

Determination of 20 Phthalic Acid Esters in Alcoholic Drinks by Ultra High Performance Liquid Chromatography/Tandem Mass Spectrometry

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# 1. Introduction

Phthalic acid esters (PAEs) are a group of commercial chemicals widely used to make plastics more malleable and help lotions penetrate skin. A number of phthalate esters are known to cause birth defects or reproductive harm. PAEs can migrate from plastic materials to the environment. They are often found in water, soil, air, food products and the human body. As well known, alcoholic drinks have always been popular around the world. In alcoholic drinks production, plastic containers are typically used in the storage and transportation process, which could make some phthalate esters leak easily from PVC tubes or vessels as well as plastic caps. The aim of this study is to determine the level of phthalate esters migration in alcoholic drinks by fast liquid chromatography-electrospray tandem mass spectrometry. This method is simple and rapid with acceptable sensitivity to meet the requirements for the analysis of PAEs in alcoholic drinks.

# 2. Method and Materials

### Sample Preparation

Accurately weigh 5.0 g of alcoholic drinks into a glass tube. After centrifugation for 20 min at 6000 rpm, the supernatant was analyzed by LC/MS/MS.

#### UHPLC/MS/MS Analysis UHPLC

The analyses were performed on a Shimadzu Nexera UHPLC instrument (Kyoto, Japan) equipped with LC-30AD pumps, a CTO-30A column oven, a DGU-30A5 degasser, and an SIL-30AC autosampler. The separation was carried out on a Shim-pack XR-ODSIII column (150 mmL. × 2.0 mmi.d., 2.2  $\mu$ m) with the column temperature at 45°C. The delay column (Inertsil ODS-4 50 mmL. × 3.0 mmi.d., 2.0  $\mu$ m) was used to reduce the PAEs interference which comes from the mobile phases. The mobile phase consisted of (A) 5 mmol/L ammonium acetate-water and (B) methanol using gradient elution of 45%-90%B at 0-6.5 min, 90%-100%B at 6.5-7.0 min, 100%B at 7.0-9.9 min, and 100%-45%B at 9.9-10 min. The flow rate was 0.4 mL/min. The injection volume was 10  $\mu$ L.

### Mass spectrometry

A triple quadrupole mass spectrometer (Shimadzu LCMS-8040, Kyoto, Japan) was connected to the Shimadzu UHPLC instrument via an ESI interface. The mass spectra were acquired in positive ion mode. The DL temperature was set at 250°C, with heat block temperature at 450°C, nebulizing gas at 3 L/min and drying gas at 15 L/min. The dwell time was 15 ms and the pause time was 3 ms. The MRM parameters were shown in Table 1.

### Standard PAEs

Dimethyl phthalate (DMP), diethyl phthalate (DEP), bis(2-ethoxyethyl) phthalate (DEEP), bis(2-methoxyethyl) phthalate (DMEP), diallyl (o-)phthalate (DAP), dipropyl phthalate (DPRP), diisopropyl phthalate (DiPRP), diphenyl phthalate (DIPP), benzyl butyl phthalate (BBP), diisobutyl phthalate (DIBP), bis(2-butoxyethyl) phthalate (DBEP), dibutyl phthalate (DBP), dipentyl phthalate (DPP), diisoamyl phthalate (DIAP), bis(4-methyl-2-pentyl) phthalate (BMPP), bis(2-ethylhexyl) phthalate (DEHP), di-n-octyl phthalate (DNOP), dicyclohexyl phthalate (DCHP), di-n-hexyl phthalate (DHXP), and diheptyl phthalate (DHP). All of the standard PAEs were purchased from ANPEL Scientific Instrument Co., Ltd.

