Effective Workflow for Pharmaceutical API Impurity Analysis using HR- LCMS and Compound Discoverer

Kate Comstock1; Caroline Ding1; Vincent Jespers2

¹Thermo Fisher Scientific, San Jose, CA, USA; ²Thermo Fisher Scientific, Erembodegem, Belgium

Overview

Purpose: Demonstrate an effective workflow for pharmaceutical impurity identification using Thermo Scientific™ Orbitrap Elite™ mass spectrometer and novel node-based small molecule structure ID software Thermo Scientific ™ Compound Discoverer™

Methods: LC-HRMS and Compound Discoverer software for Fexofenadine API impurity analysis.

Results: The Fexofenadine API impurity profile was quickly obtained.

Introduction

Pharmaceutical impurity analysis is crucial for drug R&D, production, and postmarketing surveillance. LCMS is routinely used for impurity analysis because of its speed and sensitivity. For rapid, accurate, and confident impurity ID, very high resolution mass spectrometer and effective data processing software are essential.

This study demonstrates an effective workflow for pharmaceutical impurity identification using very high resolution mass spectrometer and node-based small molecule structure ID software: Compound Discoverer software.

Methods

Sample Preparation

The commercial compound Fexofenadine (Sigma-Aldrich F9427-10MG, cas# 83799-24-0) was dissolved in 1:1 ACN/Water at a concentration of 0.3 µg/mL.

Liquid Chromatography

HPLC system: Thermo Scientific™ Accela™ 1250 pump, Open Accela Autosampler and PDA

Column: Thermo Scientific™Accucore™ C18 2.1x 150 columns, 2.6 µm. Injection

volume: 5 µl

Mobile phases: A - H₂O B - Acetonitrile

C - $\rm H_2O$ with 0.05% Ammonium Hydroxide pH 9

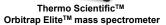
Time (min.) A% Gradient: B% C% ul/min 60 15 25 400 0.5 400 60 15 25 14.0 25 50 25 400 19.0 5.0 70 25 400 60 15 400 19.1

Mass Spectrometry

The high resolution accurate mass (HRAM) analysis was conducted on an Orbitrap Elite mass spectrometer equipped with a HESI II ion source. Full scan MS and top3 data-dependent MS/MS data were collected at resolutions of 120,000 and 15,000 respectively.

Ionization mode: ESI positive Scan range: 160-1500 amu Sheath gas flow rate (N2): 45 Auxiliary gas flow rate (\tilde{N}_2) : 10 Spray voltage (KV): +4.0 for positive Capillary temp (°C): 300 S-lens RF level: 60.0

Heater temp (°C): 450





Data Processing

Node Based Processing Workflow in Compound Discoverer

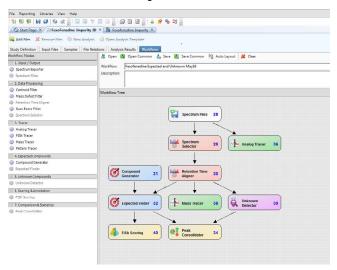
The HRAM full scan and HCD ms/ms data acquired on an Orbitrap Elite MS was processed using Compound Discoverer (CD) software for Fexofenadine API impurity profiling.



Compound Discoverer (CD) software provides flexible processing workflows which are assembled from a suite of advanced algorithms (nodes). The drag-and-drop workflow editor allows greater control and visibility in terms of how data should be processed.

Most API impurities are structurally related to the API, but unrelated unknowns do occur. In this study, the CD processing workflow included the following nodes to ensure complete impurity identification: Using "Expected Finder" to get an expected ions list from "Compound Generator" node and detect expected compounds. Using "FISh Scoring" node for fragment ion matching and fragment structure annotations on spectra. "Unknown Detector" node was added to detect structurally unrelated impurities. "Peak Consolidator" node grouped the peaks detected from both expected and unknown mechanisms for quick comparison and more confident identification. See Figure 1.

FIGURE 1. Node Based Processing Workflow



Results

FIGURE 2. Base Peak Chromatogram of Fexofenadine in CD "Specialized Traces"

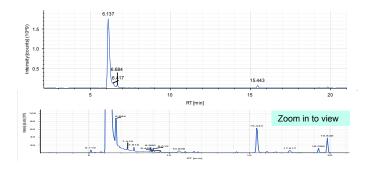


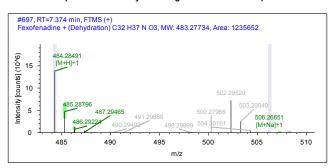
FIGURE 3. Results Review



Structure Characterization for Expected Compound Hits

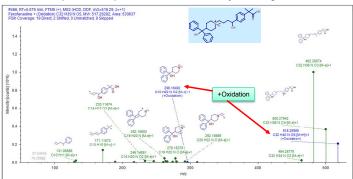
The detailed and comprehensive processing results are shown in Figure 3. It includes "Expected Compound Hits" and "Unknown Compound Hits" from Expected Finder node and Unknown Detector node respectively. An example of fine isotopic pattern confirmation of elemental formula assignment for "Expected Compound Hits" is shown in Figure 4. Color coding of isotopic fidelity gave greater confidence in elemental composition assignment from CD. Automatic adduct grouping reduced false positive hits.

