



Quantitative comparison of perfluorinated alkyl substances in drinking water between tandem triple quadrupole MS/MS and high resolution mass spectrometry using orbitrap technology – knowns and unknowns

Ali Wolfgang Haghani (Eurofins Eaton Analytical)

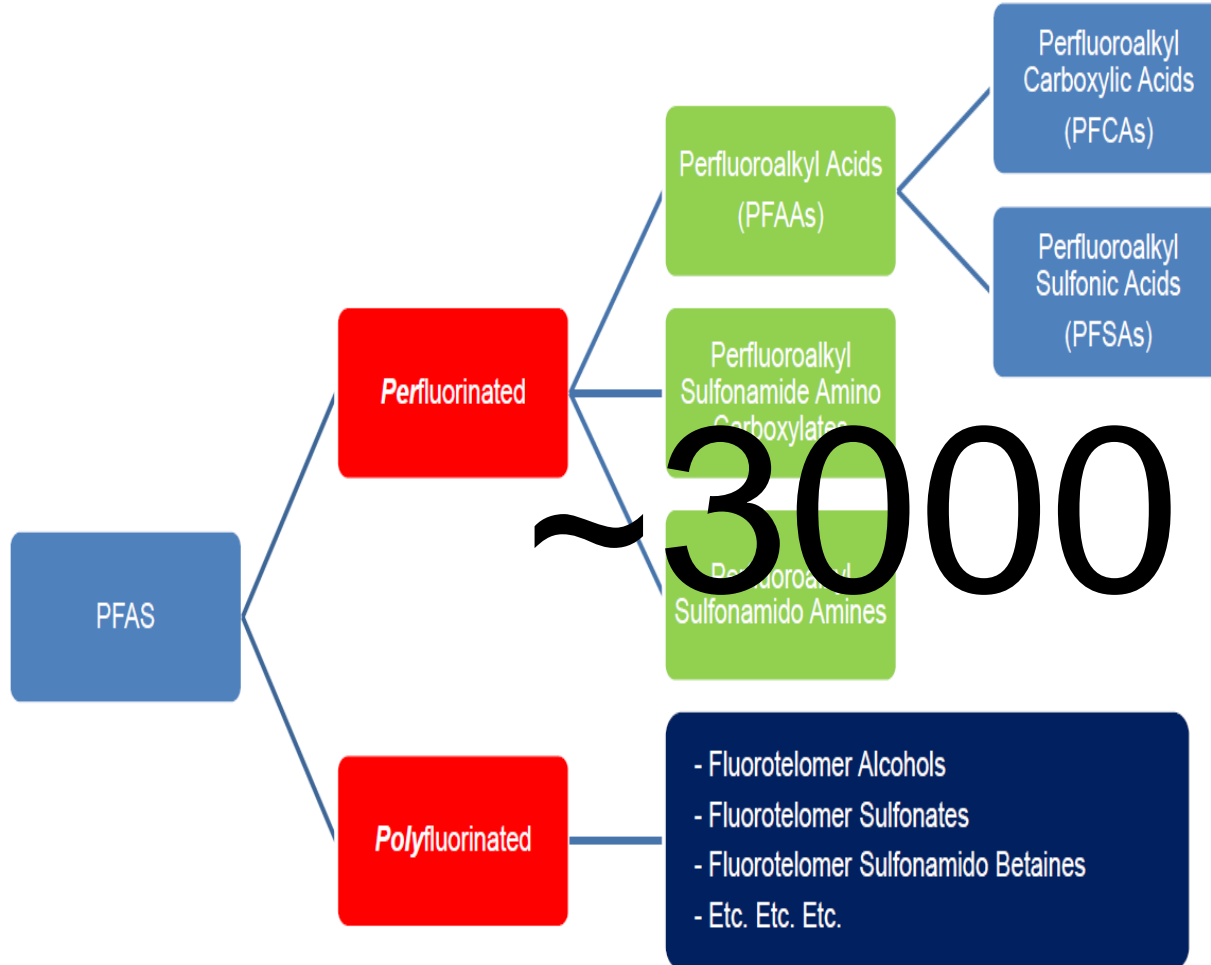
Andy Eaton, PhD, BCES (Eurofins Eaton Analytical)

Richard Jack, PhD (Thermo Fisher Scientific)

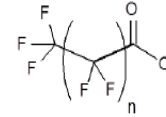
Ed George, PhD (Thermo Fisher Scientific)

Charles Yang T., PhD (Thermo Fisher Scientific)

Perfluorinated compounds as emerging contaminants

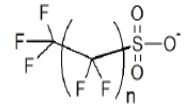


PFCAs incl. PFOA



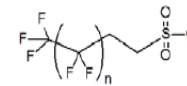
n=2, PFBA; n=3, PFPeA;
 n=4, PFHxA; n=5, PFHpA;
 n=6, PFOA; n=7, PFNA;
 n=8, PFDA; n=9, PFUnDA;
 n=10, PFDoDA;

PFSA incl. PFOS



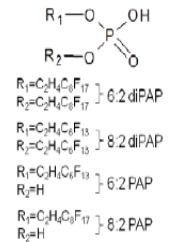
n=3, PFBS
 n=5, PFHxS
 n=7, PFOS

FTS

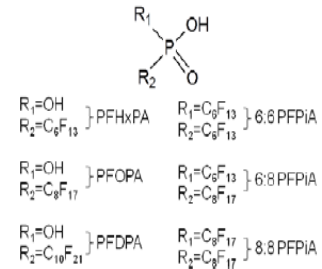


n=3, 4:2 FTS
 n=5, 6:2 FTS
 n=7, 8:2 FTS

PAP, DiPAP



PFPA/PFPiA



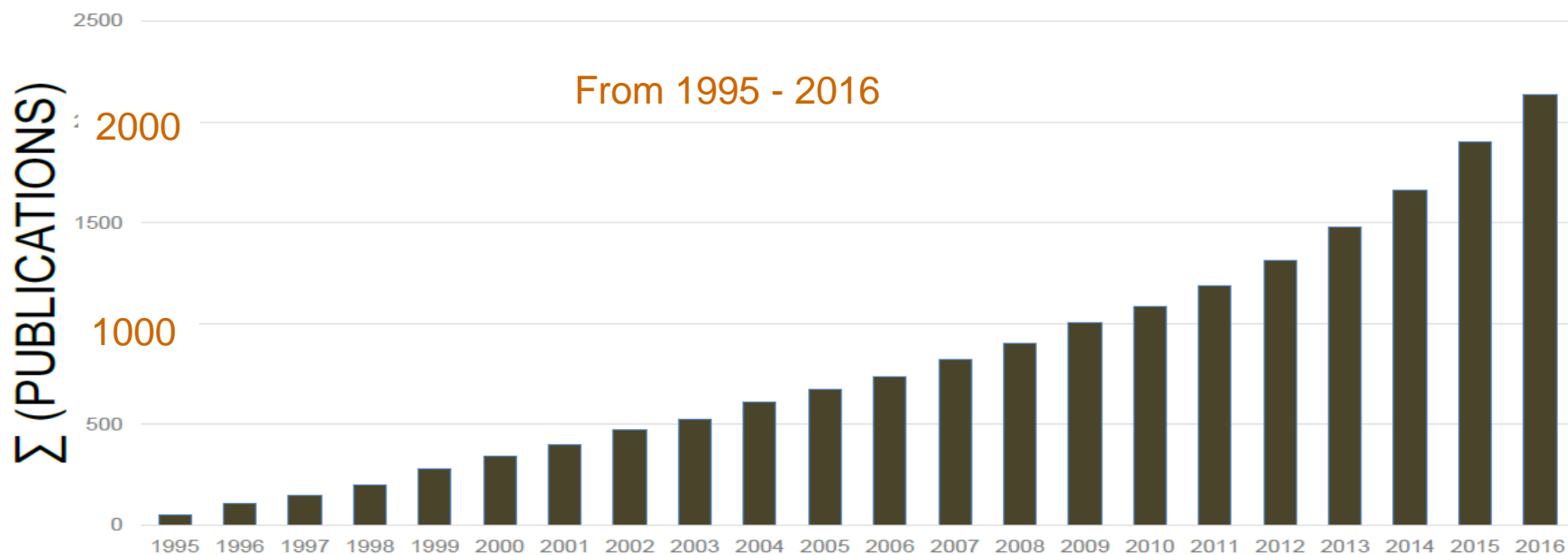
FTOH



Publications shows continued interest of PFAS.



■ Rapidly Evolving Science



Google Scholar search of "Perfluoroalkyl" in title

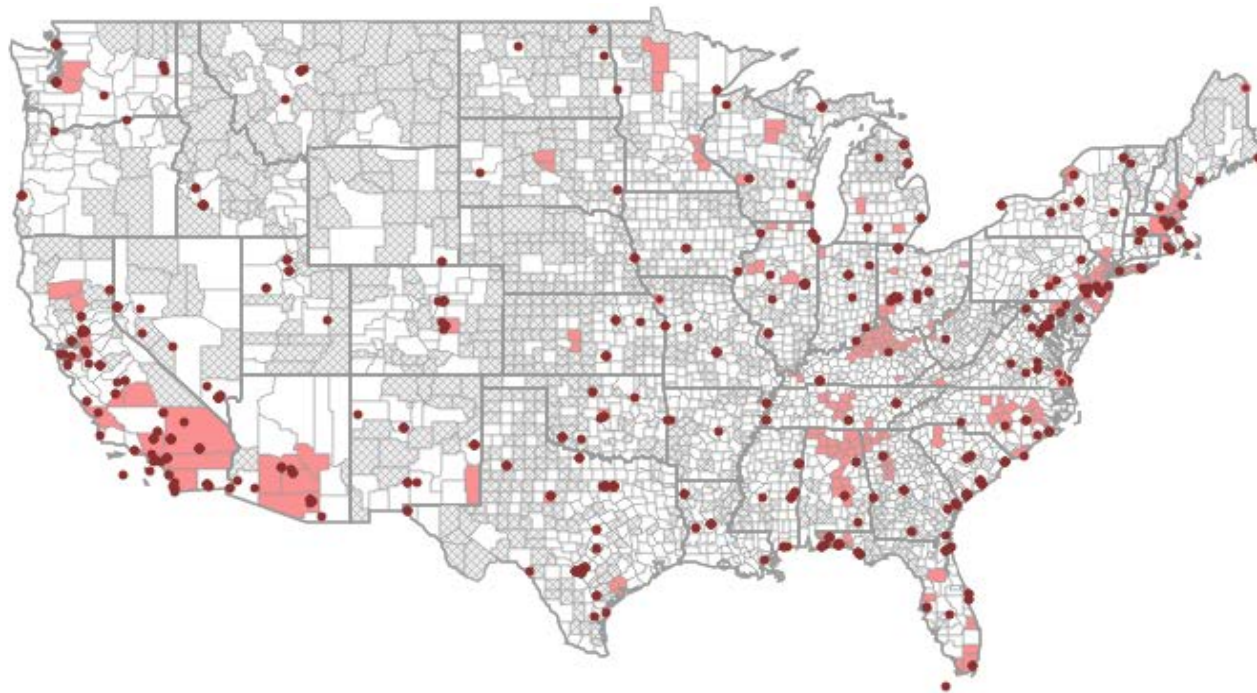
Dr. Richard (Hunter) Anderson
AFCEC/CZTE
February 2017

PFAS Occurrence



2012: Six PFASs added to Unregulated Contaminant Monitoring Rule 3 (UCMR 3) list, including PFOS and PFOA using **EPA 537 method**.