## 3. Results and Discussion

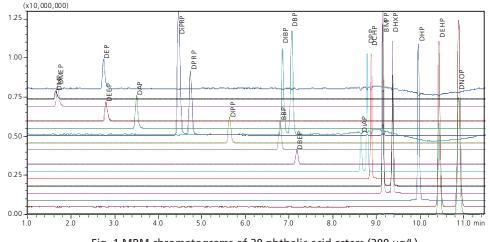


Fig. 1 MRM chromatograms of 20 phthalic acid esters (200  $\mu\text{g/L})$ 

Compound	Precursor ion ( <i>m/z</i> )	Production ( <i>m/z</i> )	Q1 pre bias (V)	CE (V)	Q3 pre bias (V)
DMP 194.7		163.0*	-23	-11	-17
	194.7	77.1	-23	-31	-14
DEMP	283.1	207.0*	-21	-6	-22
DEIVIP	205.1	149	-21	-28	-29
DEP	223.1	149.0*	-30	-20	-30
DLI	223.1	177.1	-30	-10	-18
DEEP	311.1	73.1*	-23	-13	-30
DELI	511.1	221.1	-23	-6	-24
DAP	247.1	189.1*	-28	-11	-20
DAI	247.1	149.1	-28	-16	-27
DPRP	251.1	149.0*	-30	-20	-30
DERI	231.1	191.1	-30	-9	-20
Diprp	251.1	149.0*	-30	-20	-30
DILIKI	231.1	191.1	-30	-9	-20
DIPP	319.1	225.0*	-23	-9	-24
DILL	515.1	77.1	-23	-36	-30
BBP	313.1	91.1*	-30	-20	-17
וסס	515.1	149	-30	-12	-16
DIBP	279.1	149.0*	-30	-20	-30
DIDI		205.1	-30	-8	-22
DBP	279.1	149.0*	-30	-20	-30
DDI	275.1	205.1	-30	-8	-22
DBEP	367.1	101.1*	-27	-12	-19
DDEI	507.1	249.1	-27	-7	-27
DPP	307.1	149.1*	-30	-20	-30
DIT		219.1	-30	-7	-24
DiAP	307.1	149.1*	-30	-20	-30
0111		219.1	-30	-7	-24
DCHP	331.1	149.1*	-30	-25	-28
Dem		167.1	-30	-13	-18
B MP P	335.1	149.1*	-30	-25	-27
DIVIPP		167	-30	-11	-18
DHXP	335.2	149.1*	-30	-20	-30
	333.2	233.2	-30	-8	-25
DHP	363.2	149.1*	-30	-20	-30
2111		247.2	-30	-8	-27
DEHP	391.3	149.1*	-30	-25	-30
DERF		167.1	-30	-13	-17
DNOP	391.4	149.0*	-30	-20	-30
DINOI		261.1	-30	-8	-29

Table 1 MRM parameters of 20 phthalic acid esters

\* for quantitation

20 phthalic acid esters were separated in 11 min. The MRM chromatograms in positive mode were shown in Fig. 1. A linear relationship was found between peak area and different concentrations of 20 phthalate esters within 5,

10, 20, 50 and 200  $\mu$ g/L. Correlation coefficients (r) more than 0.999, the limits of detection (LODs) and the limits of quantitation (LOQs) were obtained as shown in Table 2.

Compound	Calibration curve	r	LOD (µg/L)	LOQ (µg/L)
DMP	Y = 12673.3X + 2300.05	0.9998	0.62	1.85
D ME P	Y = 26744.3X - 12057.1	0.9999	0.23	0.70
DEP	Y = 41199.3X + 60434.8	0.9998	1.67	5.00
DEEP	Y = 24761.6X - 25617.3	0.9999	0.18	0.56
DAP	Y = 43997.2X - 53600.0	0.9999	0.12	0.36
Diprp	Y = 166476X - 70183.0	0.9999	0.05	0.15
DPRP	Y = 82151.0X - 14189.4	0.9999	0.09	0.26
DIPP	Y = 28688.5X - 89879.0	0.9990	0.16	0.47
BBP	Y = 36060.8X + 13664.7	0.9999	0.10	0.29
DIBP	Y = 105063X + 1597740	0.9999	1.63	4.94
DBP	Y = 131073X + 373016	0.9999	0.83	2.50
DBEP	Y = 16453.9X - 2429.68	0.9999	0.13	0.38
Diap	Y = 39687.3X + 274520	0.9989	0.03	0.10
DPP	Y = 115069X + 108679	0.9999	0.02	0.06
DCHP	Y = 114309X + 233683	0.9999	0.02	0.05
B MP P	Y = 143127X + 451840	0.9996	0.02	0.05
DHXP	Y = 129006X + 132748	0.9999	0.02	0.06
DHP	Y = 128802X - 125656	0.9999	0.03	0.10
DEHP	Y = 164622X - 14942.3	0.9998	0.47	1.42
DNOP	Y = 120832X - 232093	0.9998	0.46	1.40