FIGURE 4. Isotope Pattern Fidelity for Assigned Elemental Composition.



For each expected impurity hit, FISh Scoring automatically searched the fragmentation spectra, and annotated matching fragment structures directly on the spectra. The annotations are color-coded to visually indicate the transformation shifted ones for transformation localization, see Figure 5.

FIGURE 5. Expected Compound Hit with Automatically FISh Fragment Annotations



Structure Characterization for Unknown Compound Hits

For unknown compound hits, "Mass Spectrum View" showed the HRAM mass and corresponding ms/ms spectrum. The interested unknown compound s were added to a custom explanation table. Based on the HRAM fragmentation data, putative structures were propose in "Custom Explanation Editor" (Figure 7), followed by "FISh Scoring" on the fly, the unknown component ms/ms spectra were automatically annotated with matching fragment structures (Figure 6). "FISh Coverage" score indicated the percentage of fragment ion matching between experimental data and theoretical predictions from Mass Frontier™ Fragmentation Libraries™.

FIGURE 6. Unknown Compound Structure Elucidation

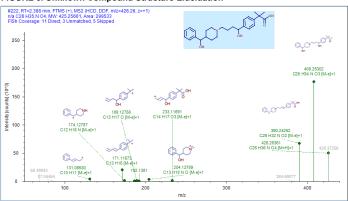


FIGURE 7. Custom Explanations Editor



The versatile and flexible "Result Filters" was used for quick data manipulation by selecting the criteria and options.

FIGURE 8. Result Filters



Data Reporting

The result was reported in the Expected and Custom explanation formats. For each identified impurity, it's isotope pattern, annotated ms/ms spectrum, transformation, Fish coverage, spectral distance, and others were included in the report, see Figure 9.

FIGURE 9. Reports of Fexofenadine Impurity: Expected and Custom Explanations



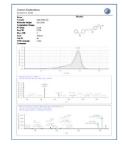
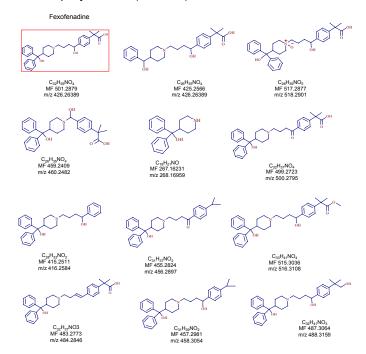


TABLE 1 Expected Compound Hits without Graphs (partial list)