October 2015 UCMR 3 data summary



Contaminant	Minimum Reporting Level
PFOS	0.04 µg/L
PFOA	0.02 µg/L
PFNA	0.02 µg/L
PFHxS	0.03 µg/L
PFHpA	0.01 µg/L
PFBS	0.09 µg/L

PFAS Contamination in US Drinking Water

- PFCs detected
- No PFCs detected
- ▨ Not tested
- U.S. Military Fire/Crash Training Area Sites

Data visualization: Moiz Syed. Sources: EPA and Department of Defense.
<https://theintercept.com/2015/12/16/toxic-firefighting-foam-has-contaminated-u-s-drinking-water-with-pfcs/>

EPA 537 Method Used - Summary

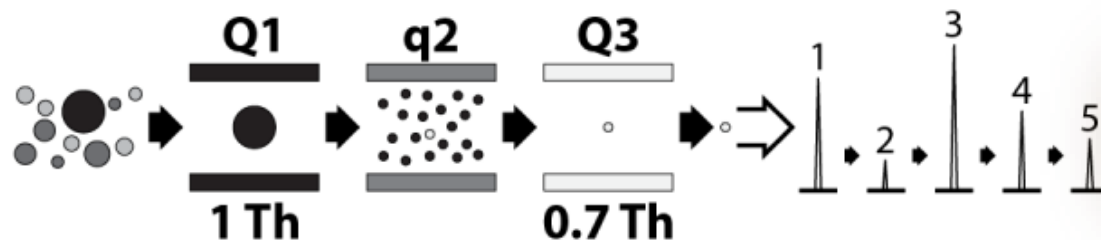


- A 250-mL preserved water sample with Trizma is fortified with surrogates and passed through a solid phase extraction (SPE) cartridge containing **Sorex HRPHS** in lieu of polystyrenedivinylbenzene (SDVB) to extract the method analytes and surrogates. The compounds are eluted from the solid phase with a small amount of methanol. The extract is concentrated to dryness with nitrogen in a heated water bath, and then adjusted to a 1-mL volume with 96:4% (vol/vol) methanol:water after adding the IS(s). A **5- μ L** in lieu of 10- μ L injection is made into an LC equipped with a C18 column that is interfaced to an
- **Q-Exactive hybrid HRAM capable of producing MS/MS data** in lieu of “low resolution triple” -MS/MS. The analytes are separated and identified by comparing the acquired mass spectra and retention times to reference spectra and retention times for calibration standards acquired under identical LC/MS/MS conditions. The concentration of each analyte is determined by using the internal standard technique. Surrogate analytes are added to all Field and QC Samples to monitor the extraction efficiency of the method analytes.

Hybrid HRAM can be used for EPA LC-MS/MS methods using Parallel Reaction Monitoring (PRM) - Targeted MS2

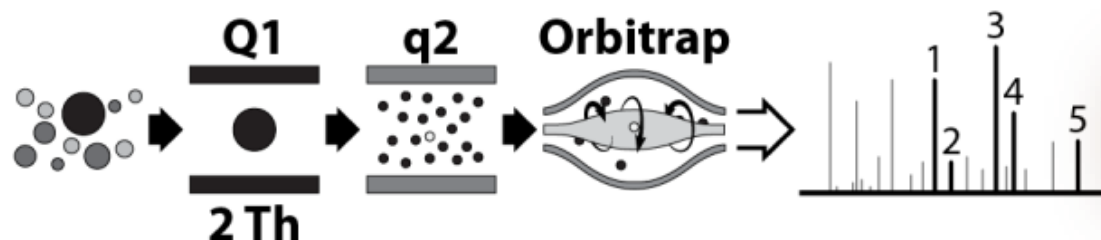


A SRM



Serial monitoring

B PRM



Parallel monitoring

Parallel reaction monitoring for high resolution and high mass accuracy quantitative, targeted proteomics. Peterson et al., MCP 2012, O112.020131

Q-Exactive tune page the set up.



040216-IDOC-PRM-FS-006
10

04/02/16 02:11:23

ICAL-5

040216-IDOC-PRM-FS-006
Q Exactive - Orbitrap MS

FULL MS - SIM

Full Scan Set up

General

Runtime

0 to 19.2 min

Polarity

Negative

In-source CID

0.0 eV

Full MS - SIM

70000 Resolution

Microscans

1

Resolution

70,000

AGC target

1e6

Maximum IT

100 ms

Number of scan ranges

1

Scan range

100 to 1100 m/z

Spectrum data type

100-1000 m/z

Profile

PRM

PRM Set up

General

Runtime

0 to 19.2 min

Polarity

Negative

In-source CID

0.0 eV

Default charge state

1

Inclusion

on

MSⁿ

Microscans

1

Resolution

35,000

AGC target

2e5

Maximum IT

100 ms

MSX count

1

MSX isochronous ITs

on

Isolation window

Isolation window

1.0 m/z

Isolation offset

0.0 m/z

Fixed first mass

-

NCE / stepped NCE

10, 55

Spectrum data type

Profile

Setup

TUNEFILES

General

Switch Count 0

Base Tunefile C:\Xcalibur\methods\600UL-EPA-537-TUNE.mstune

CONTACT CLOSURE

No need for optimization

**Targeted +
Non Targeted**

Targeted

Needs RT and specific collision E.

EPA 537 – FULL-MS at 70K resolution showing good peak shapes, and S/N for a 2.5ppt standard.

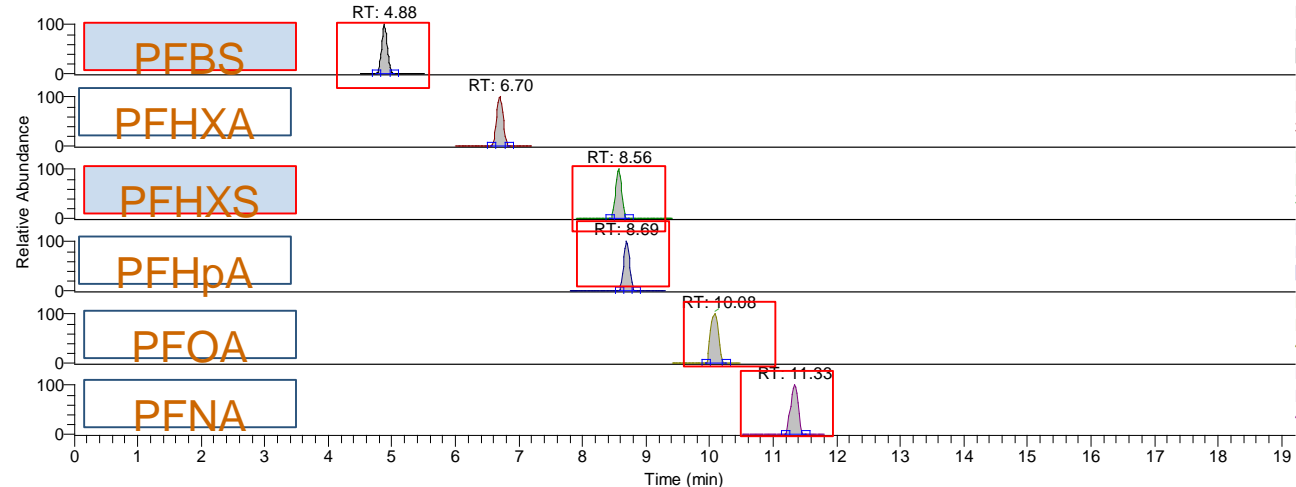


040216-IDOC-PRM-FS-003
1.0

04/02/16 01:11:20

ICAL-2

RT: 0.00 - 19.20 SM: 7B



NL: 2.83E4

Base Peak m/z= 79.95564-79.95644 F: FTMS - p ESI Full ms2 298.94@hcd55.00 [50.00-325.00] MS Genesis 040216-IDOC-PRM-FS-003

NL: 1.43E4

Base Peak m/z= 268.98166-268.98434 F: FTMS - p ESI Full ms2 312.97@hcd10.00 [50.00-335.00] MS Genesis 040216-IDOC-PRM-FS-003

NL: 2.39E4

Base Peak m/z= 318.97821-318.98139 F: FTMS - p ESI Full ms2 362.97@hcd10.00 [50.00-390.00] MS Genesis 040216-IDOC-PRM-FS-003

NL: 2.29E4

Base Peak m/z= 79.95564-79.95644 F: FTMS - p ESI Full ms2 398.94@hcd50.00 [50.00-425.00] MS Genesis 040216-IDOC-PRM-FS-003

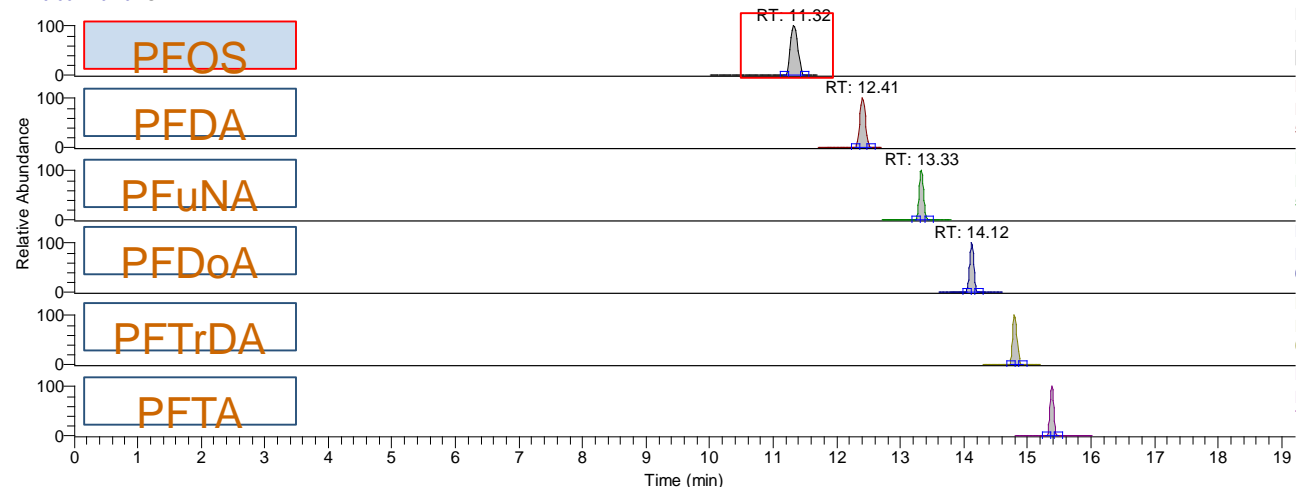
NL: 2.29E4

Base Peak m/z= 368.97481-368.97849 F: FTMS - p ESI Full ms2 412.97@hcd10.00 [50.00-440.00] MS Genesis 040216-IDOC-PRM-FS-003

NL: 2.03E4

Base Peak m/z= 418.97142-418.97560 F: FTMS - p ESI Full ms2 462.96@hcd10.00 [50.00-490.00] MS Genesis 040216-IDOC-PRM-FS-003

RT: 0.00 - 19.20 SM: 7B



NL: 2.04E4

Base Peak m/z= 79.95565-79.95645 F: FTMS - p ESI Full ms2 498.93@hcd55.00 [50.00-525.00] MS Genesis 040216-IDOC-PRM-FS-003

NL: 2.74E4

Base Peak m/z= 468.96778-468.97246 F: FTMS - p ESI Full ms2 512.96@hcd10.00 [50.00-540.00] MS Genesis 040216-IDOC-PRM-FS-003

NL: 3.13E4

Base Peak m/z= 518.96439-518.96957 F: FTMS - p ESI Full ms2 562.96@hcd10.00 [50.00-590.00] MS Genesis 040216-IDOC-PRM-FS-003

NL: 2.58E4

Base Peak m/z= 568.96097-568.96665 F: FTMS - p ESI Full ms2 612.95@hcd10.00 [50.00-645.00] MS Genesis 040216-IDOC-PRM-FS-003

NL: 2.42E4

Base Peak m/z= 618.95754-618.96372 F: FTMS - p ESI Full ms2 662.95@hcd10.00 [50.00-695.00] MS Genesis 040216-IDOC-PRM-FS-003

