

# The Use of Hydrophilic Interaction Chromatography with Mass Spectrometry Detection for the Separation of Basic Pharmaceuticals and Food Toxins

R.E. Majors\*, Agilent Technologies, Wilmington, DE, USA;

T. Yoshida and H. Kumagai, Agilent Technologies,  
Hachioji-Shi, Tokyo, Japan



# Outline of Talk

- Problems with polar compounds and RPLC
- What is HILIC?
- What types of samples can it be used for?
- What types of columns and mobile phases can be used?
- Advantages & Disadvantages of the Technique
- Comparison of RPC and HILIC
- Some applications using LC-MS/MS
- Conclusions

# Polar Compounds and Reversed-Phase HPLC

- 92% of all users employ RPLC in their laboratory at one time or another\*
- Separation based on hydrophobicity of analytes
- Polar compounds tend to elute early; the more polar, the earlier the elution
- For very polar compounds, in order to increase retention, the amount of organic modifier in mobile phase is reduced
- - When % organic becomes very low (say <5%), phase collapse (solvent dewetting) may occur
  - Quantitation is impaired for analytes with  $k' < 1$
- There are special RPC phases such as polar-embedded, hydrophilic endcapped, wide pore/lower density bonded phases, & short chain phases to increase retention at high aqueous but there may be a decrease in MS sensitivity due to poor mobile phase desolvation and ion suppression

\* R.E. Majors, LCGC Survey, 2007

# Another Way to Increase Retention for Polar, Ionizable Compounds is to Use Ion Pair RPC

- Adjust pH to ionize analyte and add to the mobile phase an alkyl counterion (e.g. sulfonate for cations or tetrabutylammonium for anions) to form ion pair which increased retention
  - Ion pair reagents modify columns
  - Equilibration time is slow, esp. for long alkyl chain IP reagents
  - MS systems don't like typical ion pair reagents typically used with UV detection



# A Third Way to Increase Retention is to Derivatize the Analytes to Provide Increased Retention

- Requires an additional chemical synthesis step; a hassle
- Derivatization is not always quantitative
- Sometimes extraneous peaks are introduced



# Could Use Normal Phase HPLC

- Must carefully control water content of mobile phase (especially with silica)
- Many chromatographers aren't familiar with the use of NP HPLC
- Some of the normal bonded phase columns aren't very stable
- Water-immiscible organic solvents are frequently used (e.g. hexane, dichloromethane)



# ...Or Use Hydrophilic Interaction Liquid Chromatography (HILIC)

- What is HILIC?
  - A liquid chromatographic technique that has been around for a couple of decades (Andy Alpert's early work on sugars on amino columns using H<sub>2</sub>O/ACN)
  - Uses a polar stationary phase such as silica, amino, mixed mode, zwitterionic, etc.
  - Uses a water-miscible, non-polar mobile phase containing a small amount of water (at least 2.5% by weight)
  - May use volatile buffers for compatibility to MS detection

# Characteristics of HILIC

- **Hydrophilic, polar and charged compounds are retained preferentially compared to hydrophobic, neutral compounds---opposite of RPLC.**  
e.g. toluene is unretained while uracil is retained
- **Addition of water to mobile phase reduces retention**
- **Provides good peak shape for strongly polar solutes compared to normal phase**
- **Complements RPLC; retains hydrophilic compounds and often reversed elution order**
- **Sometimes called “aqueous normal phase (ANP)”**





# Parameters That May Need Optimization

- Type of stationary phase
- Organic solvent concentration
- Type of buffer
- Buffer (salt) concentration
- pH
- Temperature



# Advantages of HILIC

- Good peak shape for basic compounds where RPC may give tailing and/or low efficiency
- Higher flow rates and/or long columns can be used due to low viscosity mobile phases with high organic content; greater efficiency
- Enhanced detection sensitivity with MS
  - Efficient spraying and desolvation in electrospray MS
  - As much as 3X sensitivity
- Can directly inject extracts from C18 SPE cartridges or acetonitrile precipitated plasma supernatant
- Orthogonal to RPC (2D separations); elution order reversal



# Disadvantages of HILIC

- Mechanism not entirely understood; mixed mechanisms; studies are being done here
- Column overloading can be a problem
  - Similar to silanol overloading in RPC
  - Overloaded HILIC peaks show fronting
- Equilibration times can be long for certain column types



# Typical Columns Used in HILIC

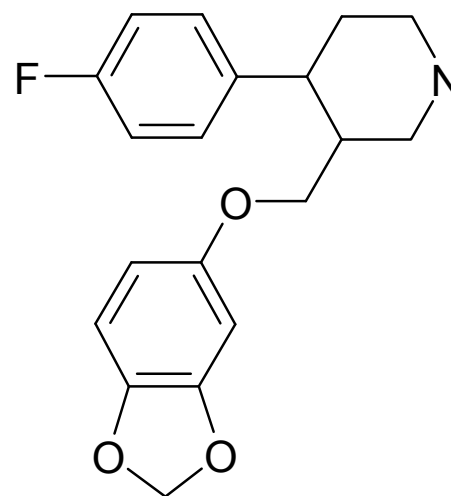
## (Silica- and Polymer-Based)

- Silica
- Amino
- Mixed-mode
  - Alkyl-diol
  - Alkyl-carboxyl
  - Aromatic-cyano
  - C18-SCX
  - C18-amide
  - Phenyl-amino
  - Alkyl hydroxyl
  - Polyhydroxyethylaspartamide
- Hydride and modified hydride
- Zwitterionic (e.g. tetraalkylamine-sulfonic)
- Proprietary phases

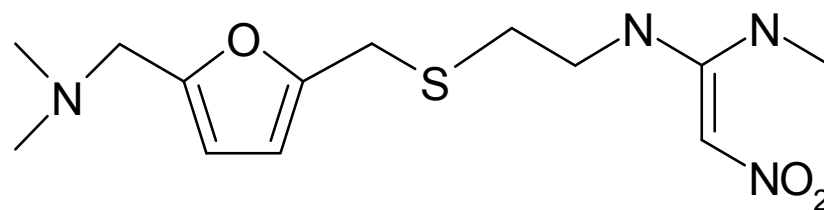


# Structures of Drug Compounds Studied

**a) Paroxetine**  
**Antidepressant**  
**MW 329.36**



**b) Ranitidine**  
**Antiulcerative**  
**MW 314.41**



# Chromatographic Conditions

## RPLC

- Instrument: Agilent Series 1100LC
- Column: ZORBAX Eclipse XDB-C18, 2.1 mm × 150 mm, 5 μm)
- Mobile phase: A: 8-mM HCOONH<sub>4</sub> in water  
B: 8-mM HCOONH<sub>4</sub> in 95% acetonitrile (ACN)/5% water
- Gradient: 5% B to 90% B in 10 min
- Column temperature: 40 °C
- Sample volume: 5 μL
- Flow rate: 0.3 mL/min

## HILIC

- Everything same as for RPLC except for column and gradient conditions:
- Column: ZORBAX RX-SIL, 2.1 mm × 150 mm, 5 μm
- Gradient: 100% B to 50% B in 10 min



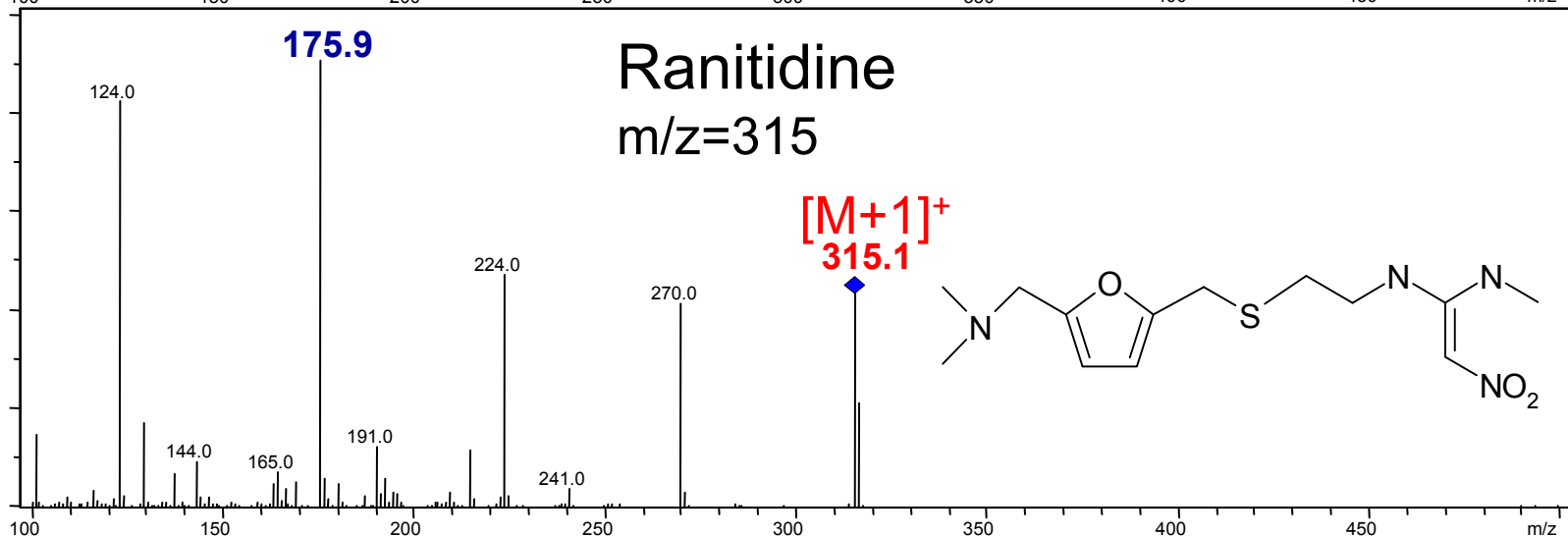
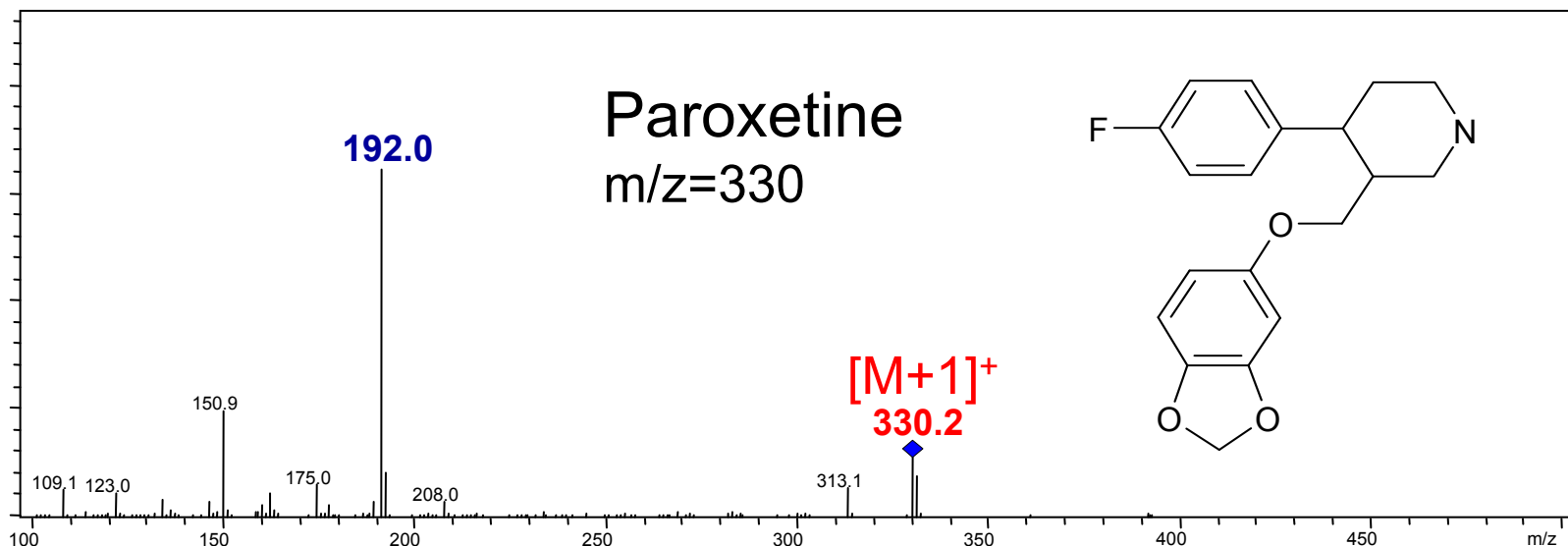
# Mass Spec Conditions

