

# Estimation of β-Sitosterol in Milk Fat (Ghee) Samples

Agilent 8890/5977B Single Quadrupole GC/MS System



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### Abstract

This application note demonstrates the use of the Agilent 8890 GC and the Agilent 5977B GC/MS single quadrupole mass spectrometer in the detection and quantification of  $\beta$ -sitosterol in milk fat samples to check for vegetable oil adulteration.

The method provides the highest confidence for routine analysis of milk fat samples in the food industry, whether it is used in manufacturing, processing, commercial testing, or academia. Sample preparation for this method involved saponification, followed by extraction of unsaponifiable matter by liquid-liquid extraction (LLE) and derivatization of sitosterol to its trimethylsilyl derivative.

### Introduction

Ghee (milk fat), also known as clarified butter, is commonly used in cuisine of the Indian subcontinent, traditional medicines, and religious rituals. Ghee is prepared by skimming the milk solids out of melted butter. Increasing demand for ghee in India has resulted in certain malpractices such as adulteration of ghee with vegetable oils.<sup>1</sup>

Plant sterols can be used as marker compounds for identification of vegetable oil adulteration in ghee. One of the common representatives of plant sterol is  $\beta$ -sitosterol.





This application note demonstrates detection of vegetable oil adulteration in ghee. The principle of detection of adulteration is based on the presence of  $\beta$ -sitosterol as a marker in the unsaponifiable matter of pure ghee and adulterated ghee samples.<sup>3</sup>

The method used for detection of the presence of any vegetable oil is based on gas chromatography coupled to mass spectrometry.

## **Experimental conditions**

#### **Chemicals required**

20% ascorbic acid (AR grade) (2 g of ascorbic acid dissolved in 10 mL water, potassium hydroxide (10 M, 56 g of KOH dissolved in 100 mL of water)(AR grade), water (Millipore, Milli-Q) *n*-hexane (HPLC grade)  $\beta$ -sitosterol reference standard, BSTFA reagent (N,O-*bis*(trimethylsilyl) trifluro), pyridine (GC grade).<sup>2</sup>

All working solutions of  $\beta$ -sitosterol were prepared in *n*-hexane. 100 µL of standard was placed into a vial, before 50 µL of pyridine was added. The vials were kept at 80 °C for 40 minutes. The final extract was taken in vial inserts and injected into the GC/MS.

### Instrument

Agilent 8890 GC system with S/SL inlet and Agilent 5977B GC/MS single quadrupole

#### Ghee sample preparation



**Figure 2.** Liquid-liquid extraction, sample preparation.

#### Table 1. GC method.

| GC Conditions        |   |  |  |  |  |  |  |
|----------------------|---|--|--|--|--|--|--|
| Column               | Agilent J&W HP-5ms Ultra Inert, 30 m × 0.25 mm, 0.25 μm (p/n 19091s-433UI)            |  |  |  |  |  |  |
| Inlet                | Agilent split/splitless inlet<br>5190-2293, splitless liner<br>Injection volume: 1 μL |  |  |  |  |  |  |
| Split Mode And Ratio | Split ratio 5:1   |  |  |  |  |  |  |
| Inlet Temperature    | 280 °C  |  |  |  |  |  |  |
| Oven                 | 80 °C for 1 minute, at 15 °C/min to 290 °C, hold 30 minutes                           |  |  |  |  |  |  |
| Carrier Gas          | 99.9995% helium at 1.0 mL/ min, constant flow mode                                    |  |  |  |  |  |  |

Table 2. MS method.

| MSD Conditions            |  |  |  |  |  |
|---------------------------|--|--|--|--|--|
| Quadrupole Temperature    | 150 °C                                   |  |  |  |  |
| Ion Source Temperature    | EI 230 °C                                |  |  |  |  |
| Transfer Line Temperature | 290 °C                                   |  |  |  |  |
| Acquisition Type          | SIM mode; ions – 396, 486, 357, 381, 129 |  |  |  |  |
| EMV Mode                  | Delta EMV: 0                             |  |  |  |  |
| Dwell Time for Each Mass  | 50                                       |  |  |  |  |

### **Results and discussion**

With the method described, instrument LOQ was determined at 200 ppb for the reference standard. Figure 3 highlights the quantifier and qualifier EIC at LOQ level. Figure 4 describes the signal-to-noise for 200 and 500 ppb level standards. Figure 4 also showcases the specificity of the method by showing a blank injection.



Figure 3. Qualifier and quantifier peaks of sitosterol at LOQ (200 ppb).



Figure 4. Sensitivity of  $\beta$ -sitosterol at 200 ppb (LOQ), 500 ppb, and blank.

### **Calibration and linearity**

A linearity plot was generated for response (peak area) across concentration levels from 200 ppb to 5 ppm (figure 6). Calibration was performed at five levels, 200 ppb, 500 ppb, 1 ppm, 2 ppm, and 5 ppm. Linearity with R<sup>2</sup> >0.999 was observed. The calibration table with one quantifier ion and two qualifier ions is shown in Figure 7, in accordance with the regulations.



Figure 5. Overlay of various concentrations of β-sitosterol.