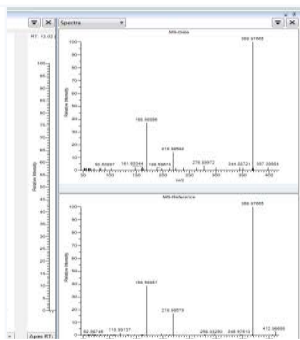
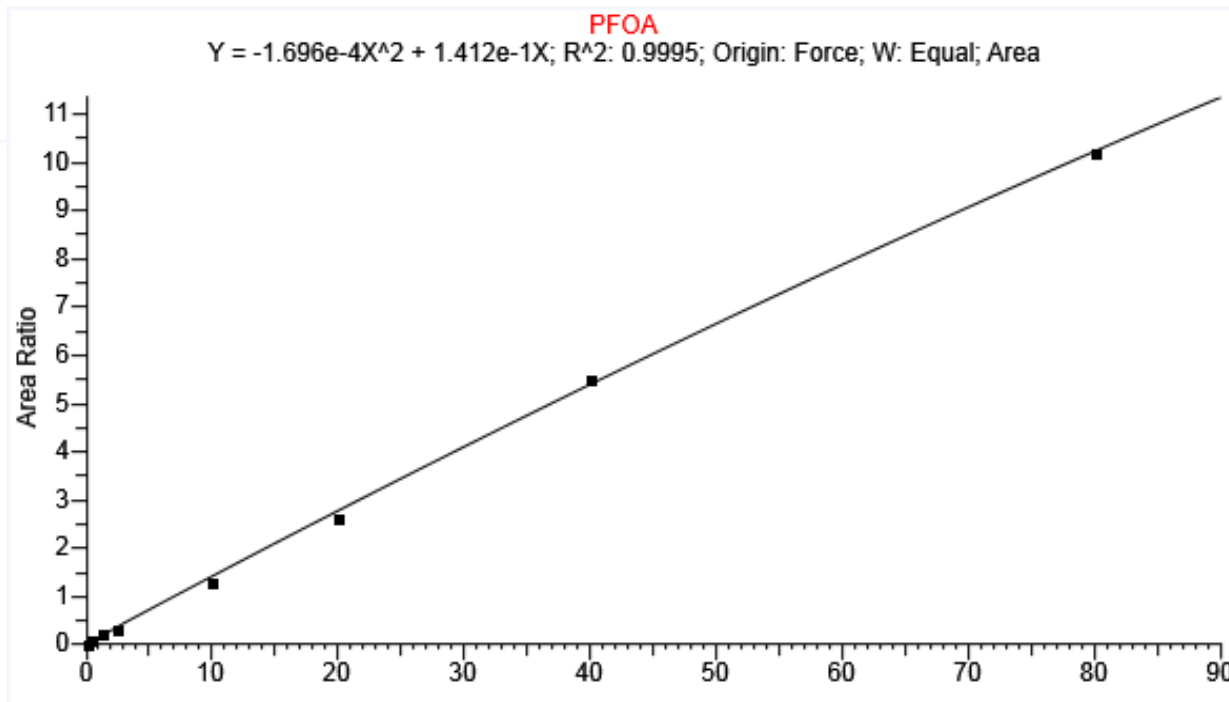
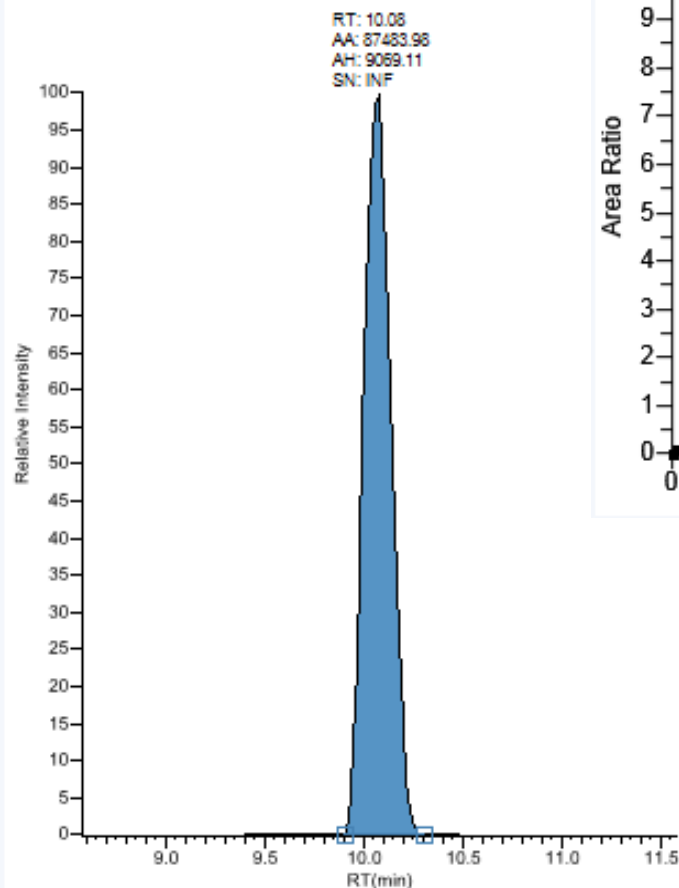
NL: 2.06E4

Base Peak m/z= 668.95430-668.96098 F: FTMS - p ESI Full ms2 712.95@hcd10.00 [50.00-745.00] MS Genesis 040216-IDOC-PRM-FS-003

EPA 537M (PRM) – targeted-MS2 for PFOA at the lowest cal std 0.5ppt shown with spectra confirmation using Trace Finder 4.1



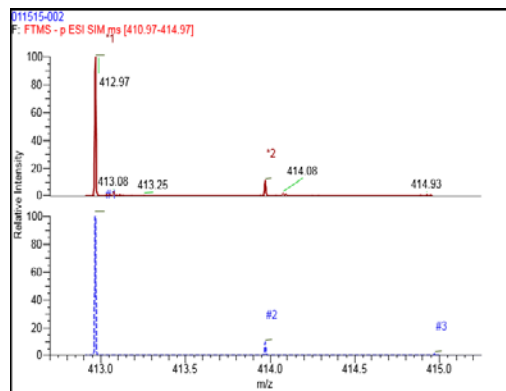
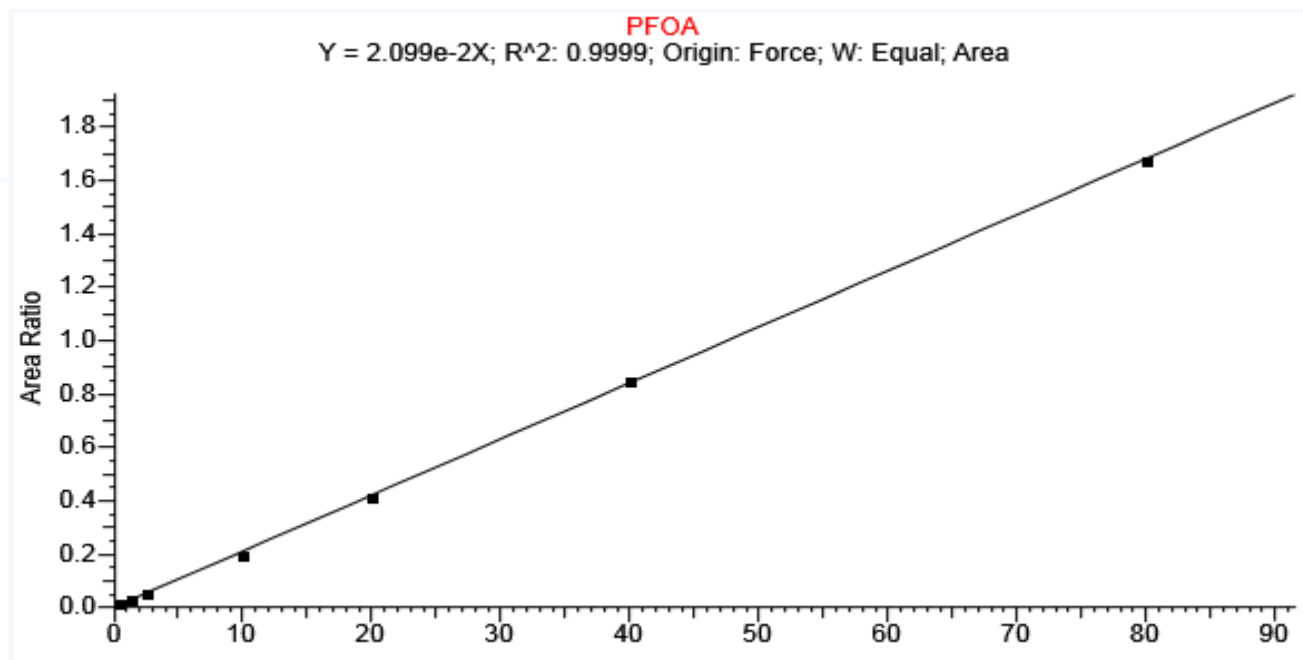
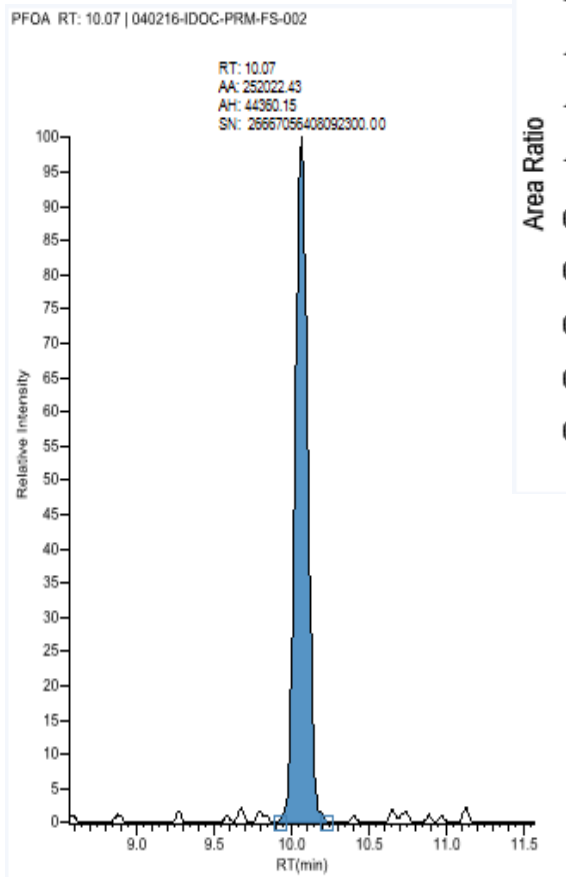
PFOA RT: 10.08 | 040216-IDOC-PRM-FS-002



← Sample MS2 Spectra

← Library MS2 Spectra

By adding Full-Scan to PRM workflow – 0.5ppt PFOA shown



← Isotopic pattern match

Determination of Minimum Reporting Limit Using LCMRL Using Regretional LOQ Calculation



Enter Test Data

Contaminant - 17 alpha-Ethynylestradiol

Method - EPA 539

Units for all measurements - Nanograms/Liter (ng/L)

NB – Calculator only works with 32 bit computers.

****Important** Please do not truncate or round any of the results. If possible, results should have a minimum of 3 significant digits.**

Use the buttons below to add a concentration (column) or measured value (row). To remove a row or column check the box next to it and click the Remove button.

↓

		Concentrations							
		<input type="checkbox"/> Conc. 1	<input type="checkbox"/> Conc. 2	<input type="checkbox"/> Conc. 3	<input type="checkbox"/> Conc. 4	<input type="checkbox"/> Conc. 5	<input type="checkbox"/> Conc. 6	<input type="checkbox"/> Conc. 7	<input type="checkbox"/> Conc. 8
		0.225	0.45	0.9	1.8	3.6	5.4	7.2	0
Measured Concentrations									
<input type="checkbox"/> Value 1	0.184	0.347	0.684	1.361	2.657	4.469	6.07	0.003	
<input type="checkbox"/> Value 2	0.184	0.35	0.728	1.365	2.819	4.551	5.814	0.000	
<input type="checkbox"/> Value 3	0.202	0.36	0.705	1.363	2.66	4.323	6.046	0.000	
<input type="checkbox"/> Value 4	0.174	0.4	0.534	1.362	2.987	4.207	5.563	0.000	

http://water.epa.gov/scitech/drinkingwater/labcert/analyticalmethods_ogwdw.cfm

<< Back

Next >>

The LCMRL is defined as the lowest spiking concentration at which recovery of between 50 and 150 percent is expected 99 percent of the time by a single analyst. The procedure requires, at a minimum, four replicates at each of seven fortification levels. Four laboratory reagent blanks should also be included. All must be processed through the entire method procedure

Target & non targeted compounds EPA 537 plus.



PRM

EPA 537 PFCA's, and PFSA'S target list

LCMRL equal or better than high end - mid range triple quads – background contamination is the limiting factor.