- Instrument: Series 1100 LC/MSD Trap
- Ionization: Positive ESI
- Scan range: 100–500  $m/z$
- SIM ions:  $m/z = 315, 330$
- Drying gas: 10 L/min at 350 °C
- Nebulizer gas: 45 psi
- Fragmentor voltage: 0.25 V



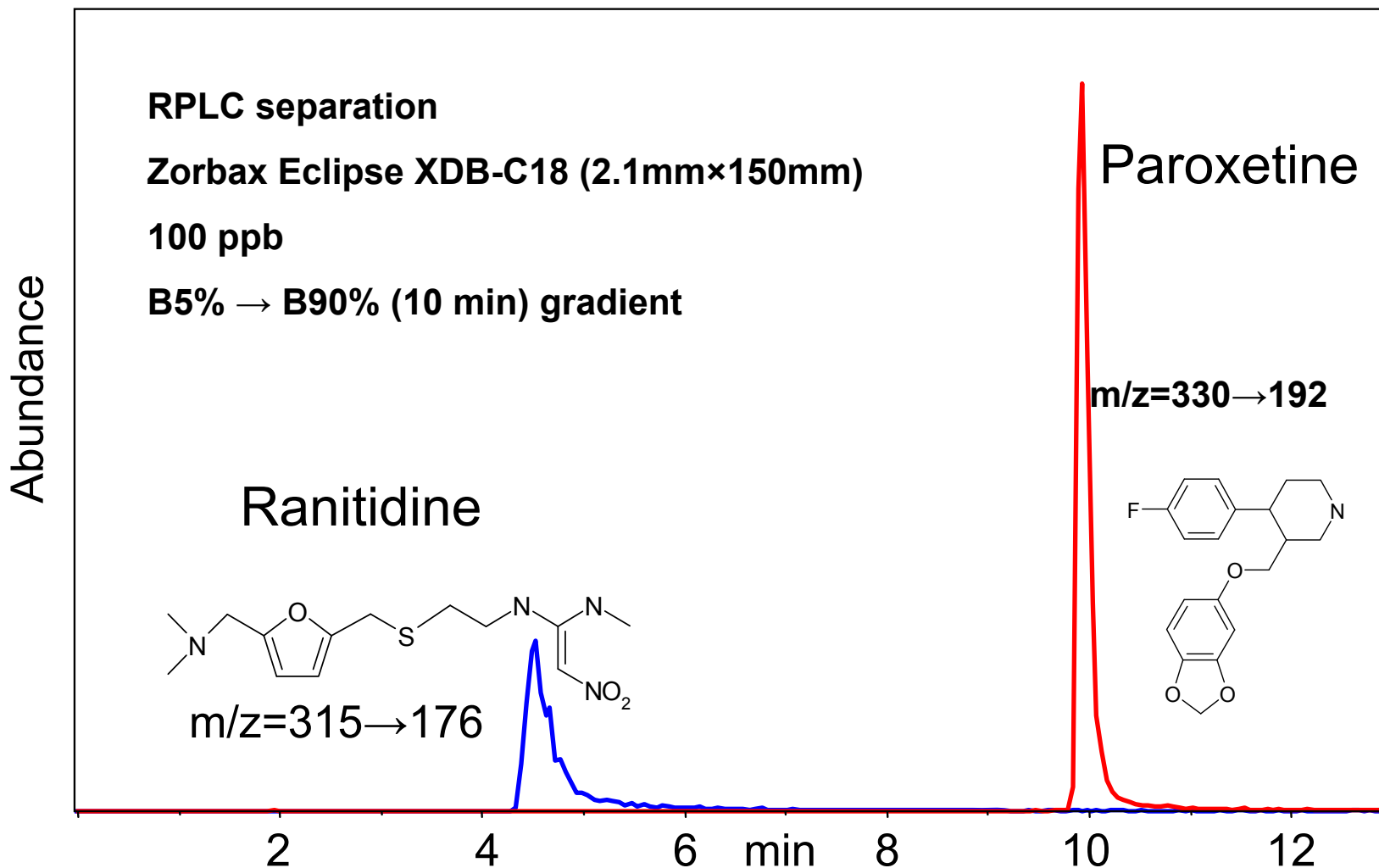
# MS/MS Spectra of Drug Standards

□ □ precursor ion selected for subsequent fragmentation

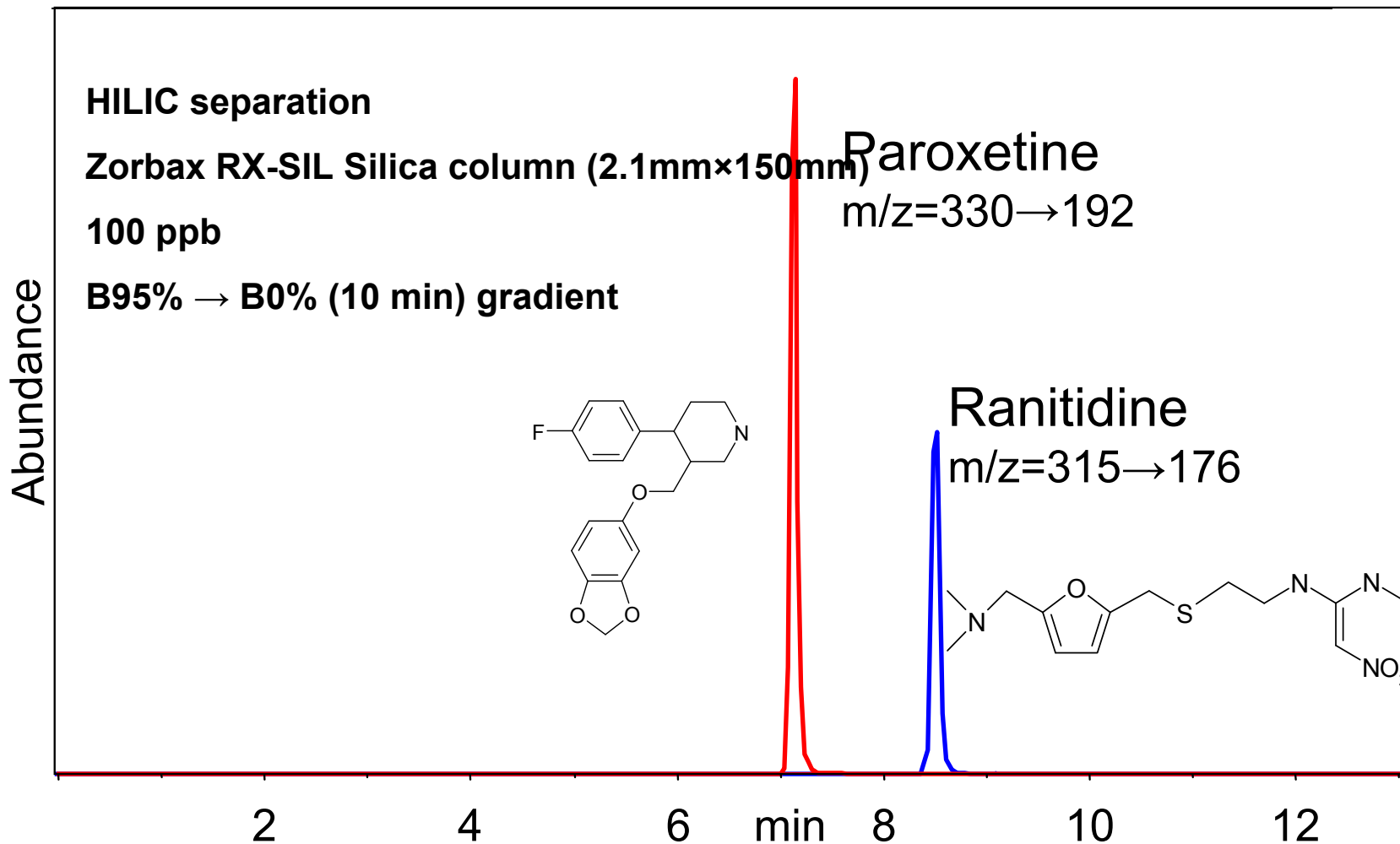




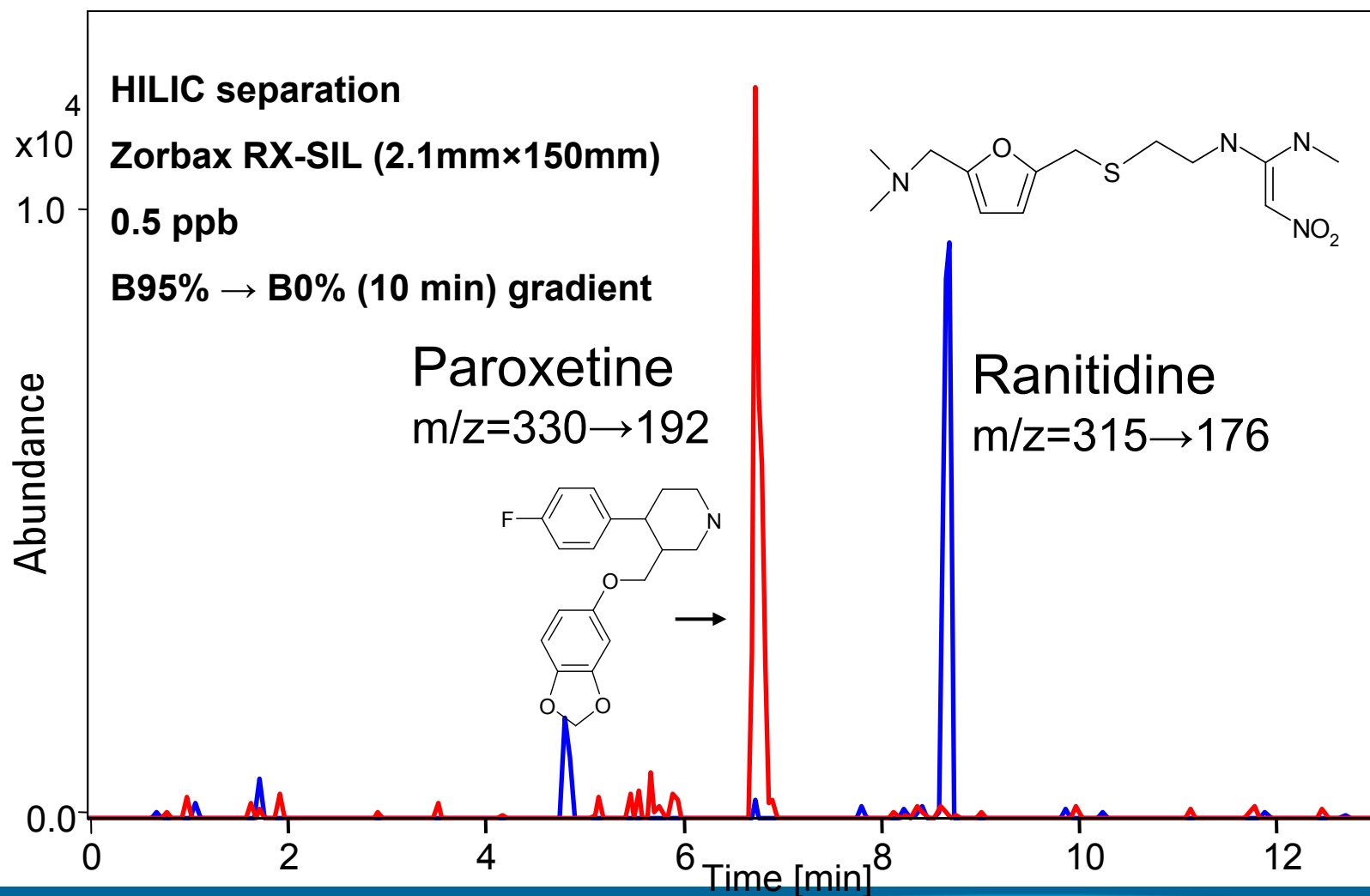
# LC/MS/MS Separation of Paroxetine and Ranitidine on Zorbax Eclipse XDB-C18 Column (RPLC Mode)



