7	23.9	414.6	213.1			
25	Methyl heneicosanoate;21:0			108.5	25.5	33.7	41.0			
26	Methyl cis-11,14-lcosadienoate;(Z)20:2n-6			36.5	13.4	282.1	238.4			
27	Methyl behenate;22:0			279.0	23.3	29.6	7.0			
28	Methyl eicoso-8,11,14-trienoate;20:3n-6			220.3	31.7	405.2	140.5			
29	Methyl erucate;22:1n-9			96.1	31.2	387.8	143.3			
30	Methyl cis-11,14,17-lcosatrienoate;(Z)20:3n-3			284.5	24.3	-	446.0			
31	Methyl tricosanoate;23:0			357.2	19.3	54.2	24.8			
32	Methyl arachidonate;(Z)20:4n-6			151.7	45.5	181.2	292.2			
33	Methyl cis-13,16-Docosadienoate;(Z)22:2n-6			128.1	315.7	335.3	283.2			
34	Methyl lignocerate;24:0			503.8	41.8	52.6	10.3			
35	Methyl cis-5,8,11,14,17-Eicosapentaenoate;(Z)20:5n-3			184.9	54.9	286.5	437.4			
36	Methyl nervonate;(Z)24:1n-9			99.4	56.5	445.2	230.7			
37	Methyl cis-4,10,13,16,19-Docosahexaenoate;(Z)22:6n-3			255.7	304.2	161.5	281.7			

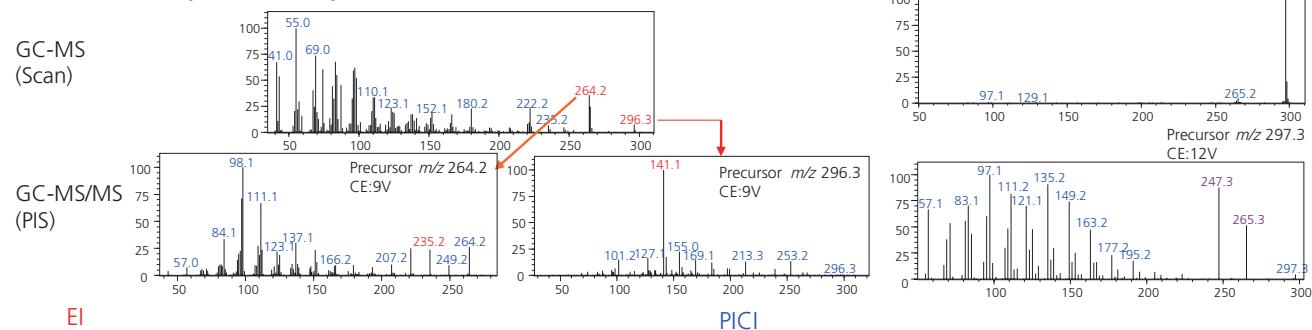
Measuring mode	Total number	Saturated	Unsaturated
EI SI	7	6	1
EI MRM	10	9	1
PICI SIM	32	13	19
PICI MRM	18	7	11

PICI was the most sensitive mode, especially in the analysis of unsaturated fatty acids