	Critical level	DL	LCMRL
PFBS	0.077	0.12	<0.5
PFDA	0.18	<0.5	<0.5
PFDoA	0.14	0.29	<0.5
PFHpA		0.35	0.97
PFHxA	0.16	0.27	<0.5
PFHxS		0.52	0.77
PFNA	0.14	0.26	<0.5
PFOA		0.36	0.5
PFOS	0.14	0.21	<0.5
PFTA		0.48	0.71
PFTrDA	0.18	0.32	<0.5
PFuNA		0.31	0.72

Full Scan

	Critical level	DL	LCMRL
PFBS	0.15	0.2	<0.5
PFDA	0.15	0.26	<0.5
PFDoA		0.47	0.73
PFHpA	0.09	0.15	<0.5
PFHxA	0.13	0.19	<0.5
PFHxS		1.7	2.4
PFNA	0.11	0.17	<0.5
PFOA		0.22	0.5
PFOS		0.26	0.5
PFTA	0.15	0.2	<0.5
PFTrDA		0.31	0.55
PFuNA		0.38	1
PFBA		0.19	0.64
PFODA		0.55	1
PFDS	0.13	0.19	<0.5
PFHxDA		0.12	0.5
PFPA	0.18	0.19	<0.5

Stock standard contained other compounds not part of EPA 537 target list which were identified and quantified using Full-MS

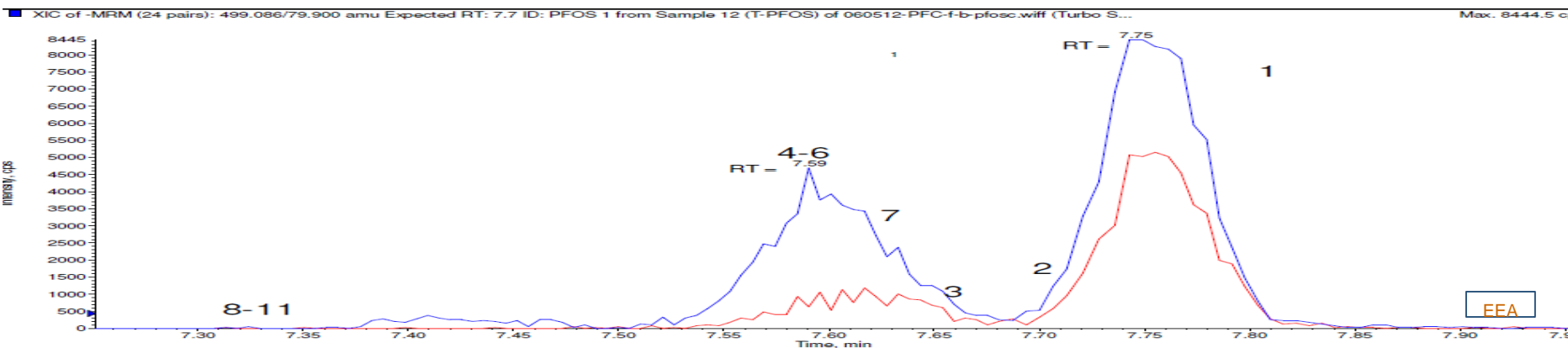


PFOS quantification is challenging: Showing a technical grade standard



Isomer 1 2 3 4 5 6 7 8 9 10 11

		MS/MS relative response factors										
↓	m/z 499 m/z 99	100	117	97	49	39	43	78	10	0	0	19
↑	m/z 499 m/z 80	100	0	78	135	241	142	123	113	118	220	90



1) $\text{CF}_3\text{CF}_2\text{CF}_2\text{CF}_2\text{CF}_2\text{CF}_2\text{CF}_2\text{CF}_2\text{SO}_3$ 2) $\text{CF}_3\text{CF}_2\text{CF}_2\text{CF}_2\text{CF}_2\text{CF}_2\text{CF}_2\text{SO}_3$ 3) $\text{CF}_3\text{CF}_2\text{CF}_2\text{CF}_2\text{CF}_2\text{CF}_2\text{CF}_2\text{SO}_3$ 4) $\text{CF}_3\text{CF}_2\text{CF}_2\text{CF}_2\text{CF}_2\text{CF}_2\text{CF}_2\text{SO}_3$



5) $\text{CF}_3\text{CF}_2\text{CF}_2\text{CF}_2\text{CF}_2\text{CF}_2\text{CF}_2\text{SO}_3$



6) $\text{CF}_3\text{CF}_2\text{CF}_2\text{CF}_2\text{CF}_2\text{CF}_2\text{CF}_2\text{SO}_3$

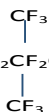


7) $\text{CF}_3\text{CF}_2\text{CF}_2\text{CF}_2\text{CF}_2\text{CF}_2\text{CF}_2\text{SO}_3$



8) $(\text{CF}_3)_3\text{CF}_2\text{CF}_2\text{CF}_2\text{CF}_2\text{CF}_2\text{SO}_3$

9) $\text{CF}_3\text{CF}_2\text{CF}_2\text{CF}_2\text{CF}_2\text{CF}_2\text{CF}_2\text{SO}_3$



10) $\text{CF}_3\text{CF}_2\text{CF}_2\text{CF}_2\text{CF}_2\text{CF}_2\text{CF}_2\text{SO}_3$



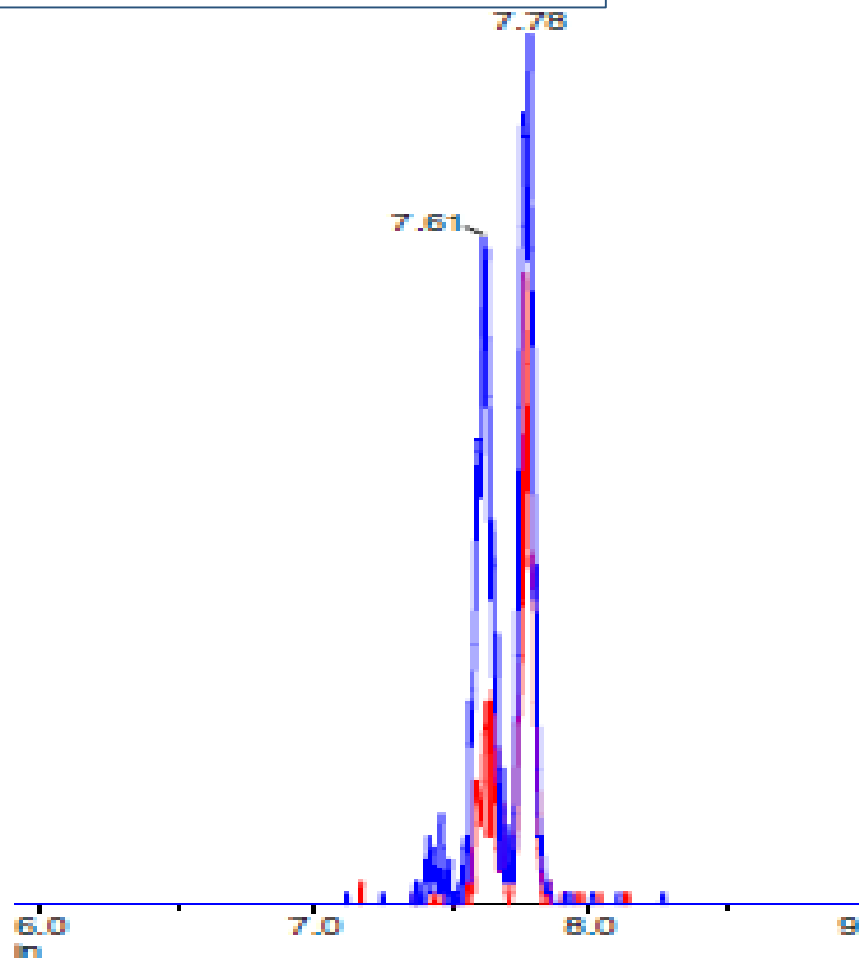
11) $\text{CF}_3\text{CF}_2\text{CF}_2\text{CF}_2\text{CF}_2\text{CF}_2\text{CF}_2\text{SO}_3$



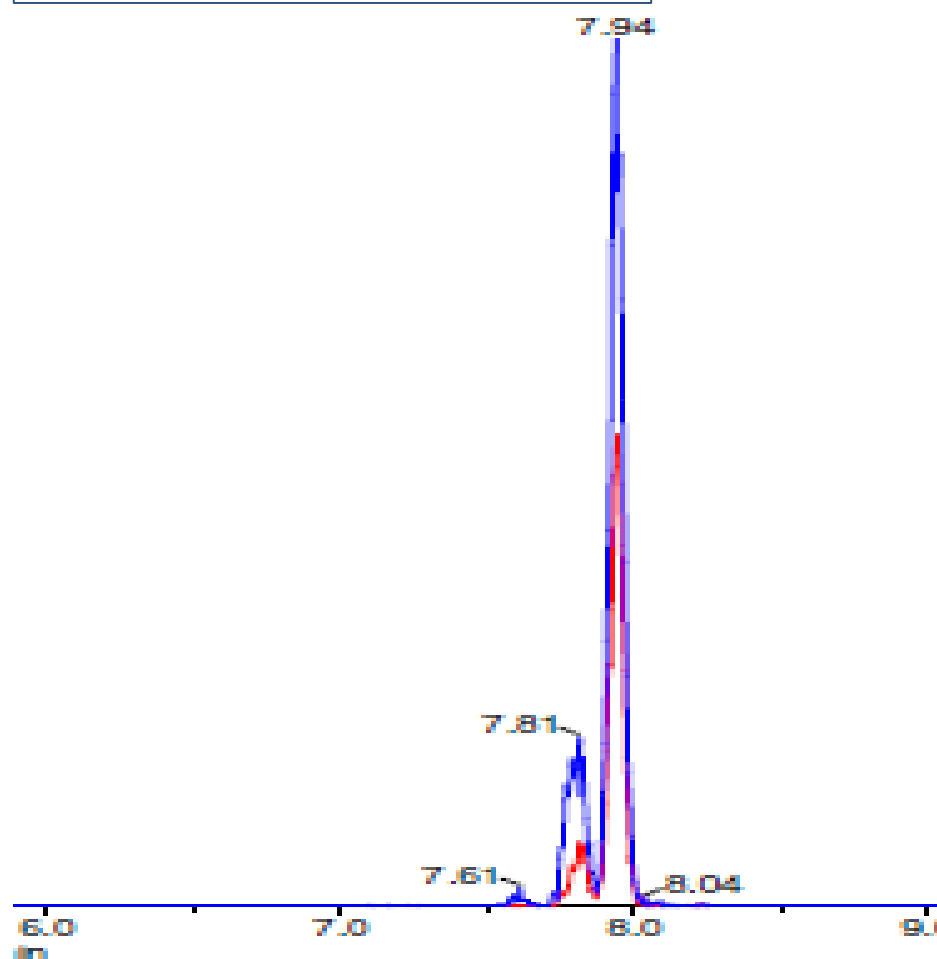
Samples from different locations can have different branch ratios



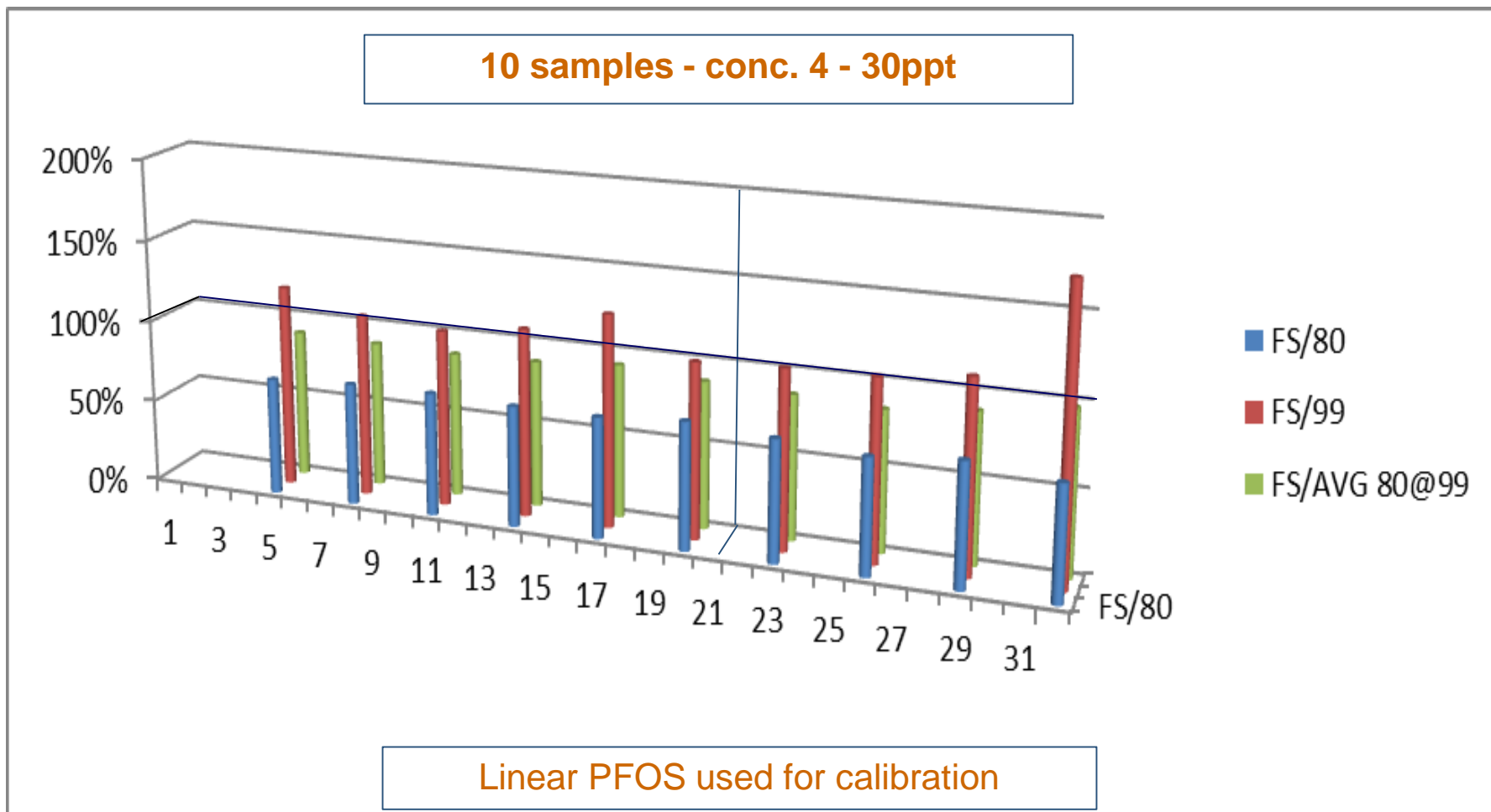
Field Sample



Std PFOS Branched

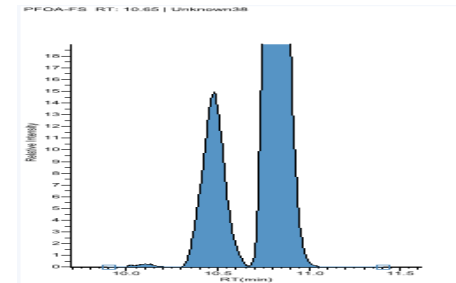
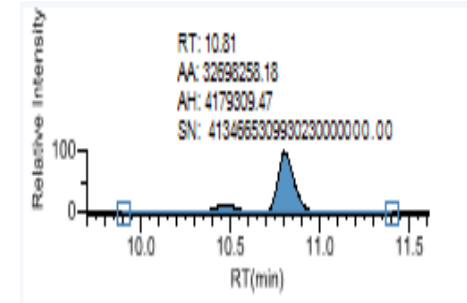


Fs scan covers all of the branches and looks to be more reliable for PFOS quant.