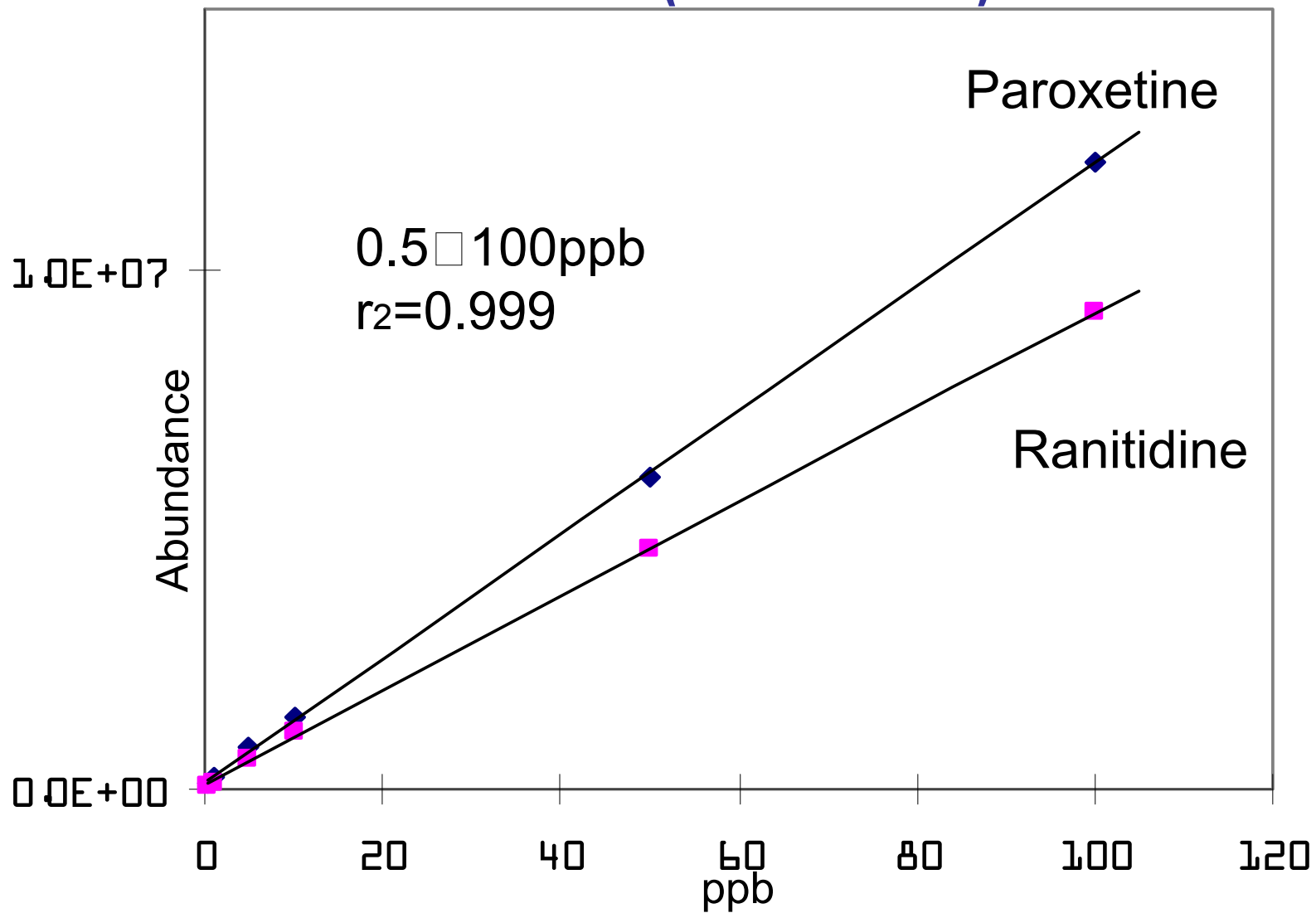
# LC/MS/MS Separation of Paroxetine and Ranitidine on Zorbax RX-SIL Column (HILIC Mode)- 100 ppb Level



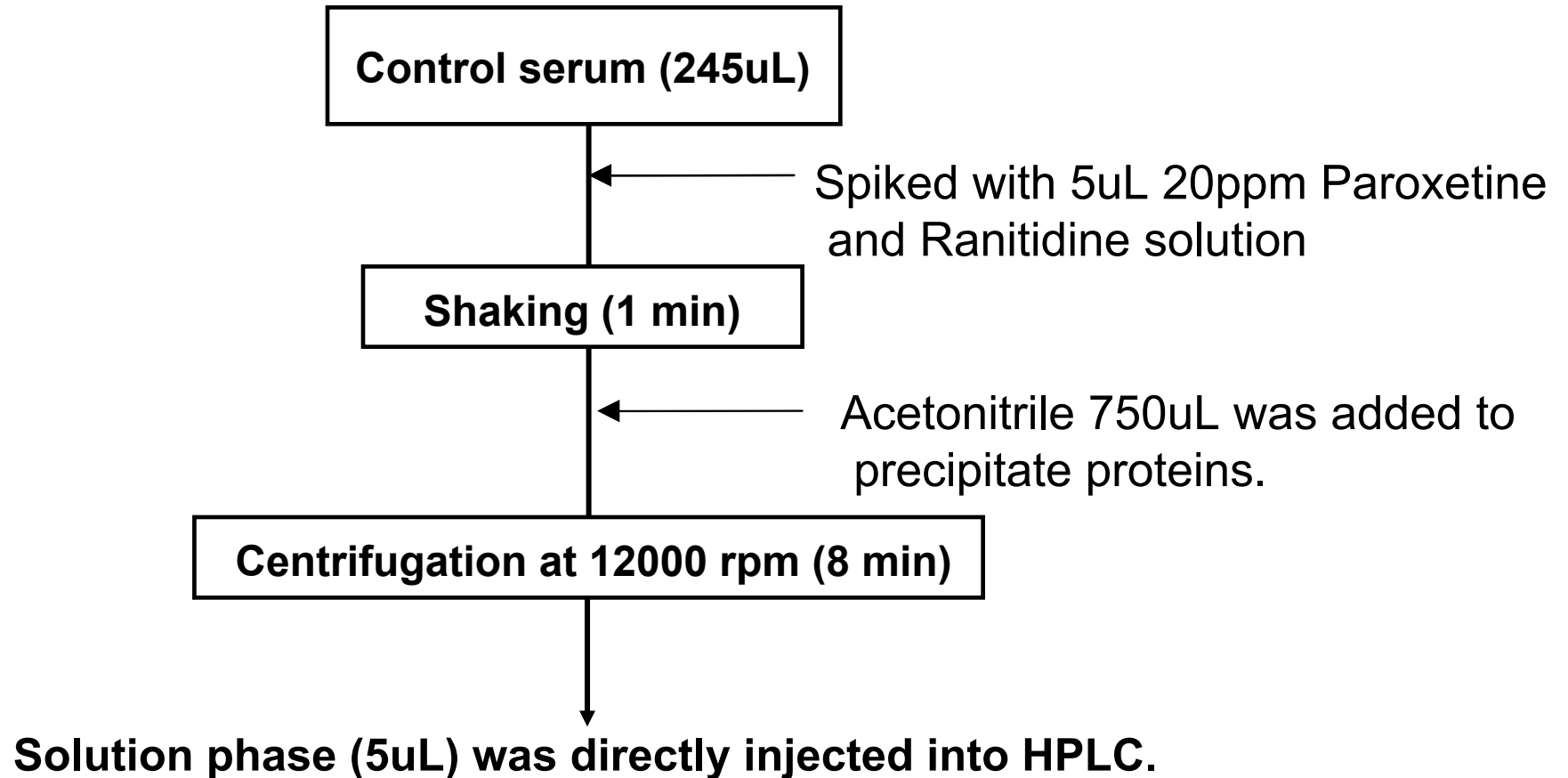
# LC/MS/MS Separation of Paroxetine and Ranitidine on Zorbax RX-SIL Column (HILIC Mode)- 0.5 ppb Level



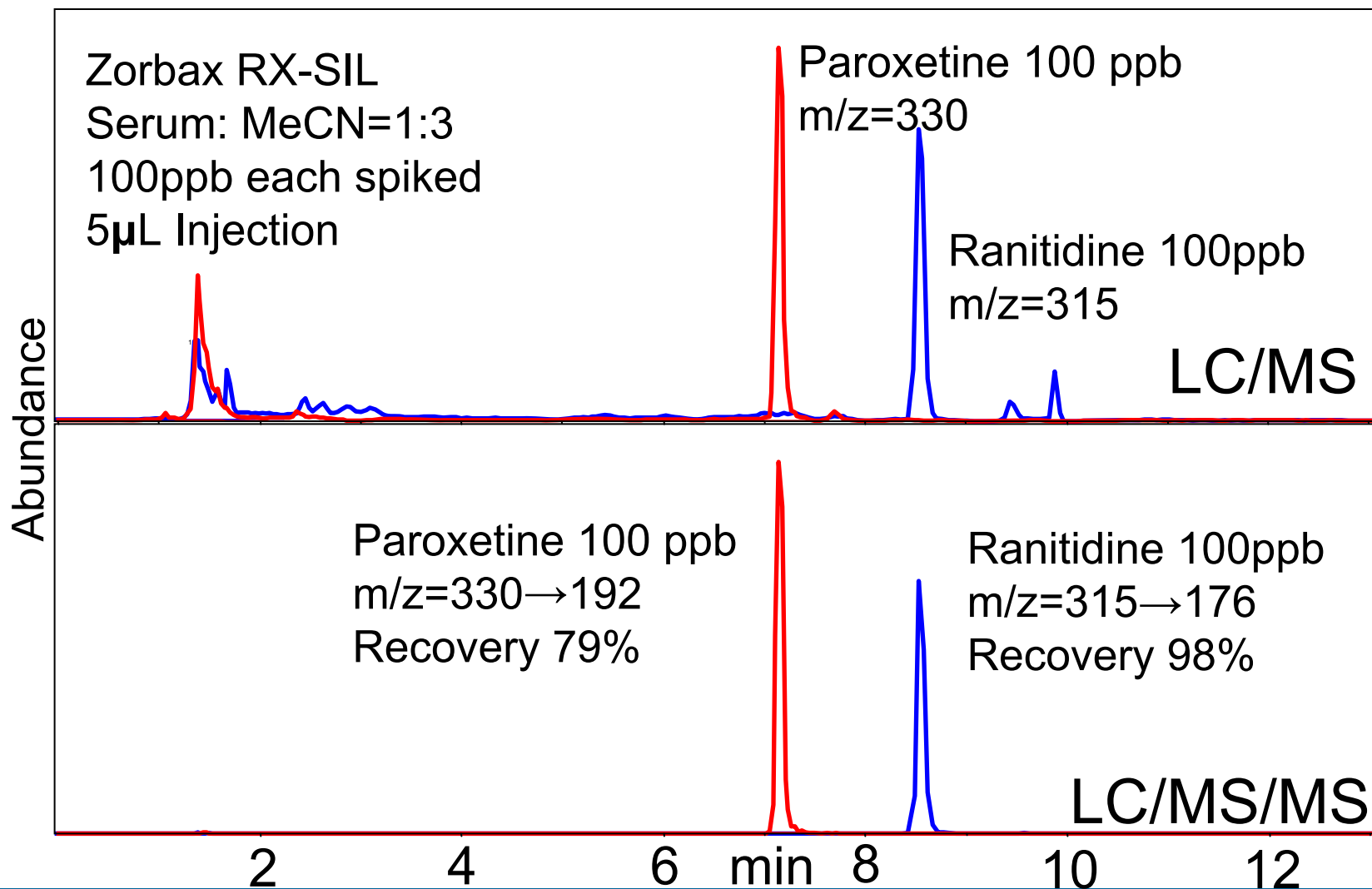
# Linearity of Paroxetine and Ranitidine on Zorbax RX-SIL Column (HILIC Mode)