#### Saturated fatty acid (methyl margarate;C17:0 M.W. 284.3)



#### Unsaturated fatty acid (Methyl oleate; (Z) C18:1n-9 M.W. 296.3)



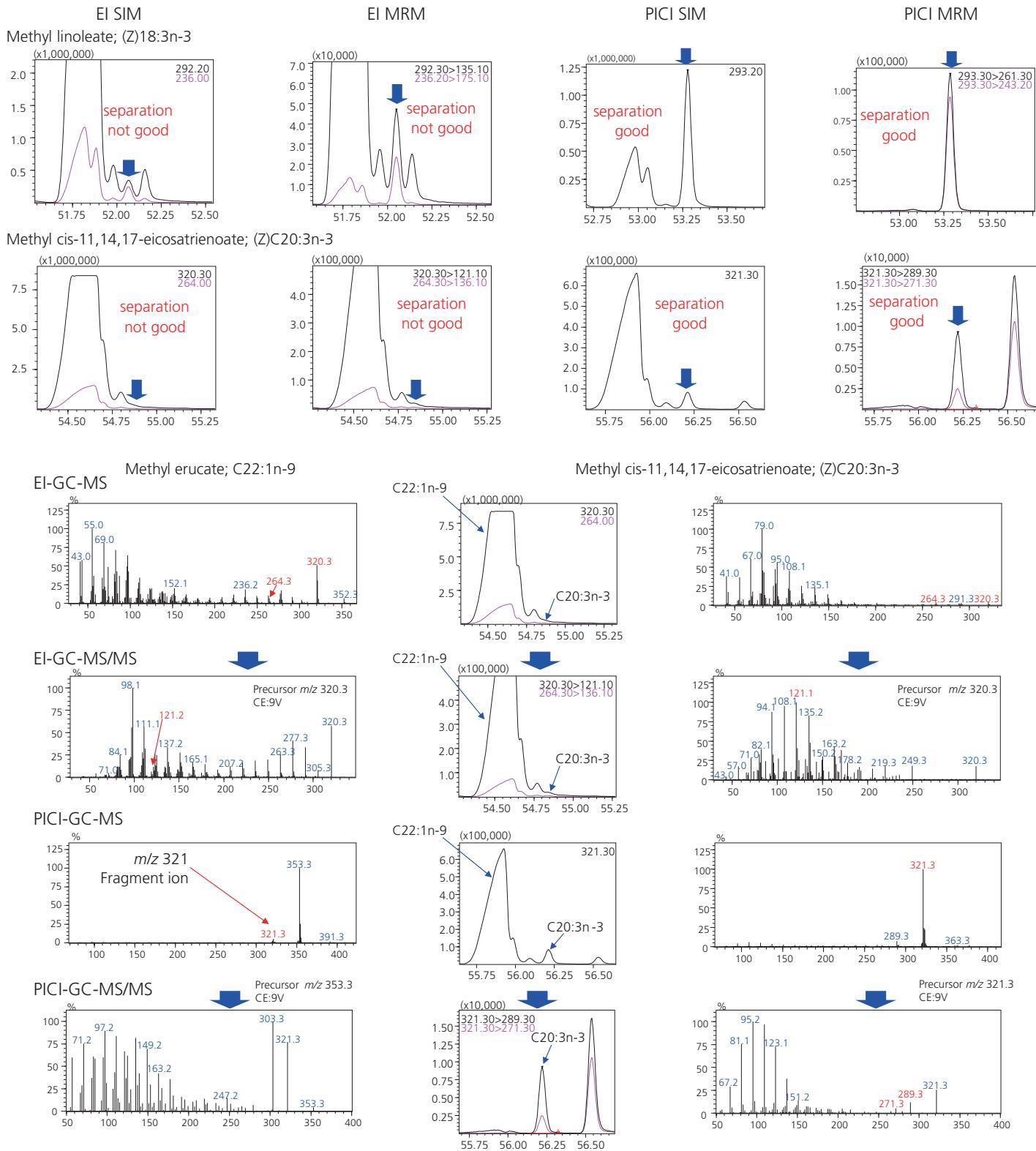
A variety of fragment ions Intensity of each ion is low (especially unsaturated fatty acids)

Unsaturated fatty acids produced intense product ions

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### 3-2. Mass Separation in Food Sample

Brevoort



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### Conclusions

- The PICI-SIM and PICI-MRM modes showed better LOQs compared to EI-SIM and EI-MRM in the analysis of 37 fatty acids, especially the unsaturated fatty acids.
- Since EI mass spectrum patterns of the fatty acids were similar, it was not easy to choose characteristic ion to separate from other fatty acids, especially in real samples.
- Although PICI-SIM was more selective compared to EI-SIM and EI-MRM, only several ions could be chosen for monitoring.
- Since PICI-MRM was more selective than PICI-SIM due to MS/MS, the fatty acids were selectively detected, which improved the reliability of identification.

First Edition: June, 2013



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