Figure 6. Calibration plot for β-sitosterol standards (concentration (ppm) versus response (peak area)).

| Sample         |        | b-Sitosterol |            | b-Sitostero | Qualifier (357 | Qualifier (381 |          |       |       |
|----------------|--------|--------------|------------|-------------|----------------|----------------|----------|-------|-------|
| Name           | Туре   | Level        | Exp. Conc. | RT          | Resp.          | Final Conc.    | Accuracy | Ratio | Ratio |
| STD-200ppb     | Cal    | 1            | 0.2000     | 22.410      | 853            | 0.2305         | 115.2    | 144.7 | 50.7  |
| STD-500ppb     | Cal    | 2            | 0.5000     | 22.410      | 1852           | 0.4681         | 93.6     | 135.2 | 51.1  |
| STD-1 ppm      | Cal    | 3            | 1.0000     | 22.410      | 4323           | 1.0554         | 105.5    | 131.4 | 51.2  |
| STD-2 ppm      | Cal    | 4            | 2.0000     | 22.406      | 7983           | 1.9252         | 96.3     | 132.5 | 50.7  |
| STD-5 ppm      | Cal    | 5            | 5.0000     | 22.406      | 21006          | 5.0208         | 100.4    | 131.7 | 51.0  |
| STD-1 ppm_rep1 | Sample |              |            | 22.402      | 4794           | 1.1672         |          | 127.1 | 53.2  |
| STD-1 ppm_rep2 | Sample |              |            | 22.401      | 4894           | 1.1909         |          | 124.6 | 53.6  |
| STD-1 ppm_rep3 | Sample |              |            | 22.397      | 4971           | 1.2094         |          | 128.8 | 52.4  |
| STD-1 ppm_rep4 | Sample |              |            | 22.397      | 5082           | 1.2357         |          | 126.8 | 50.4  |
| STD-1 ppm_rep5 | Sample |              |            | 22.393      | 5006           | 1.2178         |          | 121.0 | 51.3  |
| STD-1 ppm_rep6 | Sample |              |            | 22.393      | 5132           | 1.2477         |          | 134.3 | 53.1  |

Figure 7. Calibration table for  $\beta$ -sitosterol from 200 ppb to 5 ppm and repeatability at 1 ppm.

#### Repeatability

A repeatable response was obtained by injecting 1 ppm  $\beta$ -sitosterol standard. As shown in Table 3, the % RSD data of  $\beta$ -sitosterol are calculated from peak areas of six replicate injections at 1 ppm concentration.

#### Quantitation in ghee samples

The suggested method was extended to a ghee sample. The ghee sample was purchased from a market for the analysis and recovery study.

As shown in Figure 8, 2.24 ppm  $\beta$ -sitosterol was found in the ghee sample on which the study was performed. A chromatogram of ghee sample is shown in Figure 9.

### **Recovery study**

Spiking was done at 5 and 25 ppm respectively. Table 4 shows recovery in the spiked samples after blank subtraction. Table 3. Percentage RSD (CV) for peak area at 1 ppm  $\beta$ -sitosterol.

| Area Inj-1 | Area Inj-1 Area Inj-2 |      | Area Inj-3 Area Inj-4 |      | Area Inj-6 | %RSD |
|------------|-----------------------|------|-----------------------|------|------------|------|
| 4794 4894  |                       | 4971 | 5082                  | 5006 | 5132       | 2.48 |

| Sample        |                     |        | b-Sitosterol Results |       |             | Qualifier (357.0) | Qualifier (381.0) |
|---------------|---------------------|--------|----------------------|-------|-------------|-------------------|-------------------|
| Data File Typ |                     | Туре   | RT                   | Resp. | Final Conc. | Ratio             | Ratio             |
|               | Ghee spike 5 ppm.D  | Sample | 22.381               | 1341  | 8.5195      | 106.1             | 42.8              |
|               | Ghee spike 25 ppm.D | Sample | 22.397               | 3815  | 23.1042     | 94.4              | 41.7              |
| ►             | Ghee Sample.D       | Sample | 22.381               | 276   | 2.2418      | 107.6             |                   |

Figure 8. Quantitation in ghee samples.

Table 4. Recovery in ghee samples.

| Spiked Amount<br>(ppm) | Observed Amount<br>(ppm) | Final Amount<br>(ppm) | Recovery<br>(%) |  |
|------------------------|--------------------------|-----------------------|-----------------|--|
| 5                      | 8.52                     | 6.28                  | 125.6           |  |
| 25                     | 23.1                     | 20.86                 | 83.4            |  |



Figure 9. Chromatogram for ghee sample spiked at 25 ppm level for sitosterol.

### Conclusion

An accurate and rugged method was developed for analysis of  $\beta$ -sitosterol in ghee samples for the identification of adulteration by vegetable oils. The saponification/LLE/derivatization based sample preparation method used easy and less time-consuming steps. The lowest calibration level was 200 ppb for the standard. Repeatable results were found for six replicates of standard. Satisfactory recovery was found at 5 and 25 ppm spiked concentration in ghee samples.

### References

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