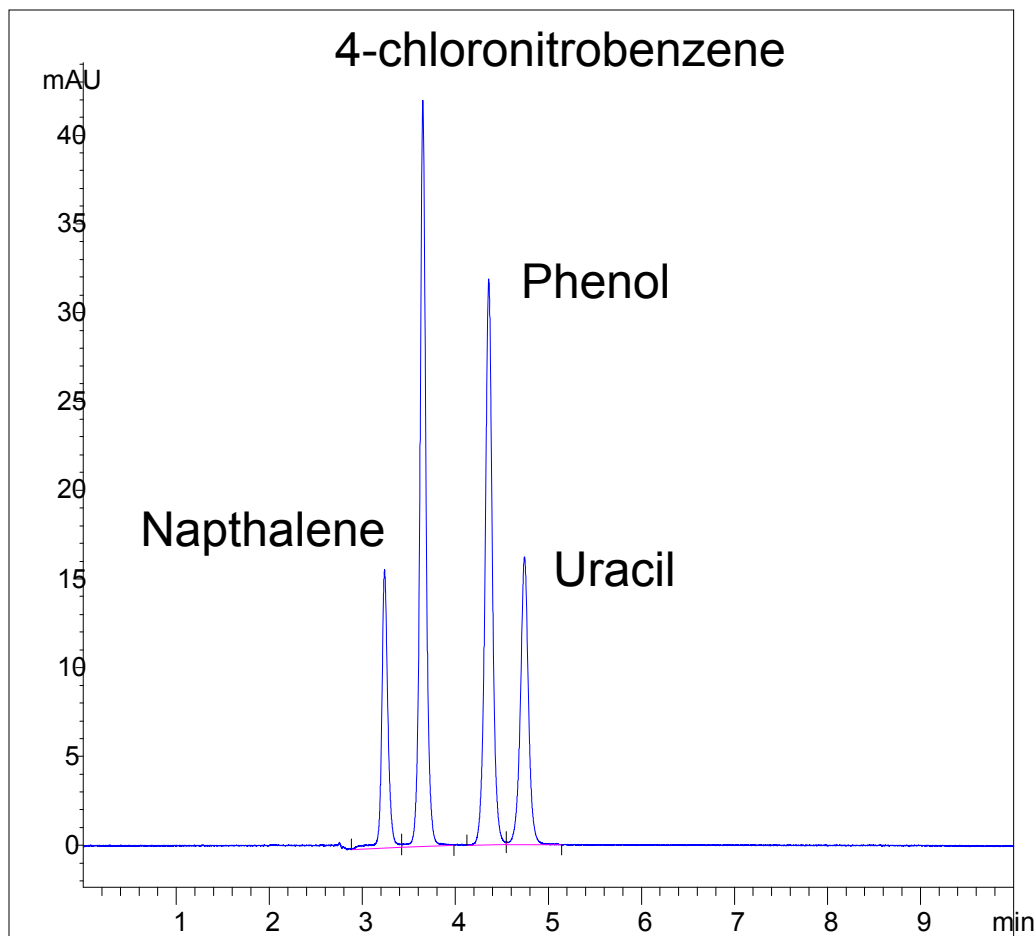
# Sample Preparation Procedure of Spiked Serum



# LC/MS and LC/MS/MS Analysis of Paroxetine and Ranitidine Spiked into Human Serum with Zorbax RX-SIL Column

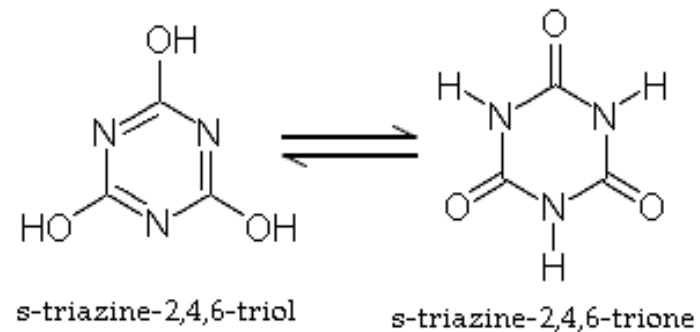
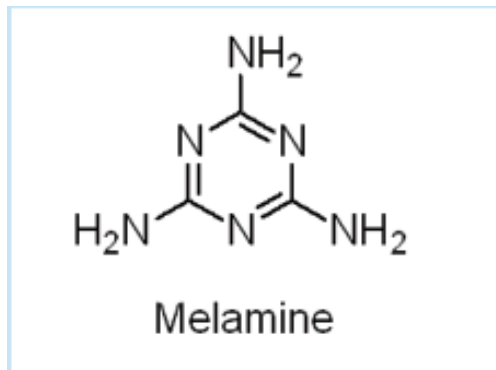


# Typical Elution Order for Test Compounds on HILIC Column



Column: Experimental Modified  
Hydride, 100 Å, 4.6 x 150 mm;  
4 µm  
Mobile phase: 75% ACN / 25% H<sub>2</sub>O  
Flow Rate = 0.5 ml/ml  
Temp = 25 °C  
Injection Volume = 5 µl

# Melamine and Cyanuric Acid Analysis in Pet Food

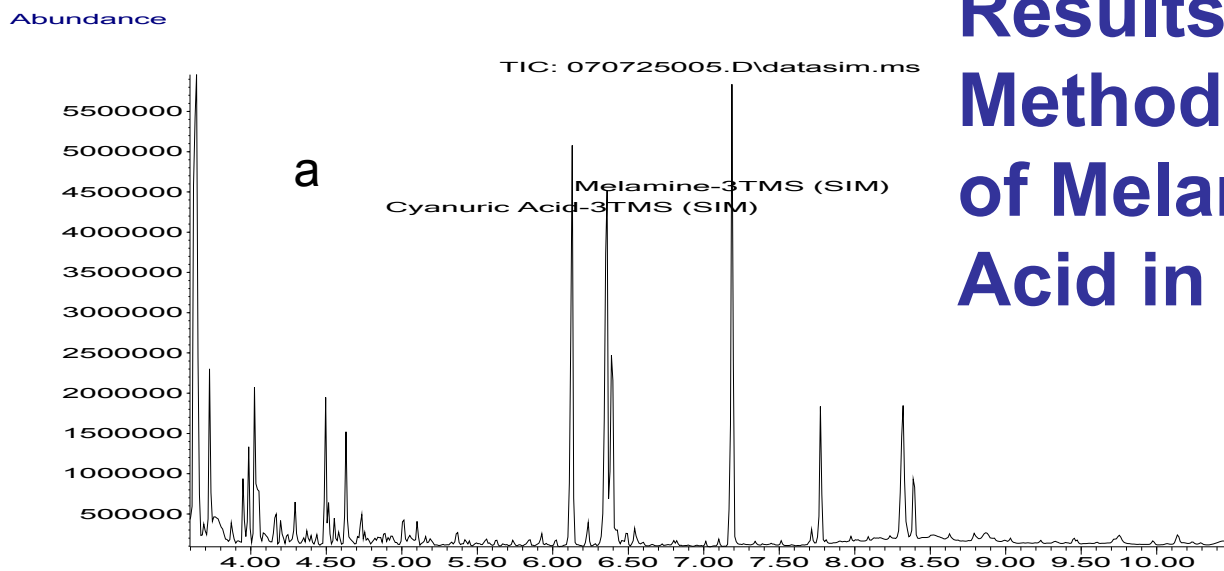


Cyanuric Acid

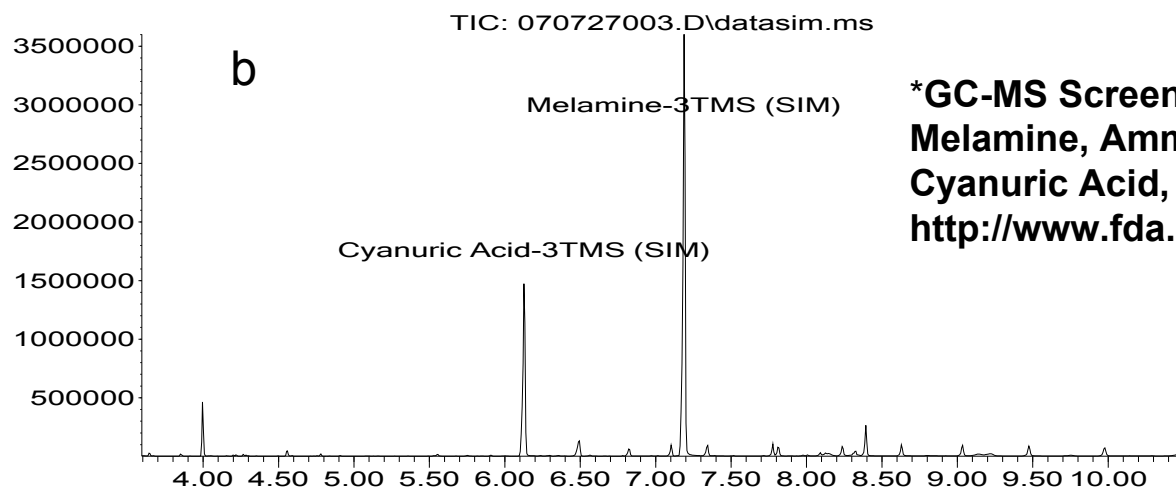
- Combination of these two resulted in renal failure of cats/dogs
- FDA published GC/MS method using silylation; 10-ug/g detection limit
  - Confirmation using SIM/SCAN



# Results from FDA GC/MS Method\* for the Analysis of Melamine and Cyanuric Acid in Pet Food



Abundance



\*GC-MS Screen for the Presence of Melamine, Ammeline, Ammelide, & Cyanuric Acid, Version 2, May 7, 2007  
<http://www.fda.gov/cvm/melaminepresence.htm>

The influence of the selection of monitoring ions in SIM mode shown with spiked dry cat food; a) the ions of highest abundance were used (cyanuric acid: m/z 73, 147, 171; melamine: m/z 327, 330, 342, 345) b) characteristic ions were used (cyanuric acid: m/z 188, 330, 345; melamine: m/z 197, 285, 327, 342)