Technical Advisory- Laboratory Analysis of Drinking Water Samples for Perfluorooctanoic Acid (PFOA) Using EPA Method 537 Rev. 1.1



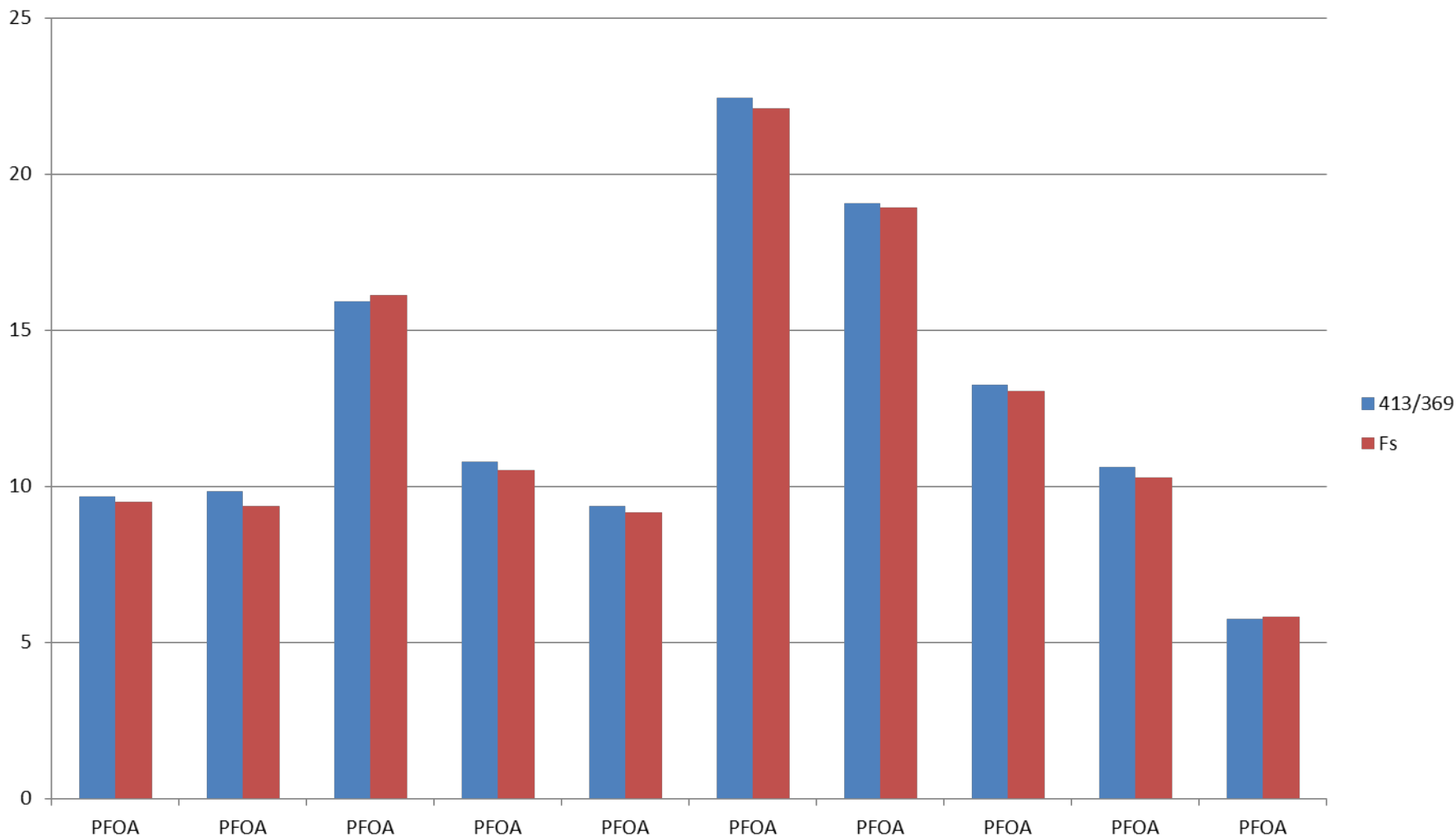
Office of Water (MS-140)

EPA 815-B-16-021

September 2016

Till around 1970 PFOA was also produced by Electrochemical fluorination (ECF) process creating branched isomers which can still be detected occasionally so it is important to also integrate the branch isomers.

