# A systematic approach to LC method transfer from 4.6 to 2.1mm ID columns

ofile remains unchanged in the new method.

 $t_{eq} = t_{ef} \times (V_{eq}/V_{ef}) \times (F_{e}/F_{e})$ 

aradionte

Results

demonstrated on Figure 2.

150x2.1mm

shown in Figure 3b

Step 4. Optimise system set-up

4.1 System dispersion

4.2 Pump dwell volume

extra-column dispersion when working with these columns.

it takes for the gradient to reach the head of the column.

(a)

Same flow rate (1 mL/min)

approximately 5 times higher than in the 4.6 mm ID column, as demonstrated in Figure 1b.

ul, dwell volume adds approximately 3 minutes to the elution time on a 2 1mm ID column in relation to the elution.

GURE 1. Isocratic method run on 150x4.6mm and 150x2.1mm columns keeping flow r Id injection volume constant (h).

150x4 6mm

## L. Pereira, D. Milton, H. Ritchie

#### Abstract

Purpose: To demonstrate a systematic approach to method transfer between 4.6 and 2.1 mm ID columns, with consideration to system set-up and the effect it has on method performance Methods: 4 step method to adjust flow rate injection volume, gradient profile and system set-up Results: Results demonstrate savings in solvent consumption, gain in sensitivity and the impact of system volumetric effects on gradient delivery to the column.

#### Introduction

There are several reasons for reducing the column internal diameter from 4.6 to 2.1 mm. The most common in recent times has been to reduce solvent costs, as mobile phase flow rate through the column is reduced by a factor of 5. Another reason is to improve analysis sensitivity as smaller internal diameter columns provide detection signal enrichment relative to normal bore columns, by eluting solutes in more concentrated chromatographic bands. A third reason is when methods are transferred from UV to MS detection, as large volumes of mobile phase are more difficult to vaporise in the LC/MS interface.

To maintain a consistent assay profile between the original and transferred method it is necessary to scale down flor rate, injection volume and gradient profile in methods that use gradient elution. To obtain the best data it is critical that the LC system is ontimized to operate under these conditions. All system components for the assay should be considered. System volume that includes connecting tubing ID and length, injection volume, flow cell volume (in UV) must be minimized, and when running gradients pump dwell volume needs to be minimized.

The work presented in this poster demonstrates a systematic approach to method transfer between 4.6 and 2.1 mm ID columns, with consideration to system set-up and the effect it has on method performance

### Materials & Methods

Instrumentation: Thermo Scientific Surveyor HPLC system (fitted with 5 cm/ 10 uL flow cell). Thermo Scientific Accela UHPLC system (fitted with 1 cm / 2 µL flow cell).

Columns: Thermo Scientific Hypersil GOLD 5 μm, 150 x 4.6 mm, Hypersil GOLD™ 5 μm, 150 x 2.1 mm, Hypersil GOLD 5 μm, 100 x 2 1 mm

Mobile phase: A - H<sub>2</sub>O + 0.1% formic acid; B - ACN + 0.1% formic acid Isocratic method - A : B (80:20) Gradient: 10 to 50% B in 10 minutes: 100% by 15 minutes Femperature: 30 °C; Detection: UV at 270 nm Flow rate : 1 ml /min or 0 210 ml /min njection volume: 20 or 4 µL Test solution: 1. Uracil: 2. p-Coumaric acid: 3. m-Coumaric acid: 4. o-Coumaric acid

Method transfer

To transfer the method geometrically to the narrower bore column and therefore ensure equivalent chromatography it is necessary to scale down the flow rate, injection volume and gradient profile:

#### Step 1. Adjust flow rate (keep linear velocity constant between original and new method)

F1 - original flow rate; F2 - new flow rate (mL/min)  $F_2 = F_1 x (dc_2^2 / dc_1^2)$ dc1 - original column ID; dc2 - new column ID (mm) Step 2. Adjust injection volume  $V_{12} = V_{11} \times (d_{c2}^2 \times L_2 / d_{c1}^2 \times L_1)$ 

#### V<sub>11</sub> - original injection volume; V<sub>12</sub> - new injection volume (mL) det - original column ID; det - new column ID (mm) L1 - original column length; L2 - new column length (mm)

Step 3. Adjust gradient profile If only the column ID has been changed from 4.6 to 2.1mm and column length remains the same then the gradient

F1 - original flow rate (mL/min); F2 - new flow rate (mL/min)

(b)

Same injection volume (20 µL)

150x2.1mm

150x4.6mm

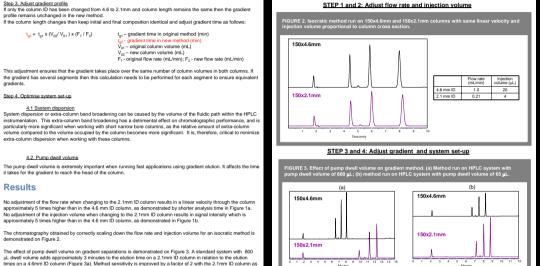
t<sub>n1</sub> - gradient time in original method (min)

gradient time in new met

Vor - original column volume (ml.) V<sub>01</sub> – new column volume (mL)

If the column length changes then keep initial and final composition identical and adjust gradient time as follows:

Thermo Fisher Scientific

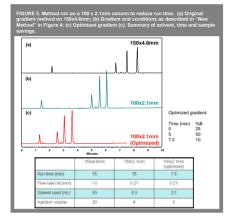


Runcorn, Cheshire, UK

Transfer method to a shorter column



The method was transferred to a shorter column, as resolution between all the sample components is high. The gradient was adjusted by using an online method transfer calculator as shown in Figure 4. The separation obtained is planet was adjusted by using an online method using the reduction as another the separation of ompared to the original gradient method on the 150 x 4.6 mm column in Figure 5 a and b. In Figure 5c the plimization of the gradient to shorten the run time is also shown.



#### Conclusions

To ensure equivalent chromatography when transferring a method to a narrower bore columns it is necessary to scale down flow rate, injection volume and gradient profile for gradient elution methods

- In gradient methods the pump dwell volume has a significant impact on the method transferability, as it adds to the time for the gradient to reach the column;
- System volume which adds to band dispersion needs to be minimized when working with narrow bore columns
- Reducing the column internal diameter from 4.6 to 2.1mm reduces solvent consumption and sample consumption by a factor of 5.
- Method sensitivity is improved with narrow bore columns. Optimization of gradient speeds up analysis.

### References

1) http://www.hpictransfer.com/

For additional information, please visit our Chromatography Resource Centre which can be found at:

All trademarks are the property of Thermo Fisher Scientific and its subsidiaries

PSGSC 0510 0210



Part of Thermo Fisher Scientific