# HPLC Parameters

|                         |  |
|-------------------------|--|
| <b>HPLC system</b>      | <b>: Agilent 1200 RRLC</b>                       |
| <b>Column</b>           | <b>: Agilent Zorbax-Rx Sil, 2.1×150 mm, 5-um</b> |
| <b>Injection Volume</b> | <b>: 10 uL</b>                                   |
| <b>Temp</b>             | <b>: 40 °C</b>                                   |
| <b>Flow rate</b>        | <b>: 0.2 mL/min</b>                              |
| <b>Mobile phase</b>     | <b>: A - 5 mM Ammonium acetate in Water</b>      |
|                         | <b>: B - 5 mM Ammonium acetate in ACN</b>        |
| <b>Isocratic</b>        | <b>: 95%B</b>                                    |



# MS Parameters



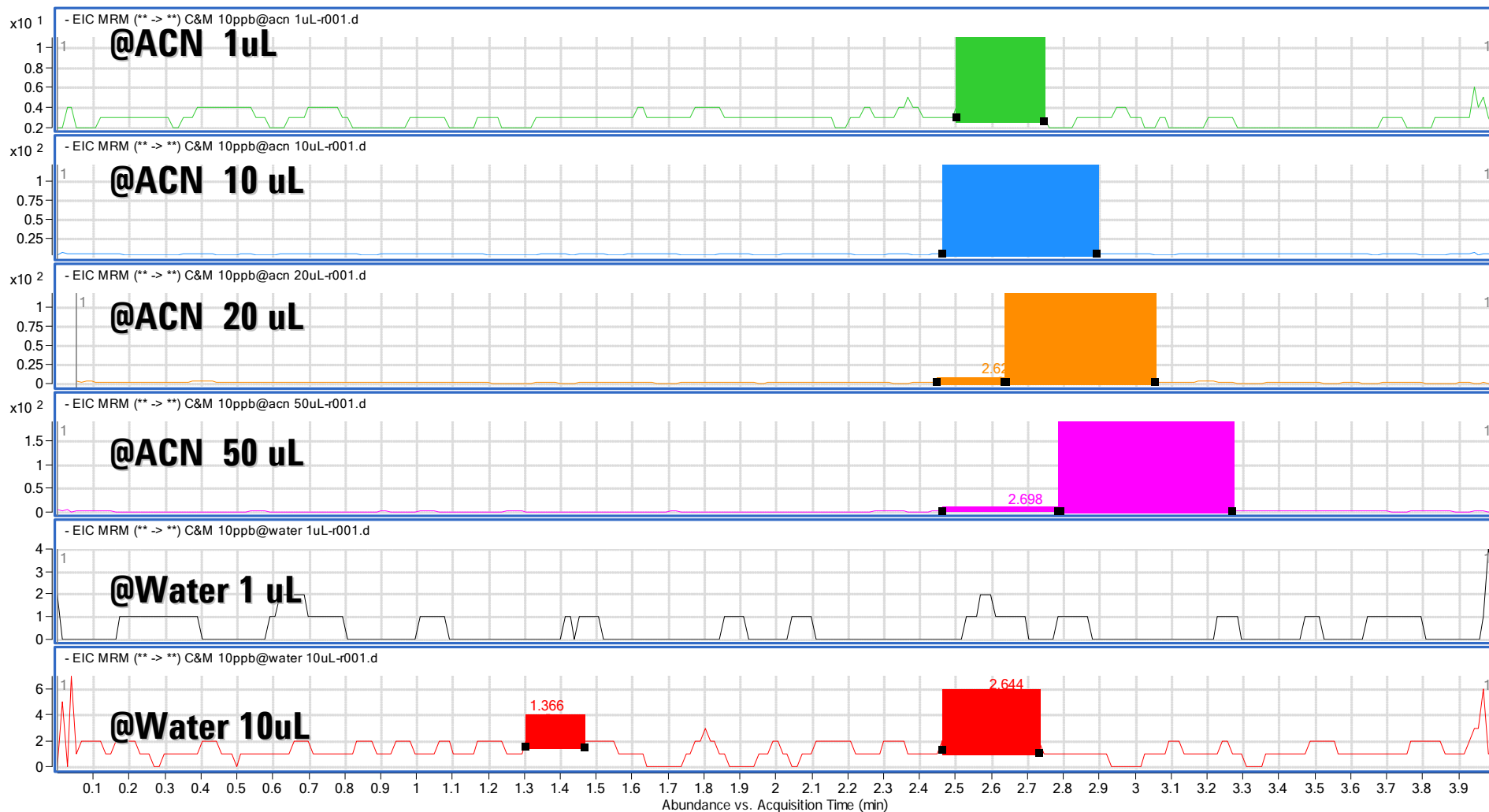
|                           |   |
|---------------------------|---|
| <b>MS system</b>          | <b>: Agilent 6410 LC/MS/MS</b>                        |
| <b>Ion source</b>         | <b>: ESI</b>  |
| <b>Polarity</b>           | <b>: Positive and Negative</b>                        |
| <b>Nebulizer gas</b>      | <b>: Nitrogen</b>                                     |
| <b>Ion spray voltage</b>  | <b>: 4000V</b>  |
| <b>Source temperature</b> | <b>: 350□</b>   |
| <b>Resolution</b>         | <b>: Q1 (unit) Q3 (unit)</b>                          |
| <b>Scan mode</b>          | <b>: Multiple Reaction Monitoring (MRM)</b>           |
| <b>Segment</b>            | <b>: Segment 1= 0~4min negative for cyanuric acid</b> |
|                           | <b>Segment 2= 4~6min positive for melamine</b>        |
| <b>Delta EMV</b>          | <b>: 600V</b>   |

# MRM Conditions

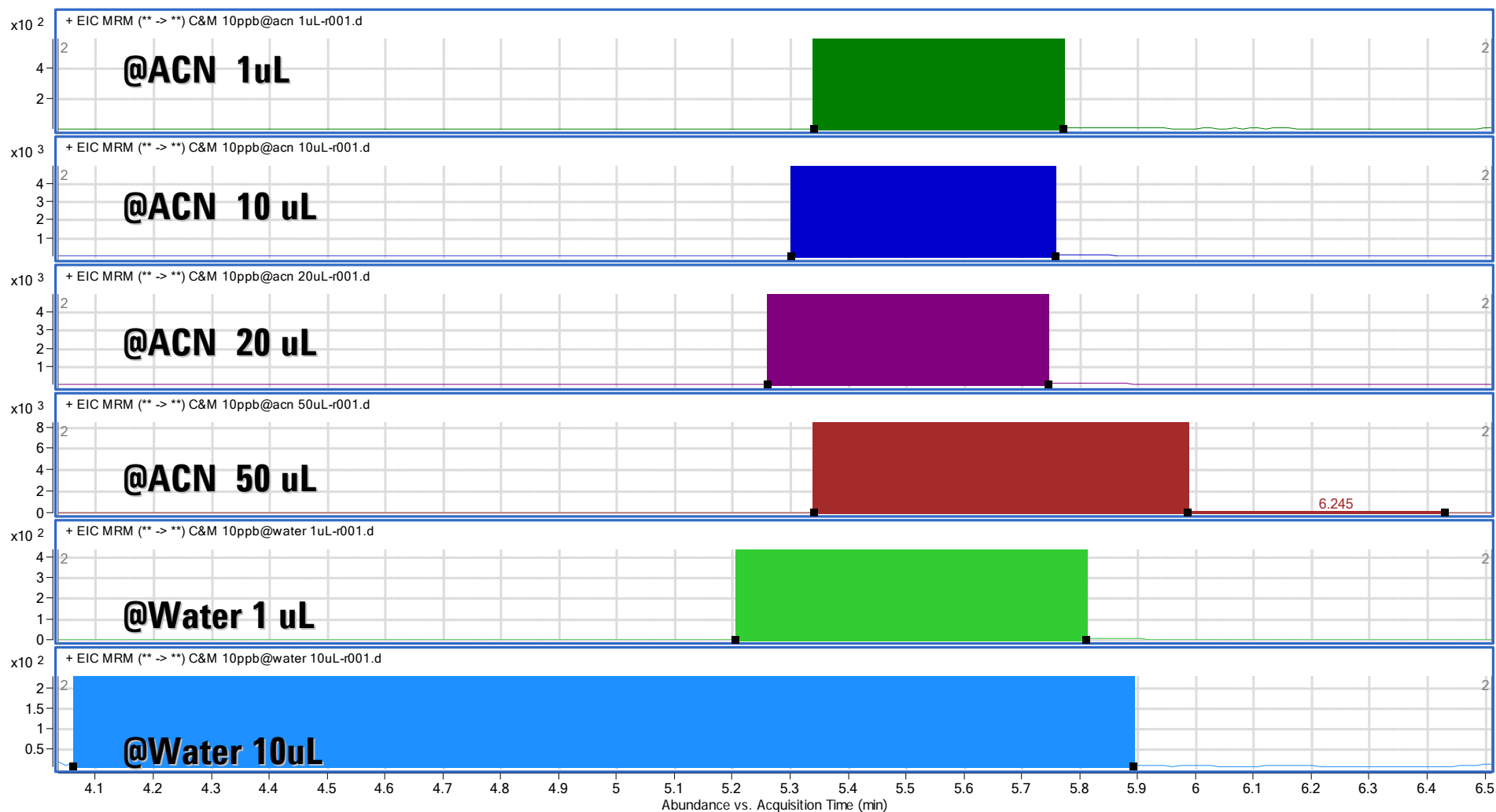


| Time | Compound      | Precursor | Product | Dwell | Fragmentor | Collision Energy |
|------|---------------|-----------|---------|-------|------------|------------------|
|      |               |           |         | (ms)  | (V)        |                  |
| 2.6  | Cyanuric Acid | 128       | 85      | 200   | 100        | 5                |
|      |               | 128       | 42      | 200   | 100        | 30               |
| 5.9  | Melamine      | 127       | 85      | 200   | 100        | 20               |
|      |               | 127       | 68      | 200   | 100        | 35               |