Table 2 The calibration curve, LOD and LOQ of 20 PAEs

Table 3 Repeatabilit	y of peak area	and RT at different	concentrations (n=6)
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Compound	%RSD (20 µg/L)		%RSD (50 µg/L)		%RSD (100µg/L)	
	Area	R.T.	Area	R.T.	Area	R.T.
DMP	3.23	0.08	3.14	0.21	1.76	0.24
D ME P	0.73	0.08	0.71	0.19	0.74	0.20
DEP	4.79	0.17	3.90	0.18	2.76	0.24
DEEP	1.04	0.18	1.18	0.19	1.22	0.26
DAP	0.85	0.18	1.25	0.18	0.81	0.26
DiP R P	0.93	0.18	0.49	0.14	1.17	0.24
DPRP	0.61	0.17	0.87	0.13	0.91	0.22
DIPP	0.84	0.16	1.00	0.10	1.27	0.19
BBP	1.29	0.13	0.36	0.07	1.19	0.15
DIBP	3.55	0.12	3.34	0.07	1.82	0.15
DBP	4.79	0.11	1.50	0.07	1.40	0.14
DBEP	1.33	0.12	0.66	0.06	0.41	0.14
DiAP	1.56	0.05	1.44	0.02	1.10	0.05
DPP	1.30	0.09	0.26	0.09	1.74	0.07
DCHP	1.51	0.04	0.75	0.02	0.56	0.05
B MP P	1.35	0.04	0.83	0.02	1.23	0.03
DHXP	1.24	0.04	0.66	0.02	1.57	0.02
DHP	1.86	0.04	0.99	0.03	1.45	0.02
DEHP	0.59	0.06	0.42	0.04	1.65	0.04
DNOP	1.76	0.08	0.85	0.06	0.98	0.06

In this study, the repeatability of 20 phthalic acid esters at different concentrations (20, 50 and 100  $\mu$ g/L) was investigated. The %RSDs of retention time were better than 0.26% and %RSDs of peak area were less than 4.79%, as shown in Table 3.

The mixed standard sample was spiked into the blank alcoholic drink at the levels of 50  $\mu$ g/kg and 100  $\mu$ g/kg to evaluate the recovery of this method developed in this study. A good recovery of 78% to 127% was obtained for

each of the compound. The results are shown in Table 4. A 50-percent alcohol distilled liquor made by Hunan-based liquor producer in China contained a maximum of 1.04 mg of DBP per kg. Three samples, including Chinese liquor, wine and whisky from the local market were chosen for analysis. The results showed that 154.1 µg/kg DBP and 18.7 µg/kg DIBP were detected in one of Chinese liquor, and no detection in other samples.

		50 µ	g/kg	100 µg/kg	
No. Compo	Compound	Measured value (µg/kg)	Recovery (%)	Measured value (µg/kg)	Recovery (%)
1	DMP	42	91	92	100
2	D ME P	48	104	101	110
3	DEP	45	97	101	110
4	DEEP	49	107	102	111
5	DAP	49	107	102	111
6	Diprp	48	104	101	110
7	DPRP	48	105	105	114
8	DIPP	54	118	117	127
9	BBP	47	102	98	106
10	DIBP	45	99	98	106
11	DBP	48	104	101	110
12	DBEP	48	104	102	111
13	DiAP	36	78	77	84
14	DPP	46	99	91	99
15	DCHP	43	92	85	93
16	B MP P	45	99	91	99
17	DHXP	46	101	91	99
18	DHP	46	99	89	97
19	DEHP	46	100	97	105
20	DNOP	45	99	96	105

#### Table 4 Recovery of 20 PAEs in an alcoholic drink

## 4. Conclusions

A UHPLC/MS/MS method has been developed for determination of 20 PAEs in alcoholic drinks. All of them were separated in 11 minutes, and analyzed in positive mode. The calibration curves of 20 PAEs were constructed over a concentration range of 50-200 µg/L with correlation coefficients (r) more than 0.999. Good repeatability on both retention time and peak area was obtained. The limits of detection (LODs) and the limits of quantitation (LOQs) for 20 PAEs were better than 2  $\mu$ g/L, 5  $\mu$ g/L, respectively. The method was established for fast, reliable quantitative determination of 20 PAEs in alcoholic drinks.

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