FS and 413/369 compares well in real field samples having branched PFOA - ppt

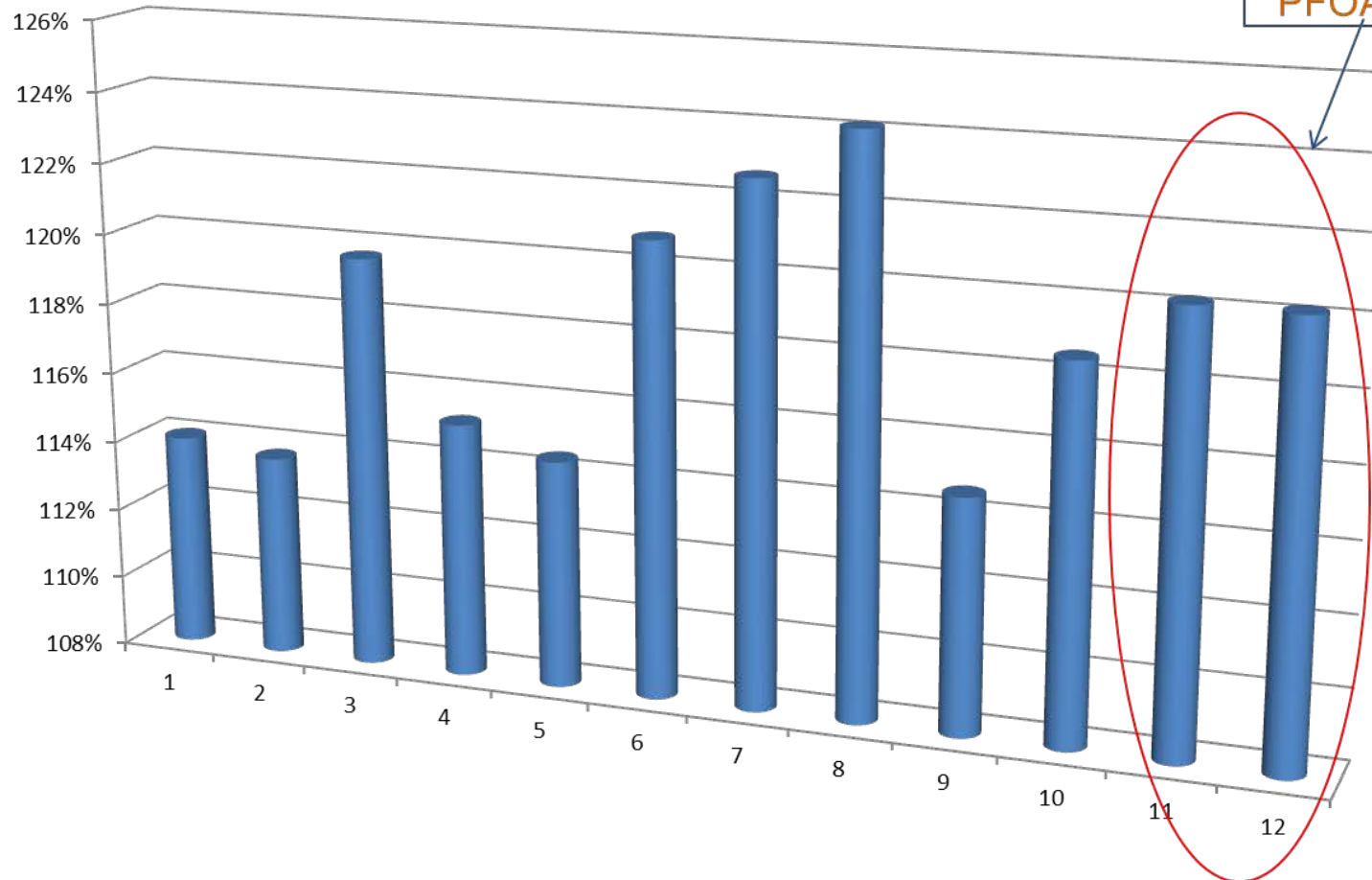


Including branched isomers when present about 20% diff. in a technical grade



PFOA – Including Branched Isomers/linear integration

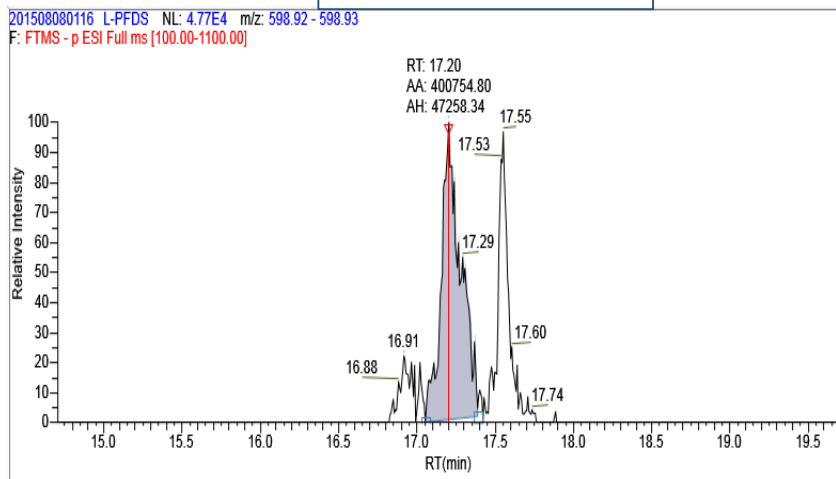
Technical grade PFOA standard



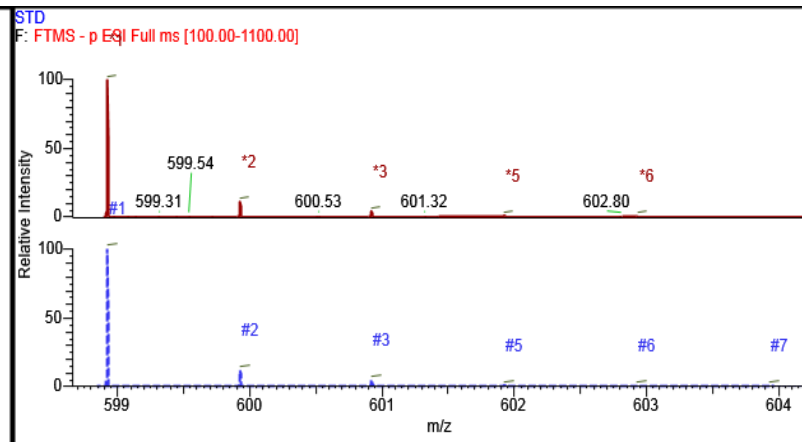
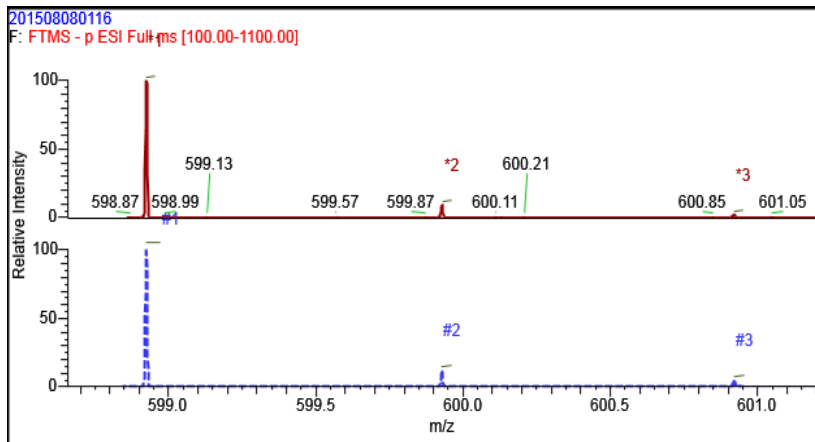
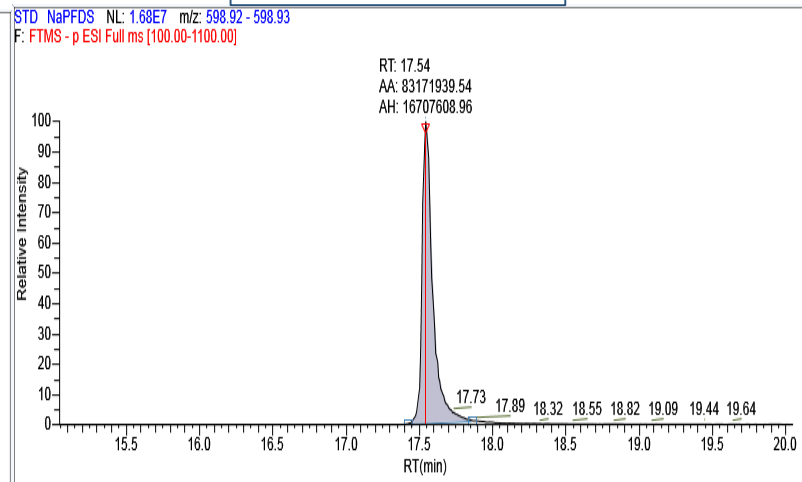
A UCMR3 sample shown having a trace hits for non-targeted known compound. PFDS



Sample



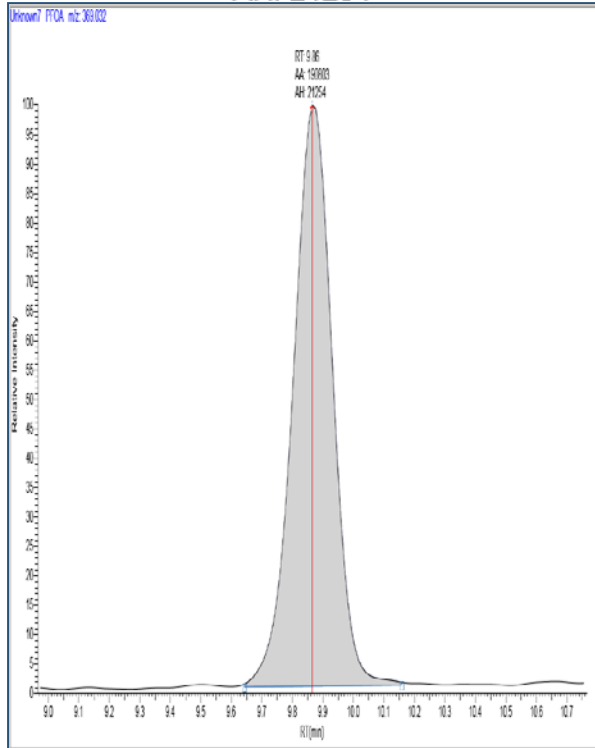
80 ppt STD



A 2.5ppt standard of PFOA. Excellent quantitation and sensitivity is obtained with HRAM in comparison to QQQ analysis.

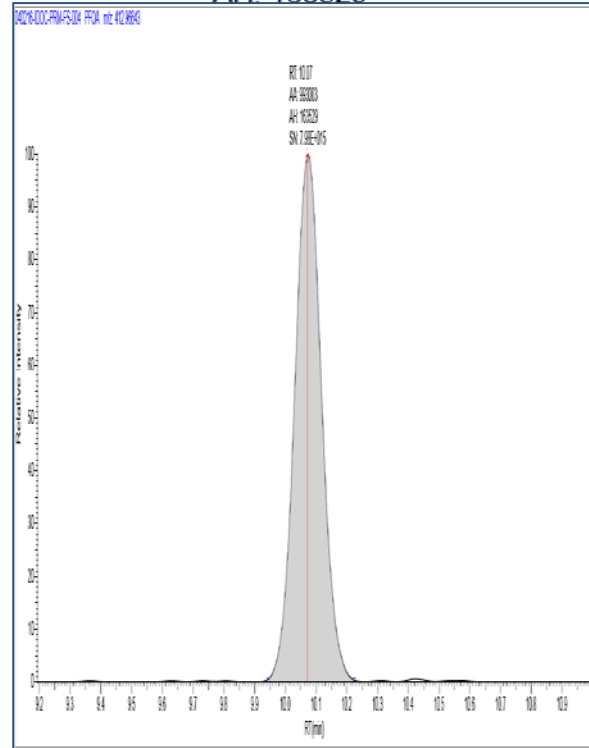


RT: 9.86
AA: 190803
AH: 21254



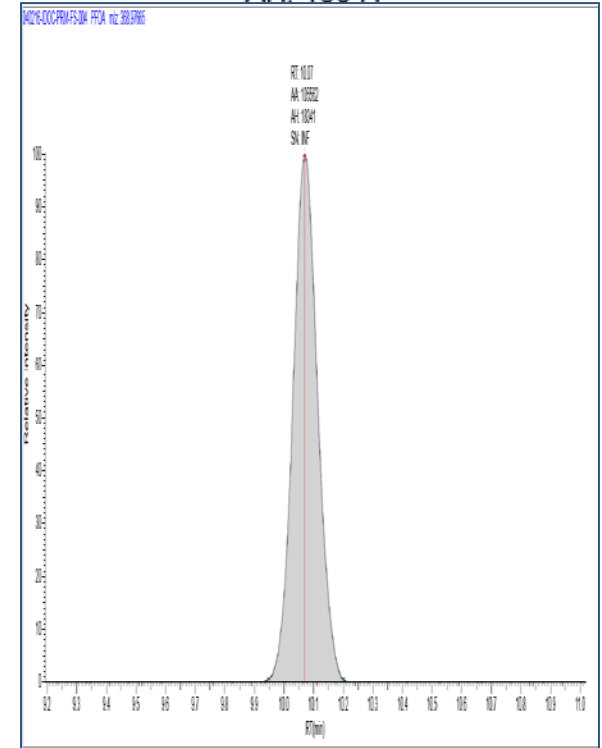
a) SRM Analysis

RT: 10.07
AA: 993003
AH: 163529



b) HRAM Full Scan

RT: 10.07
AA: 105562
AH: 18341



c) HRAM PRM

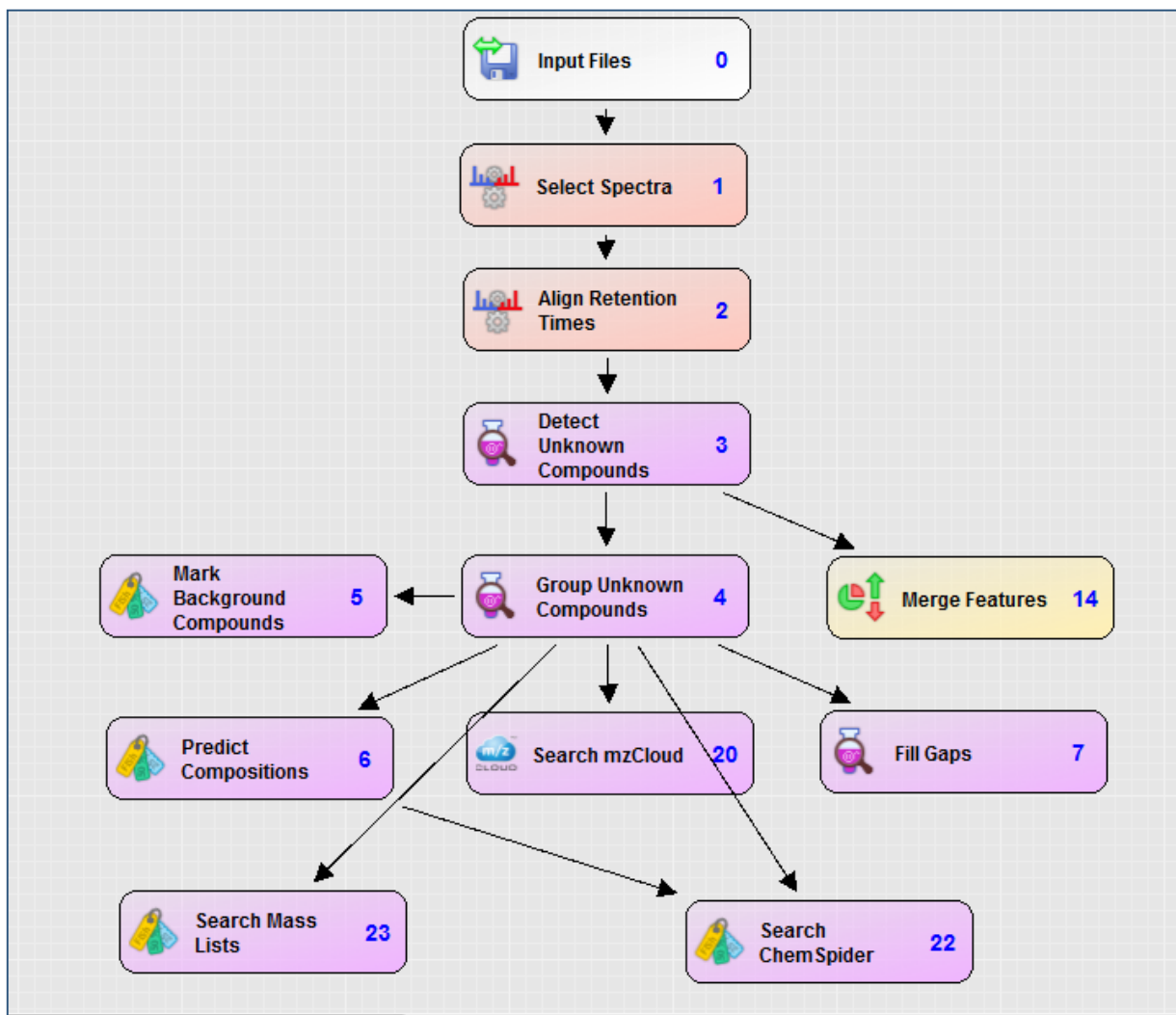
Looking for Unknowns : Contaminated GW in a AAAF site:

On-line SPE UHPLC / Fs-ddms2, top 5

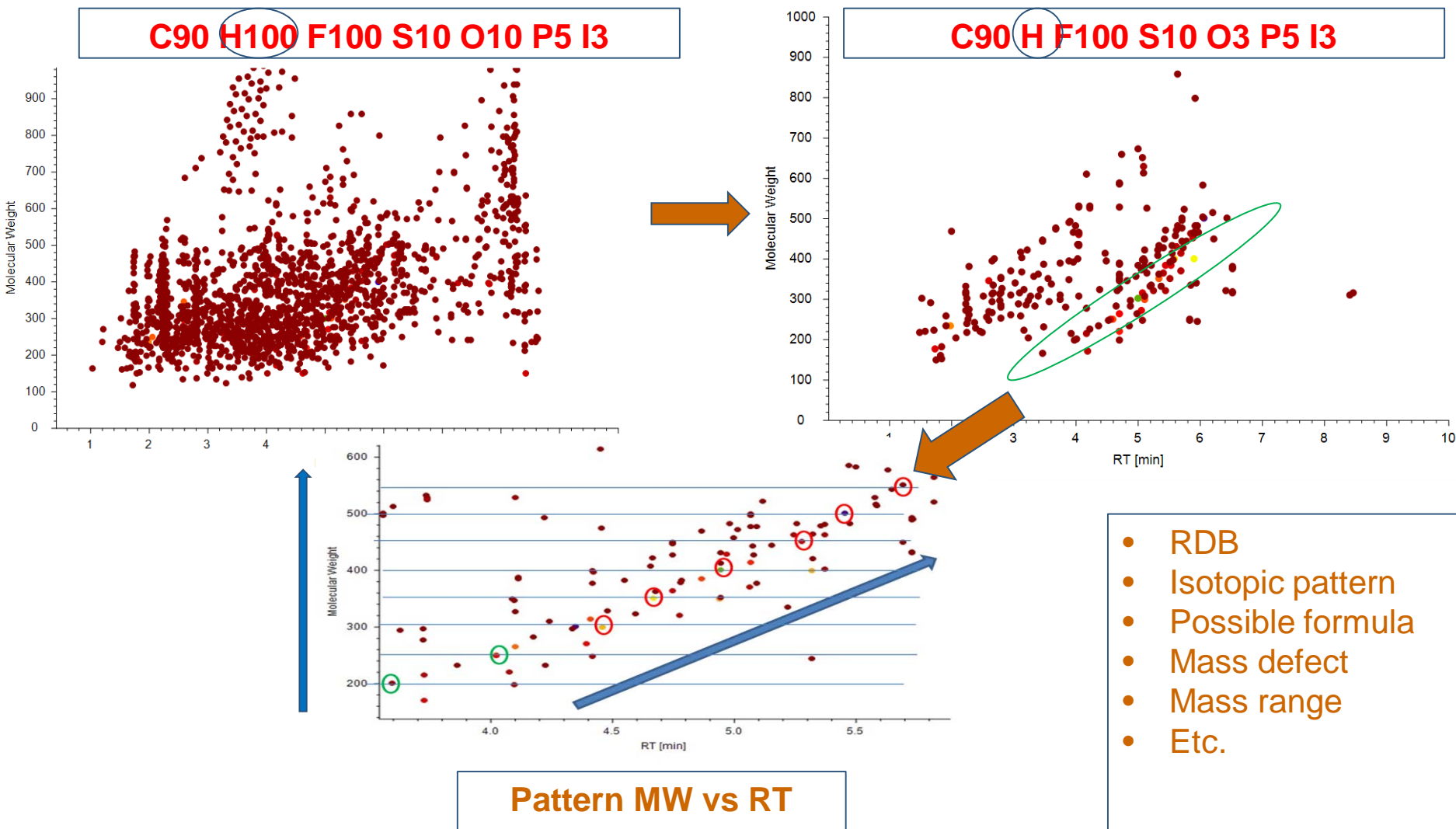


The screenshot displays a software interface for mass spectrometry. On the left, there are panels for 'Global Lists' (Lock Masses, Inclusion, Exclusion, Neutral Loss, Tag Masses), 'Tune Files' (C:\scalbur\methods\200UL-FLOW-TUNE.mstune), 'External Hardware' (Divert Valve a, b, Syringe, Contact Closure), 'Chromatogram' (Scan Groups), and 'Experiments' (General, Full MS - SIM, AIF, Full MS / AIF, Full MS / dd-MS² (TopN), Targeted-SIM, PRM, Targeted-SIM / dd-MS², Full MS / AIF / NL dd-MS², DIA). The main area shows a chromatogram with a 'Top5' scan group highlighted. Below the chromatogram is a detailed schematic of the Orbitrap Mass Analyzer, including components like the NanoSpray Source, Capillary, S-Lens, S-Lens Exit Lens, Injection Flatapole, Inter-Flatapole Lens, TK Lens, Bent Flatapole, Quadrupole, Quad Exit Lens, Split Lens, C-Trap Entrance Lens, C-Trap, and HCD Collision Cell. On the right, a 'Properties' panel shows settings for the method, including 'Global Settings' (User Role: Advanced, Use lock masses: off), 'Properties of Full MS / dd-MS²' (General: Runtime 1 to 9 min, Polarity negative, In-source CID 0.0 eV, Default charge st 1; Full MS: Microscans 1, Resolution 140,000, AGC target 1e6, Maximum IT 100 ms; dd-MS² / dd-SIM: Microscans 1, Resolution 35,000, AGC target 2e5, Maximum IT 50 ms, Loop count 5, MSX count 1, TopN 5, Isolation window 4.0 m/z, Isolation offset 0.0 m/z, Fixed first mass -, (N)CE / stepped + nce: 20, 40, 80, Spectrum data by Profile), and 'dd Settings' (Minimum AGC tai 6.00e3, Intensity threshold 1.2e5, Apex trigger -, Charge exclusion -, Peptide match -, Exclude isotopes -). The bottom status bar shows 'Experiment Setup Summary' and 'ready'.

Data mining software using “Compound Discoverer”



Refining data to explore known classes of compounds



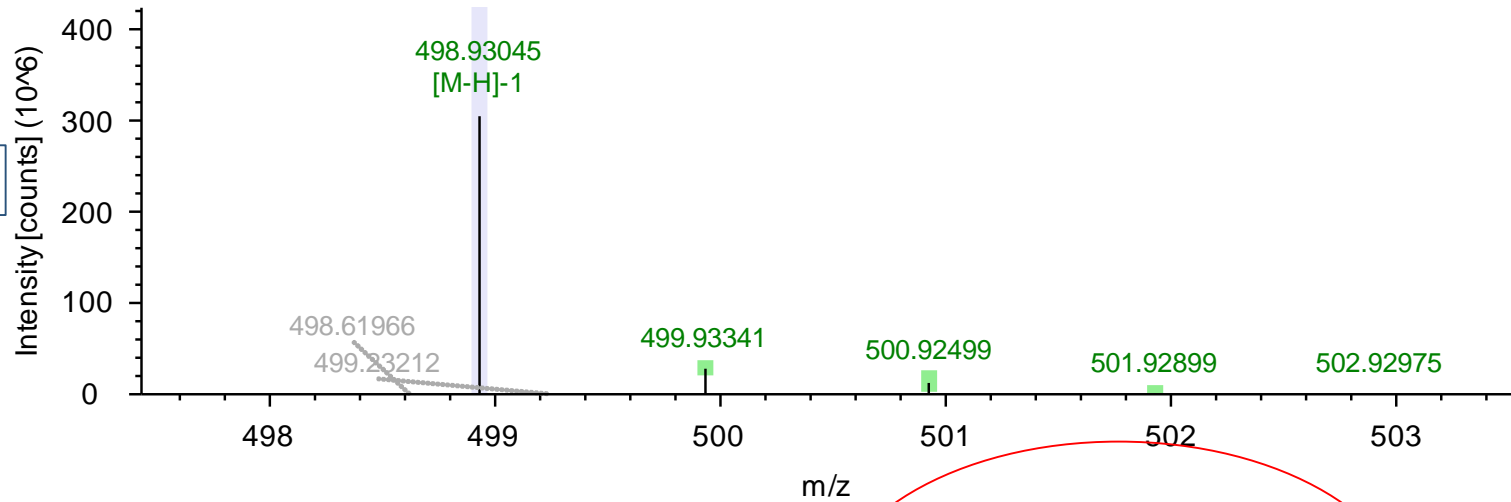
- RDB
- Isotopic pattern
- Possible formula
- Mass defect
- Mass range
- Etc.

At higher resolutions more trace isotopic pattern can be used for Ms scans.

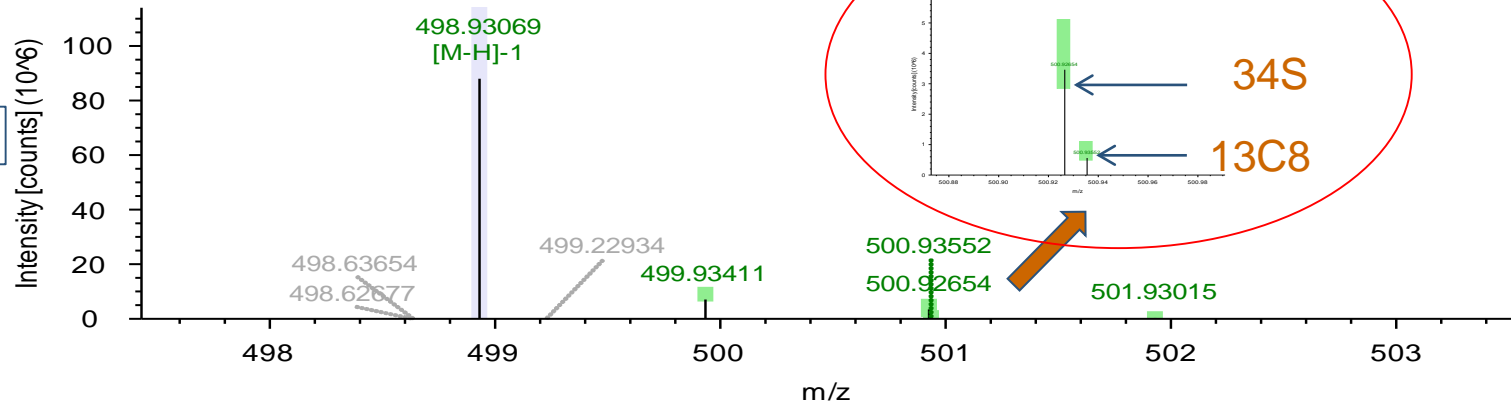


201605240377R, #2094, RT=5.457 min, FTMS (-)
C8 H F17 O3 S as [M-H]-1

70K Res.



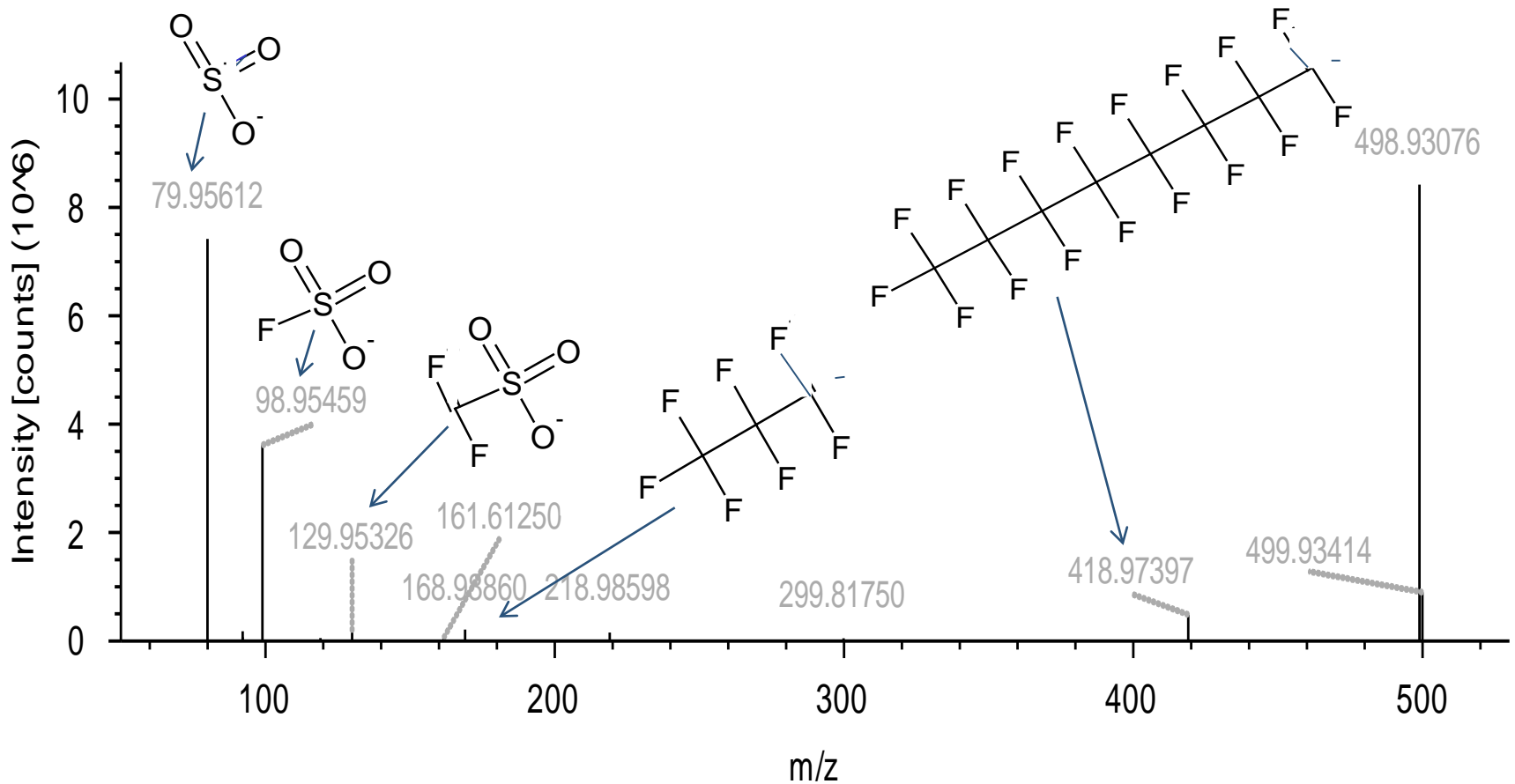
140K Res.