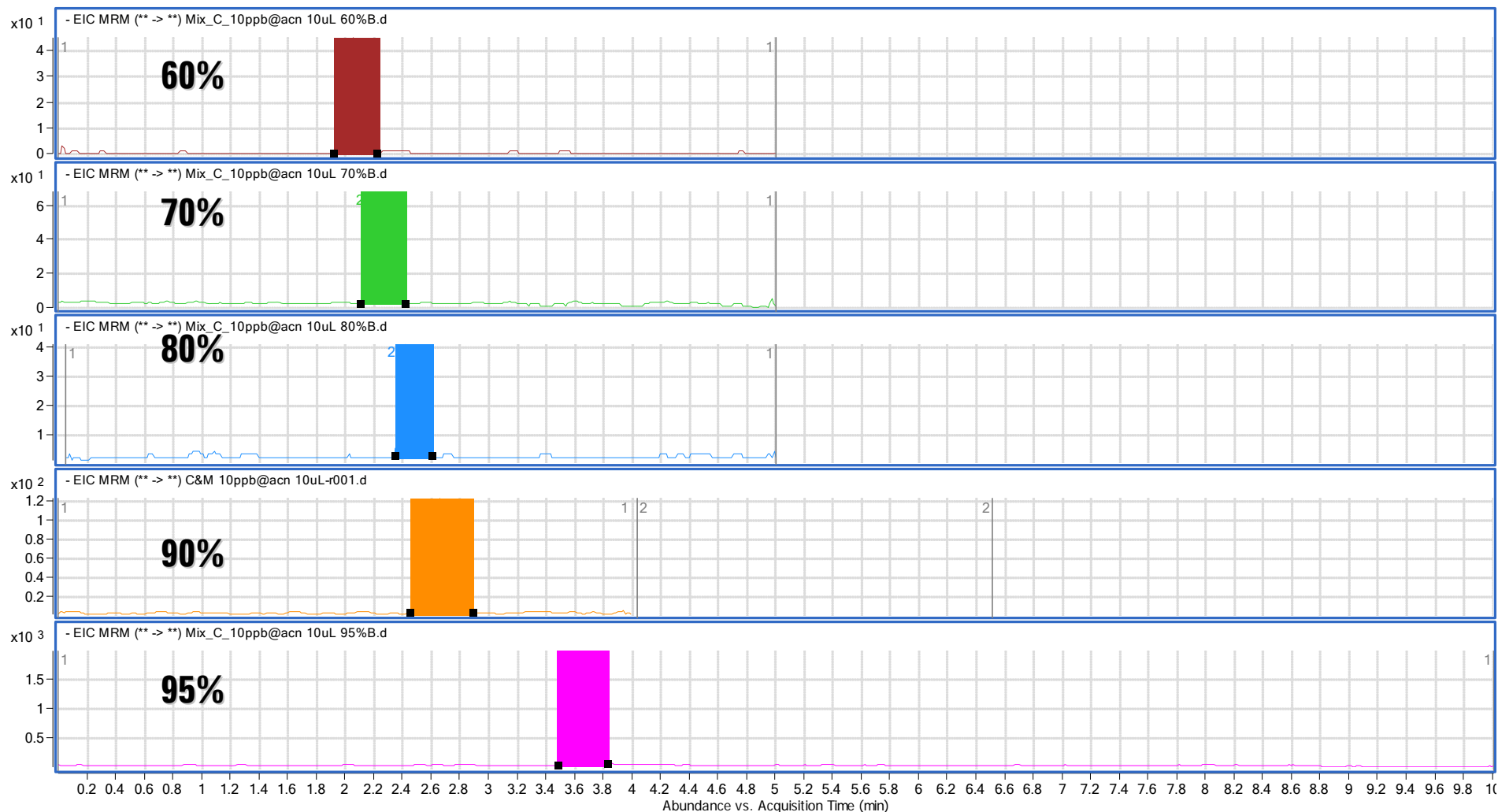
# Effect of Injection Solvent & Volume on Cyanuric Acid Retention



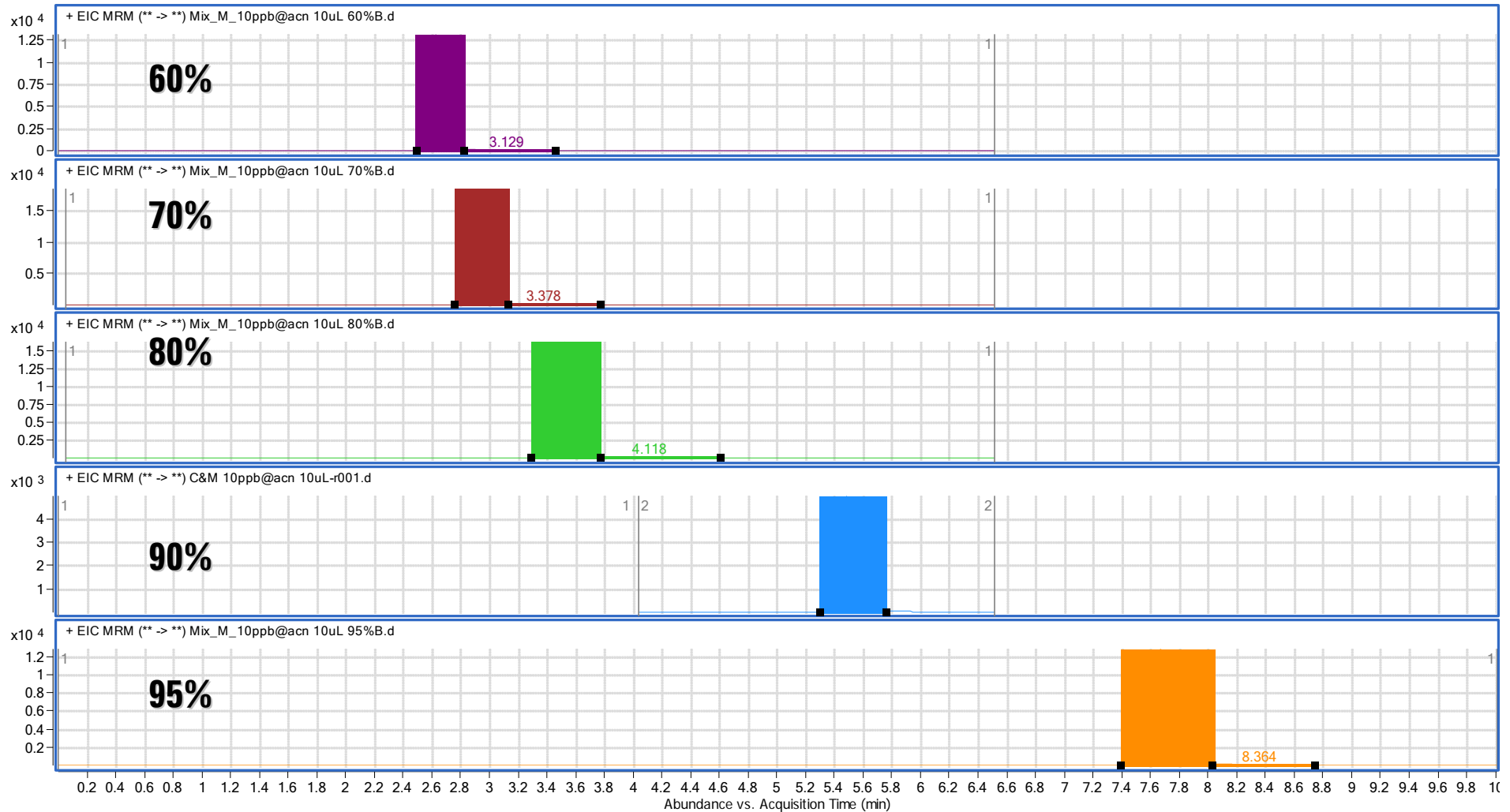
# Effect of Injection Solvent & Volume on Melamine Retention



