

# Identification of Leachable Impurities from Pharmaceutical Container Closure Materials using Orbitrapbased GC-MS

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## Overview

The results of this study show that the Q Exactive GC system is an ideal analytical tool for extractables and leachables studies where both known and unknown compounds are encountered. Software tools enable fast and accurate differential analysis to be performed to isolate unique features in extracts. Routine mass resolution of 60,000 FWHM and consistent sub-ppm mass accuracy ensures selective and confident compound detection and identification.

## Introduction

The investigation of potentially toxic chemical impurities leaching from a wide variety of plastics, polymers and packaging products destined for pharmaceutical products has received a great deal of attention and remains a challenging analysis for chemists. Often termed extractables and leachables (E/L) studies, their aim is to identify, quantify and ultimately minimize any impurities that can migrate from packaging into a final product or drug. "Extractables" are those chemicals that can extract from components of a container closure system into solvents under accelerated laboratory conditions, such as elevated temperature and aggressive solvent, with the aim to extract the maximum compounds without deforming or degrading the material. "Leachables" are defined as chemicals that can migrate from the packaging into a drug product over the course of its shelf life.

Gas Chromatography-Mass Spectrometry (GC-MS) has been widely used in extractables studies as it provides analytical advantages of chromatographic resolution, reproducibility, peak capacity and importantly extensive spectral libraries to aid in identification. As packaging products can contain a large number of volatile and semi-volatile constituents it is well suited to analysis by GC-MS. In this study we seek to take advantage of a new class of GC-MS system with high mass resolution performance and exceptional mass accuracy for the detection and identification of compounds in polymer gaskets (O-rings) used in container closure systems and production seals. This work aims to demonstrate the application of a complete untargeted workflow to detect and identify chemical components in the O-rings. This work focuses on analyzing the samples using a full scan non targeted acquisition and using high mass resolving power to obtain accurate mass measurements.

# Methods

## Sample Preparation

A total of four O-ring samples were included in the leachable study; A - Red Ring, B - Brown Ring, C - White Ring and D - Black Ring.

An accelerated leachable study was performed, following BPOG guidelines. Samples were cut into 20 mm sections and submerged in 10 mL 100% ethanol, 50% ethanol, WFI and 5M NaCl for 30 days at 40°C in a sealed crimped cap vial. A solvent blank was used, for chromatographic comparison, was also treated following the same protocol. An aliquot of each sample extract was transferred to GC vial for analysis.

## Gas Chromatography-Mass Spectrometry

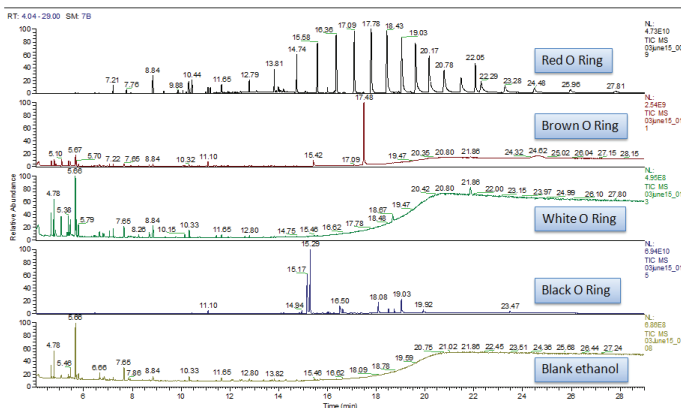
1  $\mu$ L of sample was injected into a splitless injector and compound separation was achieved using a Thermo Scientific™ TRACE™ 1310 gas chromatograph and a Thermo Scientific™ TraceGOLD™ TG-5SILMS 30 m length  $\times$  0.25 mm inner diameter  $\times$  0.25  $\mu$ m film thickness column. A Thermo Scientific™ TriPlus™ RSH autosampler was used for sample introduction (Table 1). High resolution EI spectra were acquired using 60,000 FWHM resolution (measured at  $m/z$  200) with a mass range of 50–600  $m/z$ . An internal lock mass was used throughout the acquisition.

## Data Analysis

Data was acquired and processed using the Thermo Scientific™ TraceFinder™ software. TraceFinder allows easy data reviewing and data reporting.

# Results

Figure 1. GC-MS total ion chromatograms of the 100% ethanol leachate from four O-rings and blank (control).



## Isolating Unique Components

Full scan chromatograms were obtained for each sample and the total ion chromatograms (TICs) are shown in figure 1. The first step in this analysis was to quickly identify the peaks in each of the samples that were unique or significantly elevated when compared to the blank.