Ms2 Data Shows CmF2m+1- “9 series” and CnF2nSO3- radical “0 series” as expected.



201605240376, #1272, RT=6.094 min, FTMS (-), MS2 (HCD, DDF, 498.93@50.00, z=-1)



The structure of selected compound can be drawn in “Custom Explanation” using Mass Frontier to check against MS and MS2 collected data



Chromatograms

Group By: Sample Type (1/1), Sample (1/1)

Filter By: Sample Type, Sample

C6HF11O4S C6 H F11 O4 S MW: 377.94199
File: 201605240383.raw (F10) FTMS (-) MS1

Mass Spectrum

#1111, RT=5.451 min, FTMS (-)
#1115, RT=5.467 min, FTMS (-)
#1118, RT=5.480 min, FTMS (-), MS2 (HCD, DD)

201605240383, #1118, RT=5.480 min, FTMS (-), MS2 (HCD, DDF, 376.94@50.00, z=-1)
C6HF11O4S C6 H F11 O4 S, MW: 377.94199, Area: 331907
FISH Coverage: 3 Direct, 23 Unmatched, 8 Skipped

Custom Explanation Editor

Description: Molecular weight: 377.94199, Formula: C6 H F11 O4 S, Name: C6HF11O4S

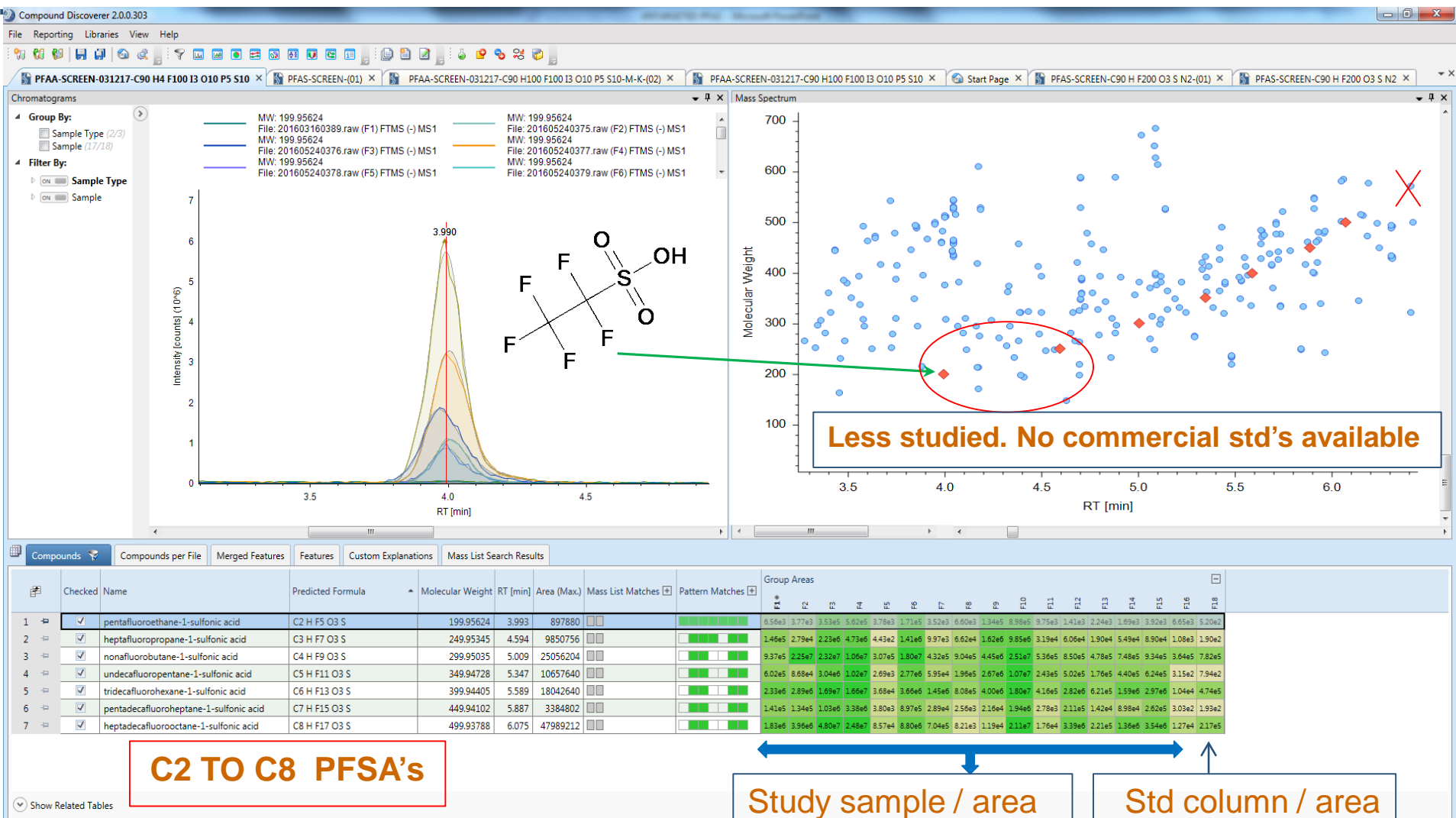
FISH Scoring: Molecular weight (original): 377.94246, Formula (original): C6 H F11 O4 S

Save Cancel

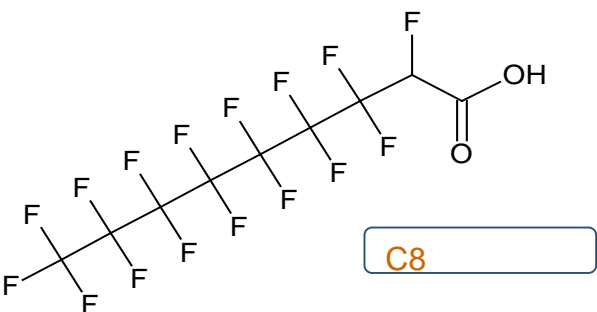
Checked	Structure	Name	Formula	Molecular Weight	Comments
<input type="checkbox"/>		C9 H3 F16 O3 P		493.95643	NO MATCH MS2
<input type="checkbox"/>		C9 H2 F16 O2		445.97993	MS2 NOT MATCHES
<input checked="" type="checkbox"/>		C6HF11O4S	C6 H F11 O4 S	377.94199	
<input type="checkbox"/>		C6HF11O3S	C6 H F11 O3 S	361.94707	C4F7O=280.98346, 9
<input type="checkbox"/>		C4HF7O4S	C4 H F7 O4 S	277.94838	4,798

(min)	Best Sfit [%]	Max. # MI	# Adducts	Area	Study File ID	FISH Coverage
54	38	2	1	191959	F2	0.00
54	54	2	1	108988	F2	0.96
52	52	4	1	331907	F10	11.54
48	48	3	1	395253	F10	15.38
54	54	3	1	49103	F10	

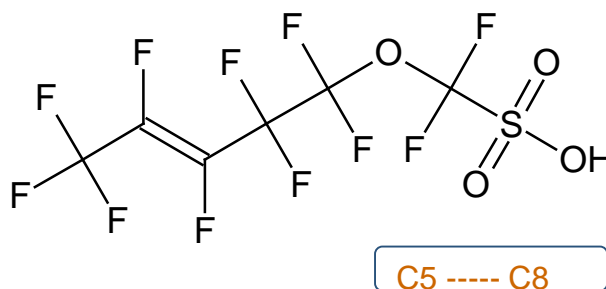
Summary of all PFSA's found for the studied group.



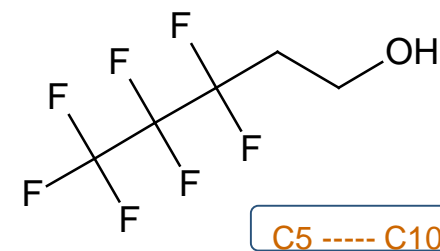
Few identified suspects using prescribed workflow which were missing from the built in library. Some need additional confirmations.



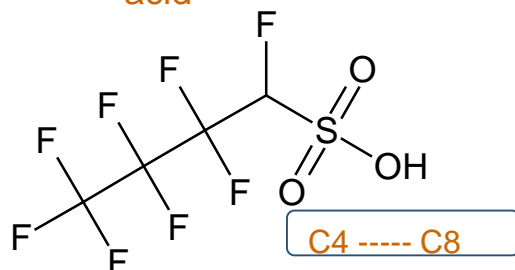
hexadecafluorononanoic acid



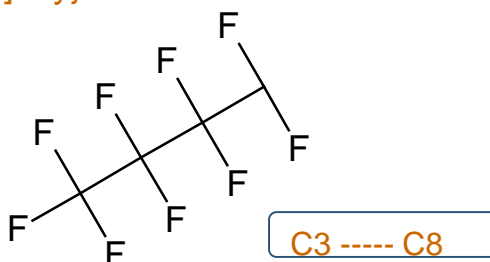
difluoro[[[(3E)-1,1,2,2,3,4,5,5,5-nonafluoropent-3-en-1-yl]oxy]methanesulfonic acid



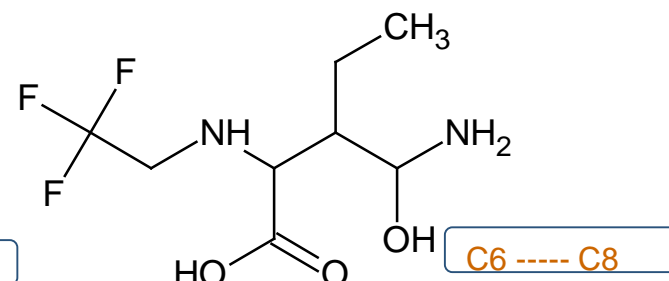
3:2 FTOH



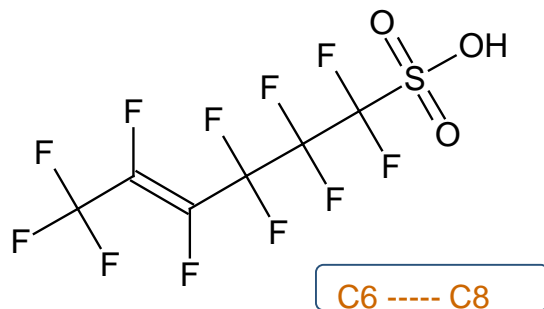
octafluorobutane-1-sulfonic acid



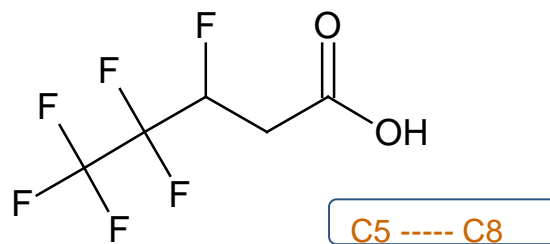
nonafluorobutane



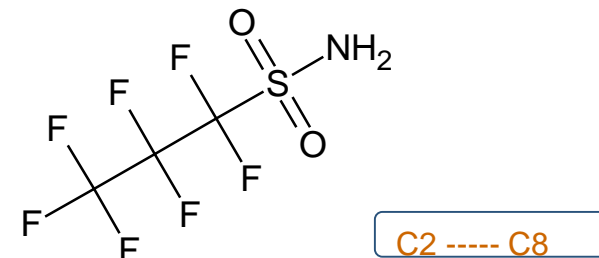
3-[amino(hydroxy)methyl]-2-[(2,2,2-trifluoroethyl)amino]pentanoic acid



undecafluorohex-4-ene-1-sulfonic acid



3:2 FTA



heptafluoropropane-1-sulfonamide

Conclusion



- **Q-Exactive HRAM instrumentation in the PRM scan mode can be used for quantitation with performance like a triple quadrupole in SRM mode with added specificity, selectivity and comparable sensitivity.**
- **Full scan HRAM can likely produce more accurate quantitative data for compounds that contain branched isomers such as PFOS.**
- **Routine quantitative workflows and non-target analysis can be performed in a single analysis.**
- **HRAM data processing using Thermo Fisher Scientific Compound Discoverer software can simplify complex data reduction and save time.**
- **Other techniques may be necessary for further confirmation of suspects/unknowns structures such as MSⁿ, ¹³C and ¹⁹F NMR, when standards are not commercially available.**

Any Questions?



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