# Effect of Increasing % ACN on Cyanuric Acid Retention

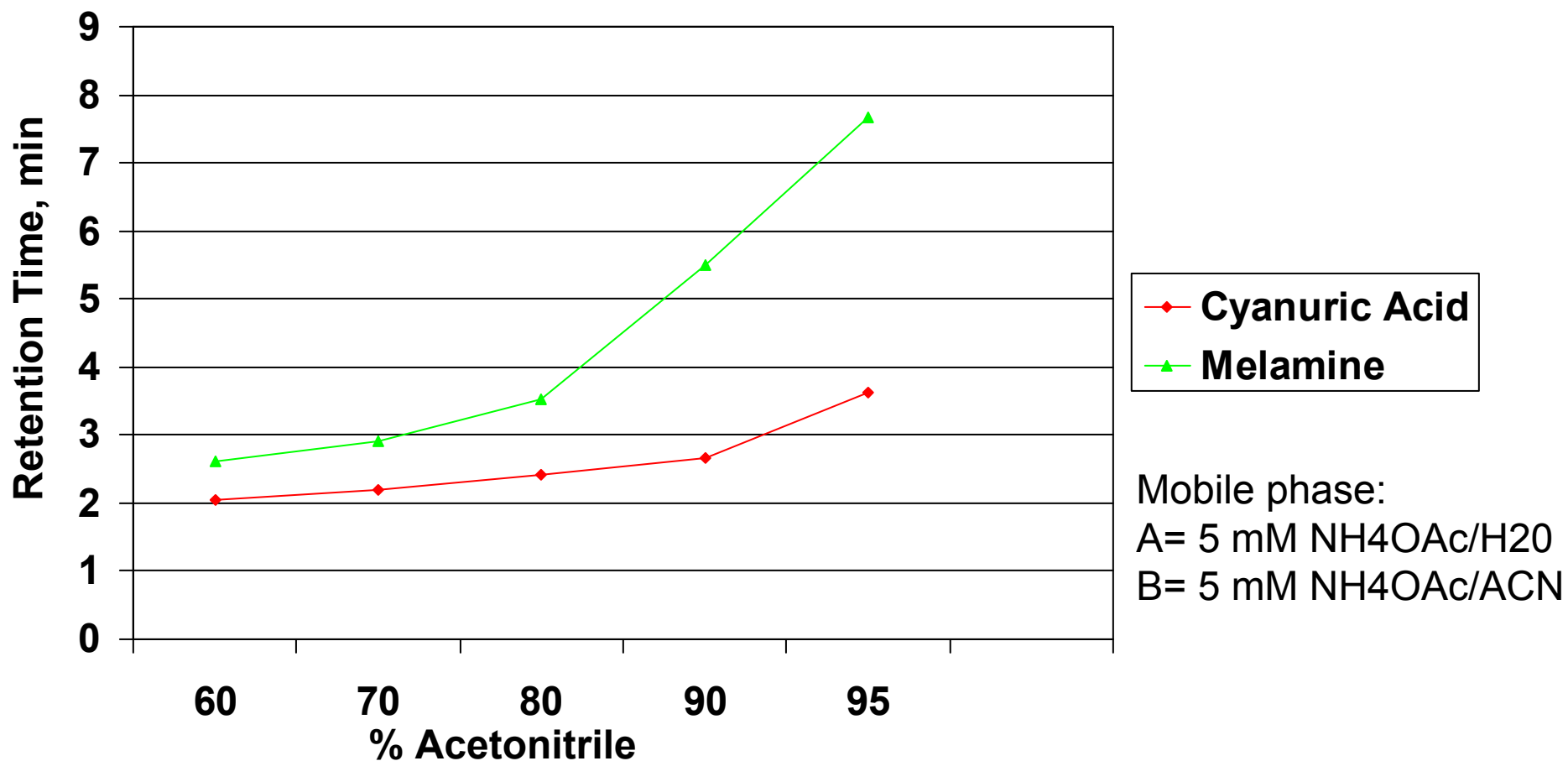


# Effect of Increasing %ACN on Melamine Retention

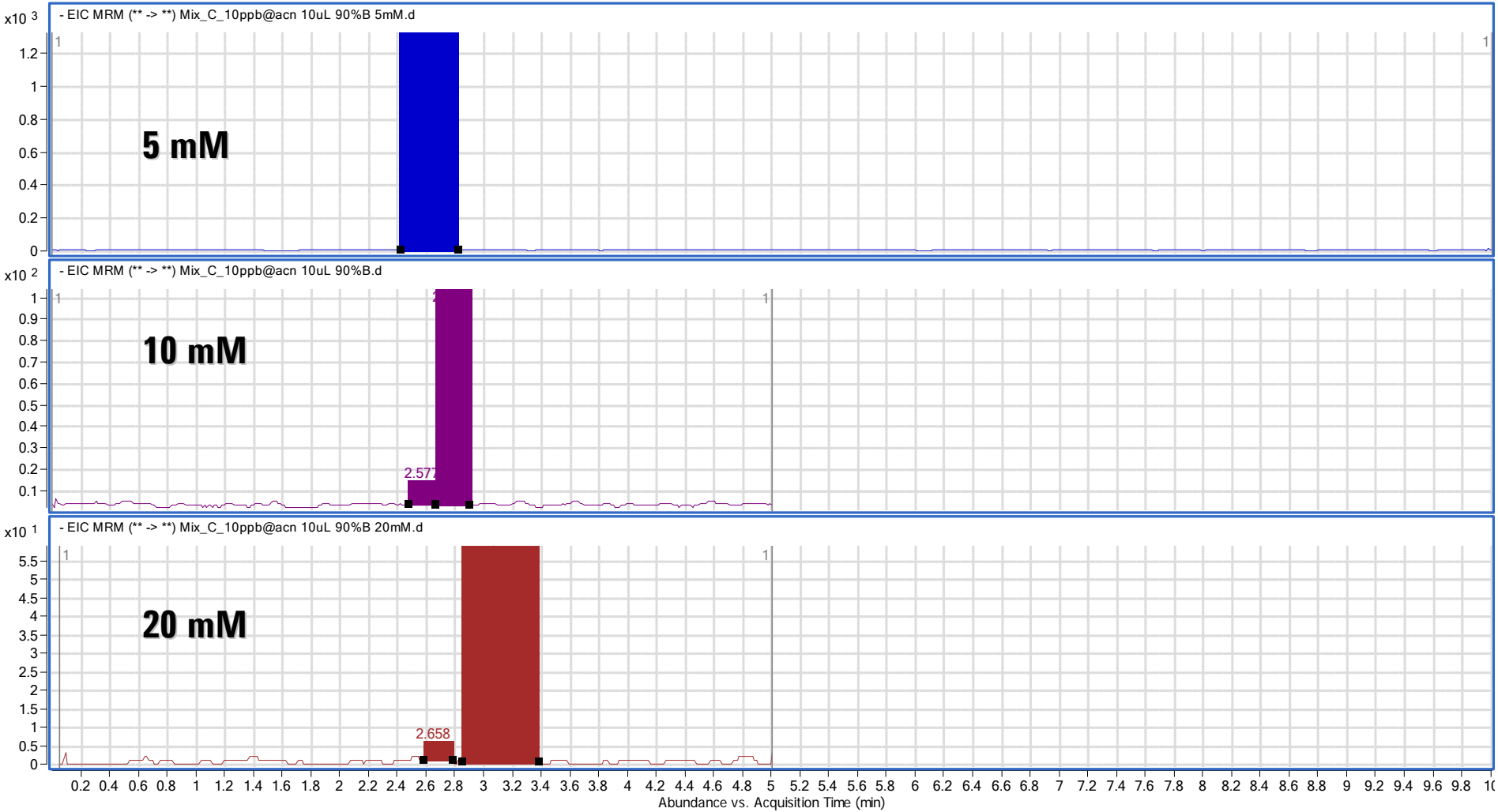




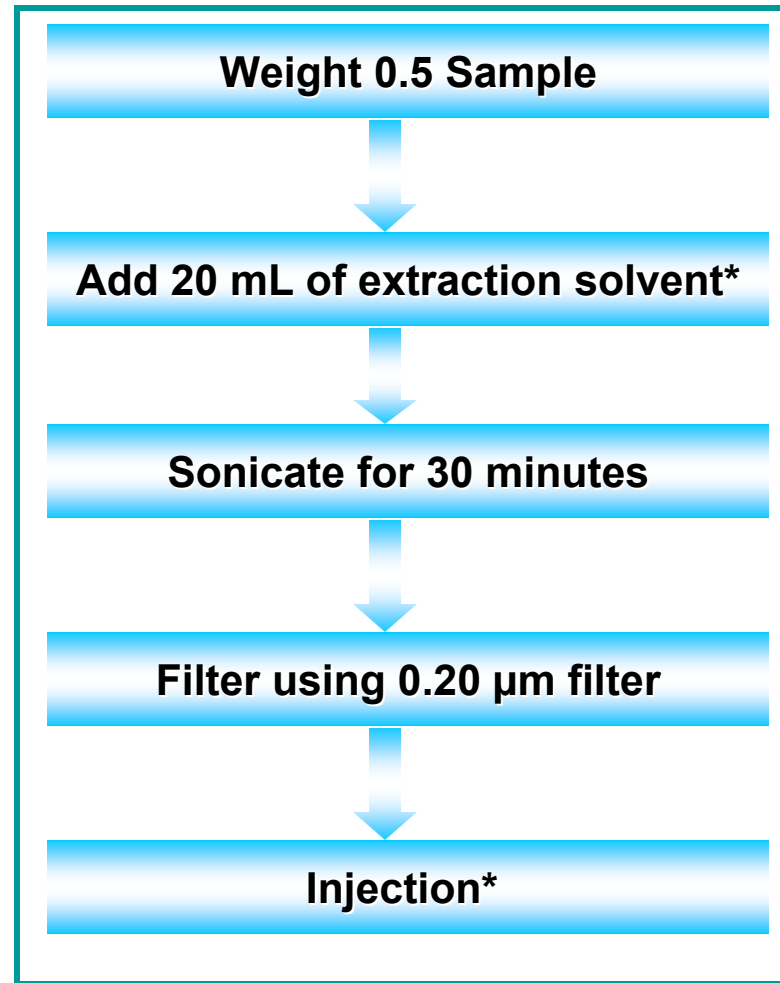
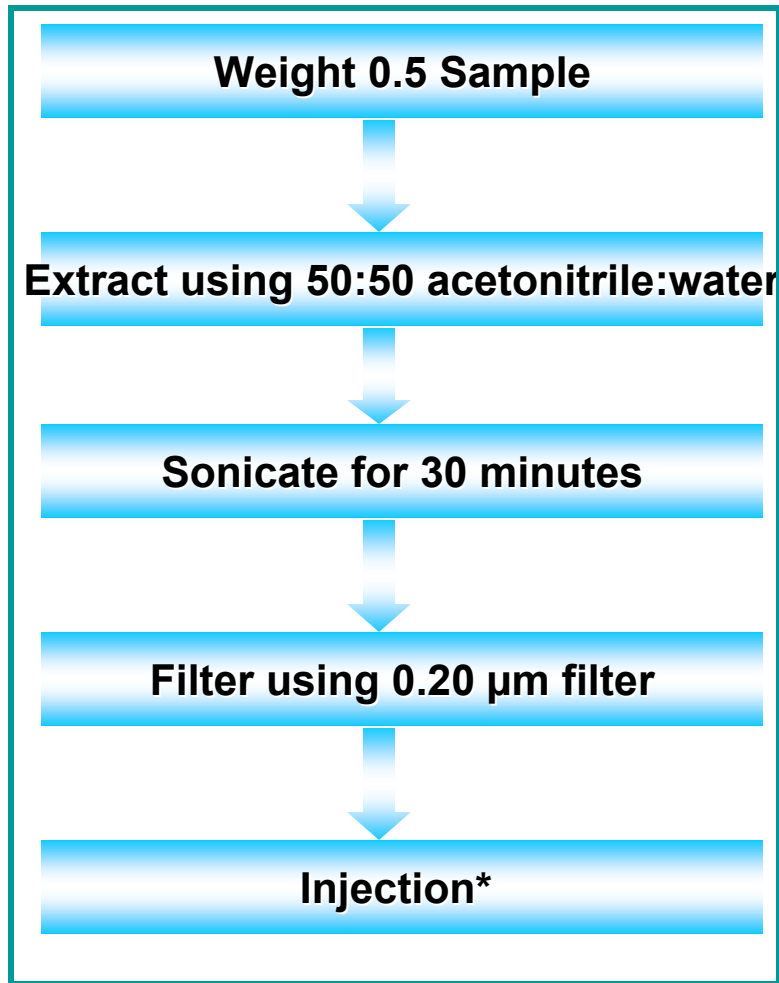
# Retention Time Changes with Changes in Mobile Phase Composition



# Effect of CH<sub>3</sub>COONH<sub>4</sub> Concentration on Cyanuric Acid Retention



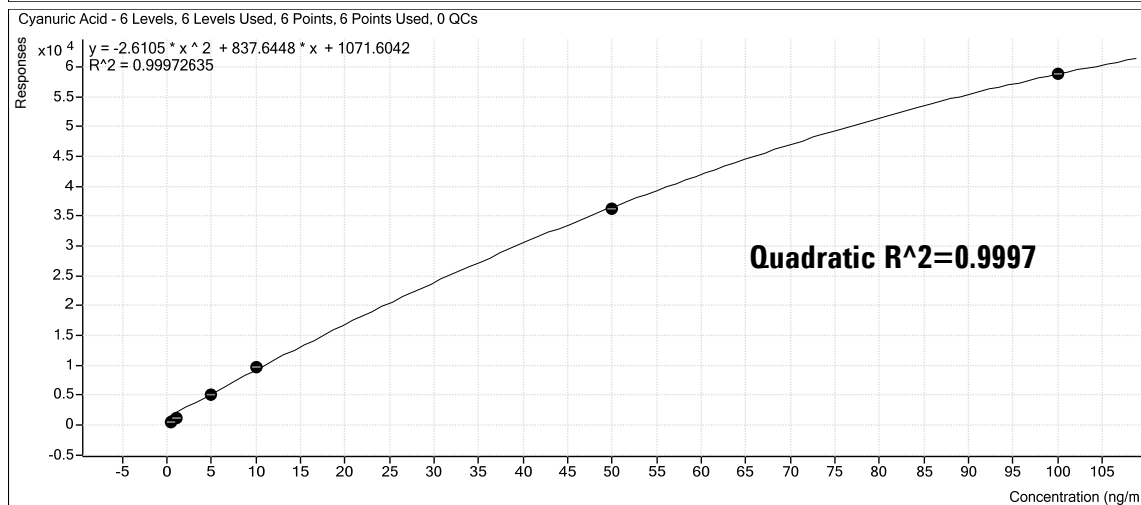
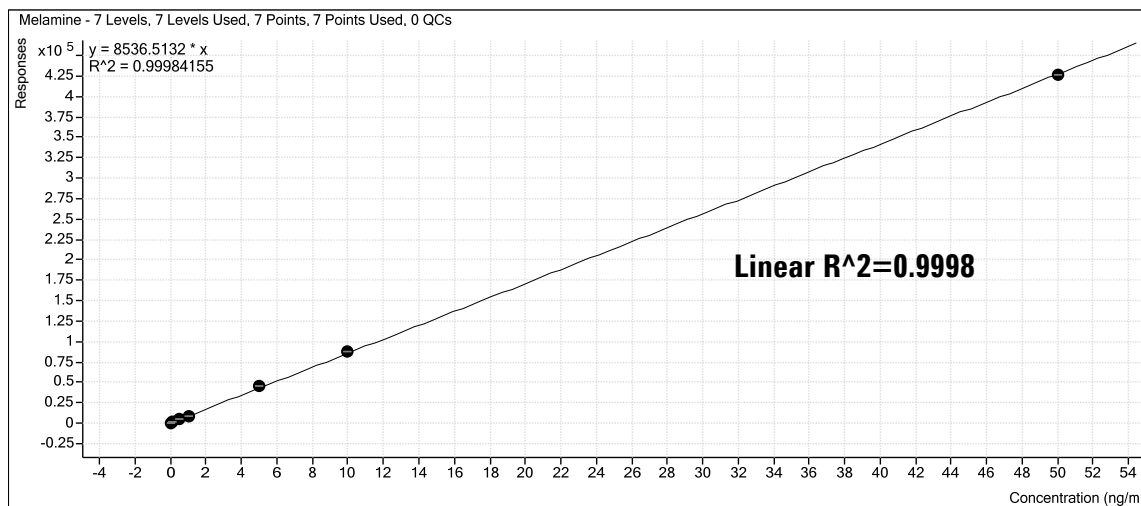
# Sample Treatment