This was achieved by binary comparison between the test sample and the blank in TraceFinder. The software first performs an accurate mass deconvolution of the data. An example deconvoluted peak cluster for ethyl octanoate is shown in figure 2. A heat map is shown in TraceFinder to quickly identify the peaks that are elevated in the test sample (figure 3). For example, the peak at 17.49 minutes with a base peak of  $m/z$  277.07800 is a peak elevated in the brown O-ring.

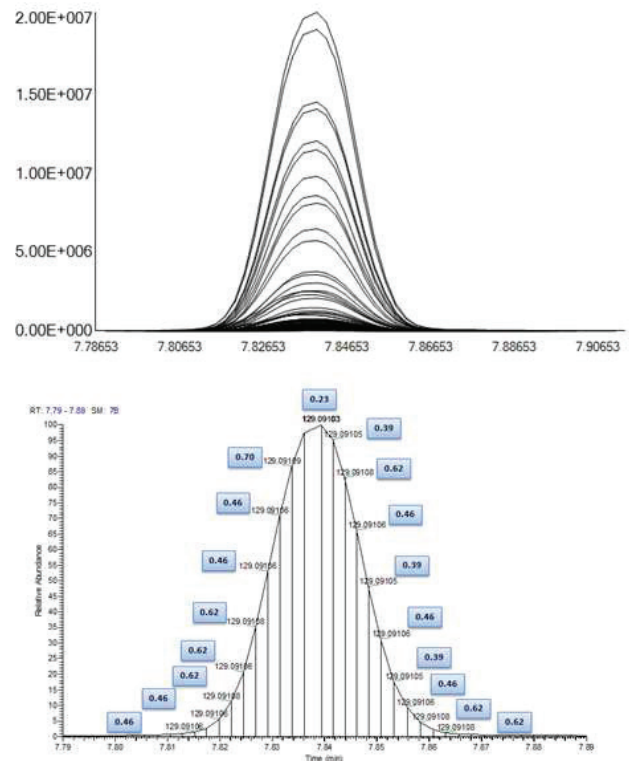
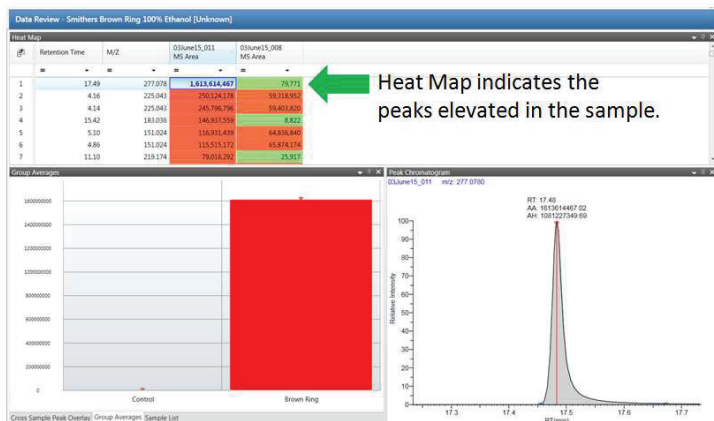


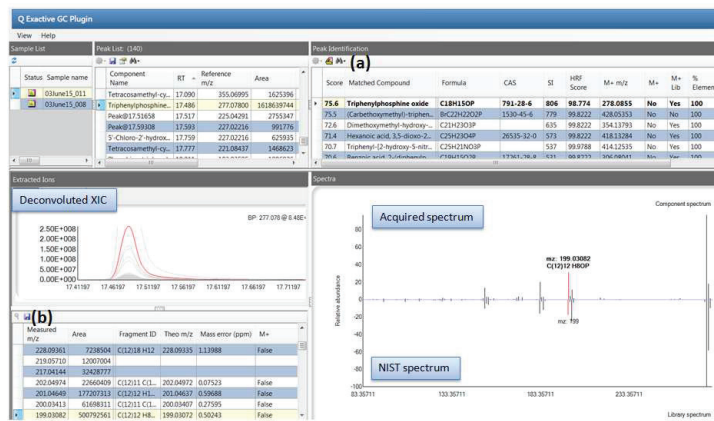
Figure 2. Deconvoluted peak cluster (upper) identified as ethyl octanoate from the black O-ring (D). Extracted ion chromatogram (lower) for ethyl octanoate ion  $m/z$  129.0910 ( $\pm 5$  ppm mass window) in black O-ring showing 18 scans/peak. Excellent accurate mass stability is shown for each individual scan (ppm mass error).



**Figure 3.** TraceFinder unknown screening window showing a section of the peak list for the Brown O-ring (B) and blank (control). The heat map (upper window) is used to isolate the peaks that are elevated in the sample. The group averages window (bottom left) shows the intensity of the peak at 17.49 minutes with base peak of m/z 277.078 in the two samples.