| Expected Compound Hits | | | | | | | | | | | | 90 |
|------------------------|--------------|-----------|--|---------------|---------------------|-----------|----------|---------|------|-----------|----------|------|
| Parent Compand | | | Transformations | Composition | AMass (pom) per lon | WY Inches | | Rest SD | | Агна | Commund | Få |
| Parent Compound | Formula | Molecular | Transformations | Change | AMuss (ppm) per lon | KI (mn) | Coverage | Best SU | E MI | Area | Area [3] | Pile |
| Excleration | C32 H35 N D4 | 501.28791 | | | 0.77 (Metget | 6.000 | 94.44 | 0.256 | 3 | 406031450 | 94,119 | |
| Fextferadine | C32 H37 N D4 | 499.27226 | Dehydration | -(H2) | 0.11 (M+H)+1 | 6.649 | 80.00 | 0.208 | 2 | 8865095 | 2.051 | |
| | | | | | 1.53 (M+166)+1 | | | | | | | |
| Fexiferadire | | 499.27226 | Dehydration, Oxidation | (H2) | 0.11 (M+H)+1 | 6.649 | 90.00 | 0.208 | 2 | 8865095 | 2.051 | |
| | | | | | 1.53 (M=Na)=1 | | | | | | | |
| Festileradine | C32 HG7 N G3 | 483.27734 | Dehydration | -(H2 O) | -0.10 (M+H0+1 | 6,078 | 86.21 | 0.140 | 3 | 6927294 | 1.605 | |
| Fexoferadine | C32 H39 N D4 | 501.28791 | | | 0.87 [M=Na]+1 | 6.128 | 33.33 | 0.104 | 3 | 2102793 | 0.486 | |
| Fextferadine | C32 H37 N O3 | 483,27734 | Dehydration | -(H2 O) | -0.60 (M+H0+1 | 7.763 | 97.27 | 0.084 | 3 | 1383870 | 0.322 | |
| Fexefenadine | C18 H19 N | 249.15175 | Dehydration | (C14 H20 O4) | Q 61 (MHPQ+1 | 10,582 | 77.78 | 0.125 | 2 | 1309300 | 0.303 | |
| Feederadine | C32 H37 N O3 | 483.27734 | Dehydration | (H2 O) | -0.60 (M+H)+1 | 7.366 | 72.37 | 0.237 | 3 | 1235652 | 0.286 | |
| | | | | | Q.11 [M+Na]+1 | | | | | | | |
| Fextferadine | C18 HQ1 N O | 267.16231 | | -(C14 H18 O3) | 0.70 (M+H)-1 | 10.500 | 61.02 | 0.154 | 2 | 1221143 | 0.293 | |
| Feroferadine | C32 H35 N O2 | 465,26678 | Dehydration, Dehydration | -(H4 O2) | -0.38 (M+H)+1 | 6.064 | 92.31 | 0.103 | 3 | 1007980 | 0.233 | |
| Fexuleradine | C32 H39 N O5 | 517.28282 | Denyarasun | -(0) | 0.36 (M+H0+1 | 5.074 | 95.12 | 0.156 | 3 | 530637 | 0.123 | |
| | | | | | 0.47 (M+Ne)+1 | | | | | | | |
| Feroferadine | C32 H39 N O5 | 517.28282 | Oxidation | +(0) | 0.35 (M+H0+1 | 5.074 | 97.56 | 0.156 | 3 | 530637 | 0.123 | |
| | | | | | Q.47 (M+Na)+1 | | | | | | | |
| Fexofenadine | C32 H39 N O5 | 517.28282 | | +(0) | 0.00 (M+HE+1 | 2.662 | 87.16 | 0.111 | 3 | 156288 | 0.000 | |
| Feederadine | C32 H39 N O5 | 517.28282 | Oxidation | +(0) | 0.00 (MHPQ+1 | 2.682 | 87.18 | 0.111 | 3 | 156268 | 0.036 | |
| Festignadine | C32 H39 N O5 | 517.20202 | | +(0) | 0.00 (M+H)+1 | 0.578 | 93.75 | 0.103 | 2 | 113600 | 0.006 | |
| Fexteradine | C32 H39 N O5 | 517.28282 | Oxidation | +(0) | 0.00 (M+H)+1 | 0.578 | 50.63 | 0.103 | 3 | 113600 | 0.026 | |
| Fecoferadine | C32 H39 N D5 | 517.20202 | Oxidation | -(0) | 1.65 (M+H)-1 | 5,540 | 00.06 | 0.234 | 3 | 102966 | 0.024 | |
| Fexofenadirie | C32 H39 N O5 | 517.28282 | | +(0) | 1.65 (M+H)+1 | 5.546 | 86.05 | 0.234 | 3 | 102966 | 0.024 | |
| Fexefenadine | C32 H39 N O5 | 517.28282 | Oxidation | +(0) | 0.00 (M+H)+1 | 4 225 | 55.74 | 0.132 | 3 | 56538 | 0.013 | |
| Festionadine | C32 H39 N O5 | 517.28282 | | +(0) | 0.00 (M+Hp+1 | 4.225 | 95.74 | 0.132 | 3 | 56538 | 0.013 | |
| Fexclenatine | C32 H35 N O3 | 481.26169 | Dehydration, Dehydration | -(H4 D) | 42.78 [Merget | 6.645 | | 0.254 | 3 | 54751 | 0.013 | |
| Fexoferadine | C32H35N 03 | 481,26169 | Dehydration, Dehydration, Oxydeton | -(H4 O) | -0.70 (M++Q+1 | 6.645 | | 0.254 | 3 | 54751 | 0.013 | |

TABLE 2. Impurity Structures (Partial List)



Conclusion

- •Effective and confident impurity analysis was achieved using very high resolution LCMS from the Orbitrap Elite mass spectrometer and Compound Discoverer software.
- ■Powerful workflow options in Compound Discoverer software detect components with targeted and untargeted mechanisms, and utilize very high resolution to quickly perform fine isotope searches. The determination of the structures of impurities is simplified with automatic FISh (fragment ion search) annotations.

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