### Identifying compounds with confidence

The deconvoluted spectrum is first searched against commercial nominal mass spectral libraries (e.g. NIST 2014) and the list of hits are scored based on a combination of the search index (SI) score and high resolution filtering (HRF) value (figure 4). The HRF value is the percentage of the spectrum that can be explained by the chemical formula in the library search. The top hit for the peak at 17.49 minutes was for the compound triphenylphosphine oxide, where 98.8% of the spectrum can be explained based on accurate mass. The fragments observed are matched to the elements in the proposed compound with sub 1ppm mass accuracy which adds confidence in the identification.



**Figure 4.** Identification of triphenylphosphine oxide. Screenshot of the deconvoluted data and library match in TraceFinder. (a) List of library hits sorted by score (combination of SI and HRF). (b) List of fragment ions from EI spectrum and elemental composition based on elements in top hit.

Sample	RT (min)	Base Peak (m/z)	Compound Name	Formula	Base Peak Mass Accuracy (ppm)	Molecular Ion Mass Accuracy (ppm)
Black O Ring	15.17	178.07754		C <sub>20</sub> H <sub>20</sub> O <sub>4</sub>	0.9	0.7
	15.29	178.07754		C <sub>20</sub> H <sub>20</sub> O <sub>4</sub>	0.1	0.2
	18.08	171.13806	Tetraethylene glycol bis (2-ethylhexanoate)	C <sub>24</sub> H <sub>46</sub> O <sub>7</sub>	0.6	-
	23.47	219.17435	Irganox 1076	C <sub>33</sub> H <sub>62</sub> O <sub>3</sub>	0.0	1.0
Brown O Ring	14.94	280.10939	ethyl-1-hydroxy-2,3-diphenylcycloprop-2-ene-1-carboxylate	C <sub>18</sub> H <sub>16</sub> O <sub>3</sub>	0.4	0.4
	16.50	126.09145	9-Octadecanamide	C <sub>18</sub> H <sub>35</sub> NO	0.9	0.6
	17.48	277.07790	Triphenylphosphine oxide	C <sub>18</sub> H <sub>15</sub> OP	0.9	0.1
	15.42	183.03595	Triphenylphosphine	C <sub>18</sub> H <sub>15</sub> P	0.7	0.9
	11.10	219.1743	4-tert-butyl-2,6-diisopropylphenol	C <sub>18</sub> H <sub>26</sub> O	0.2	0.2
Red O Ring	11.35	149.02341	Diethyl phthalate	C <sub>12</sub> H <sub>14</sub> O <sub>4</sub>	0.6	0.9
	13.57	185.04198	Diphenylsulfide	C <sub>12</sub> H <sub>10</sub> S	0.2	0.1
	11.93	263.20074	1,4 Dihydrophenacetic acid, 3,5-di-t-butyl, ethyl ester	C <sub>18</sub> H <sub>30</sub> O <sub>2</sub>	0.7	0.4
White O Ring	7.65	101.02344	Butanedioic acid, diethyl ester	C <sub>8</sub> H <sub>14</sub> O <sub>4</sub>	0.5	-
	10.44	163.07549	Ethanone, 1-[4-(1-hydroxy-1-methylethyl)phenyl]	C <sub>11</sub> H <sub>14</sub> O <sub>2</sub>	0.9	0.4
	15.09	87.044	Methyl stearate	C <sub>19</sub> H <sub>38</sub> O <sub>2</sub>	1.3	0.1
16.00	155.07025	di(butoxyethyl)adipate	C <sub>18</sub> H <sub>34</sub> O <sub>6</sub>	0.1	1.0	

**Table 1** Summary of the peaks elevated in the four O ring samples and the tentative identification of the compounds

## Conclusion

GC Orbitrap mass spectrometer in combination with TraceFinder is an extremely effective tool for the profiling of extractables samples and in the identification of unknown peaks.

- Reliable and robust chromatographic separation in combination with fast data acquisition speeds make the Q Exactive GC an ideal platform for chemical profiling of complex samples.
- The consistent sub 1 ppm mass accuracy in combination with excellent sensitivity makes confident identification of all components in a sample possible. Routine resolving power of 60,000 FWHM and a wide dynamic range eliminates isobaric interferences, increasing confidence in results when compounds are identified in complex matrices.
- The EI and PCI data obtained was used for tentative compound identification against commercial libraries. Where no library match was made the mass accuracy allowed for elemental compositions to be proposed with a high degree of confidence. Proposed identifications can be quickly confirmed or eliminated based on accurate mass.

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