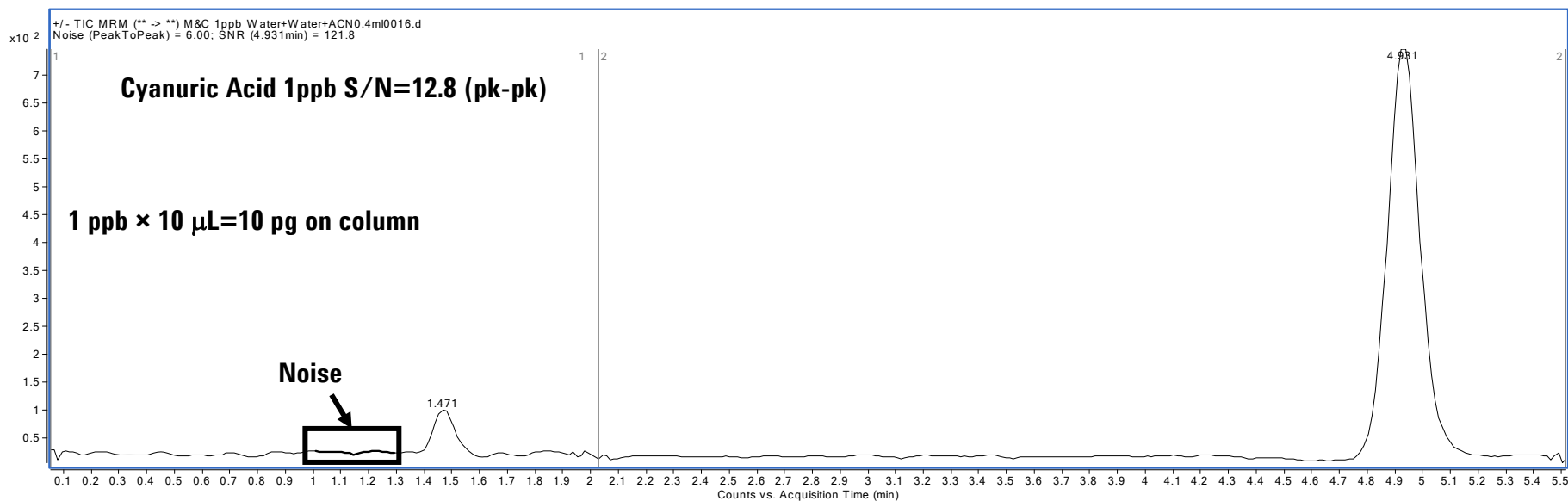
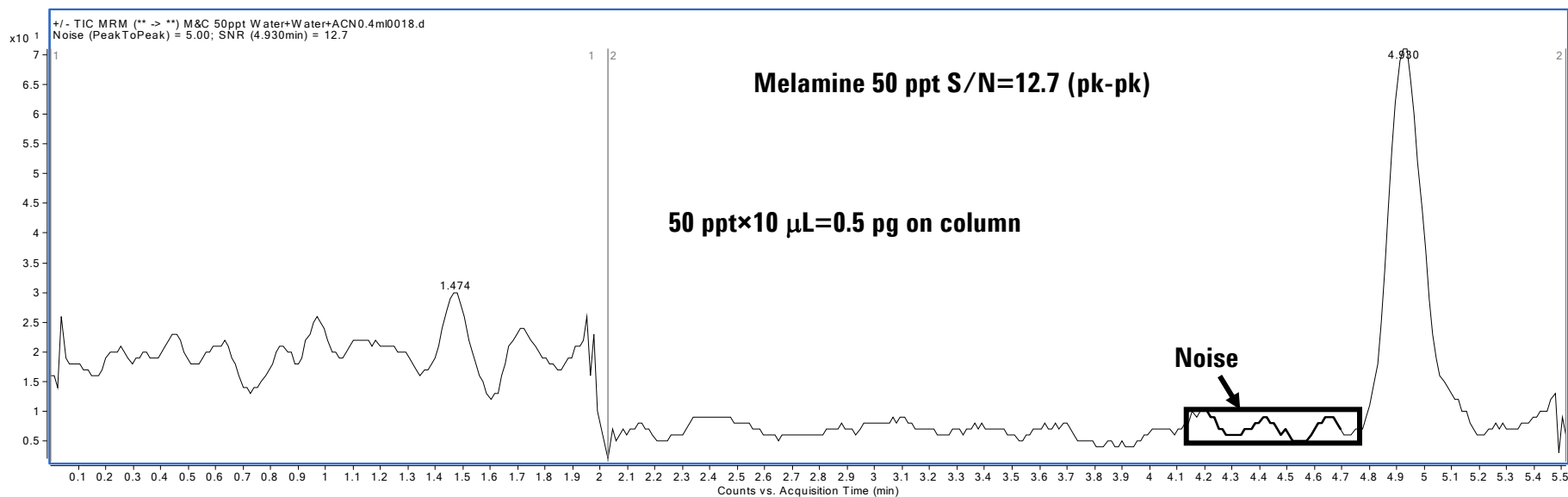
Matrices studied: wheat gluten, corn meal  
And rice protein 1

\*10:40:50 TEA : H<sub>2</sub>O : Acetonitrile  
2

# Calibration Curves for Melamine (Top) and Cyanuric Acid (Bottom) By LC-MS/MS

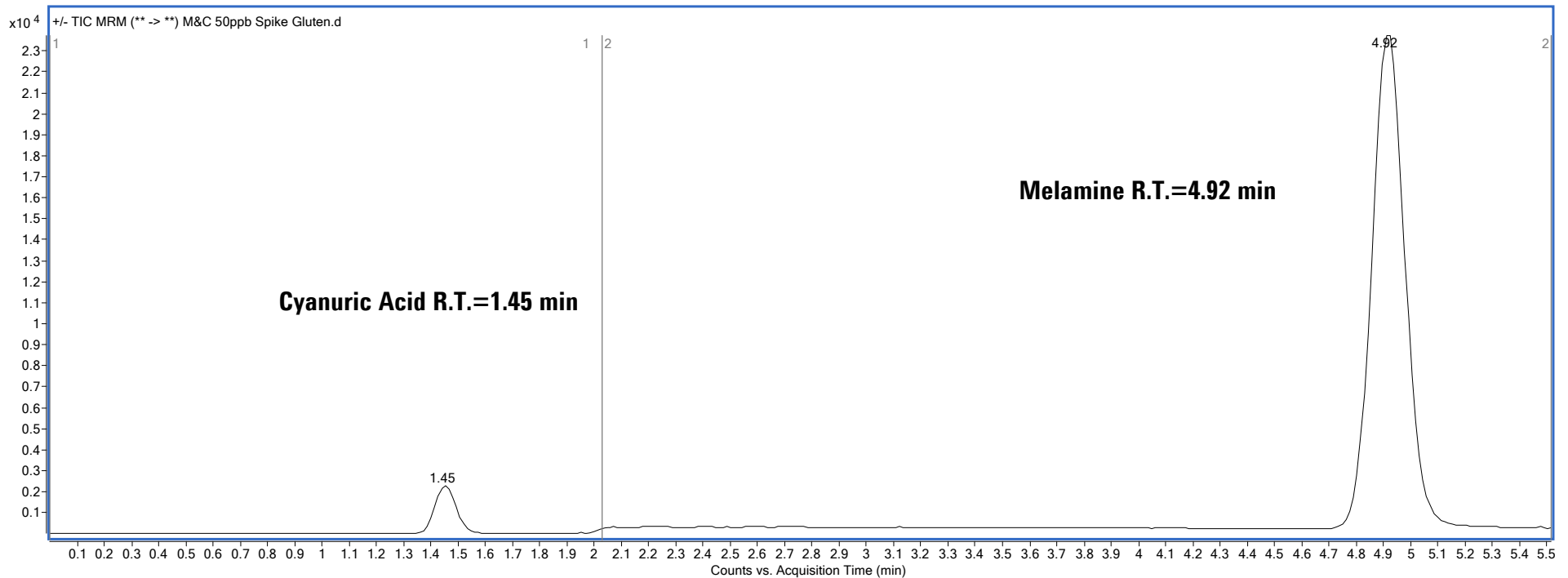


# Detection Limits for Melamine and Cyanuric Acid



# Total Reaction Monitoring Chromatogram for Spiked Wheat Gluten Sample

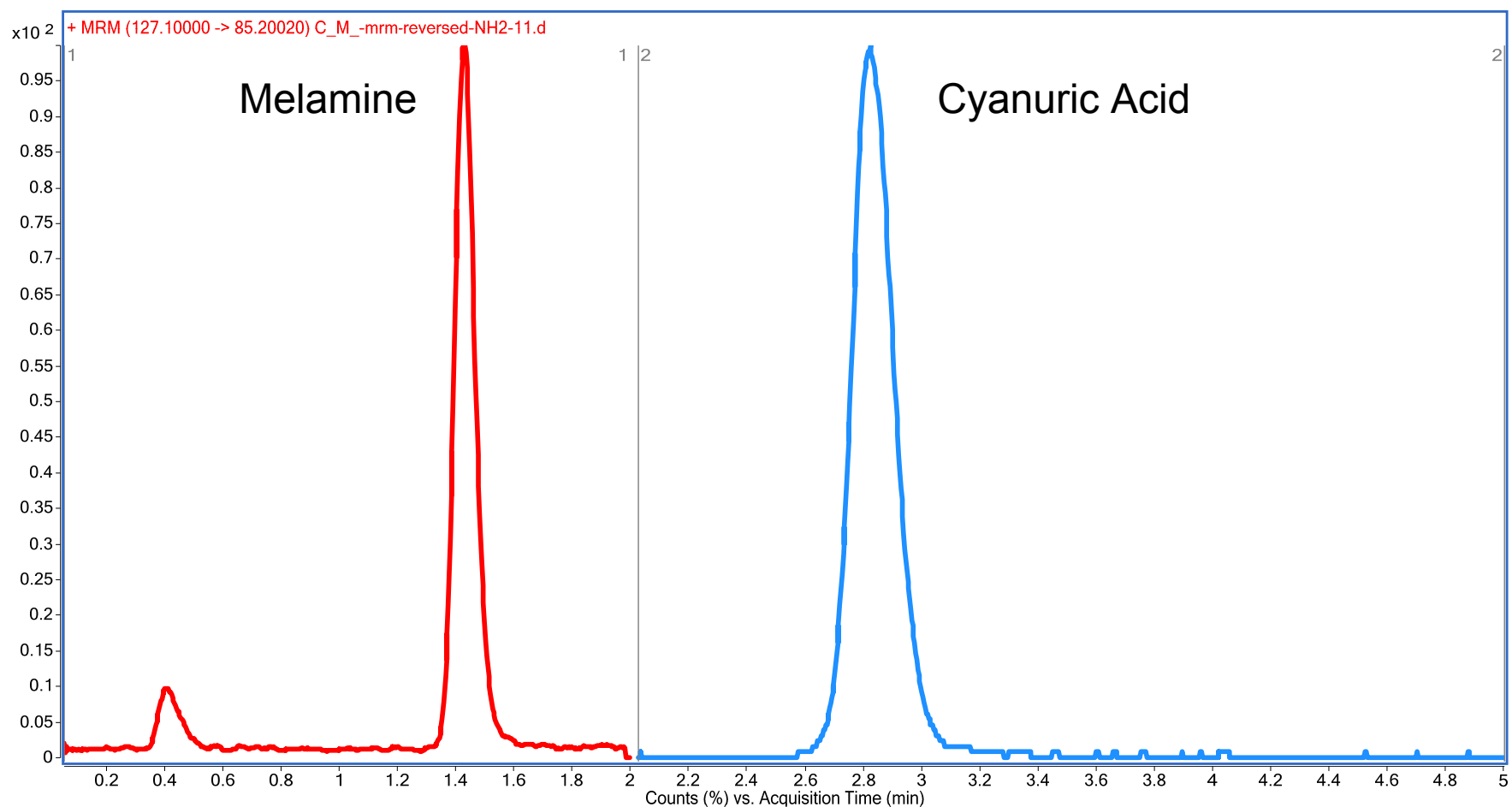
(50 ppb level)



# Comparison of Silica and Amino HILIC Columns

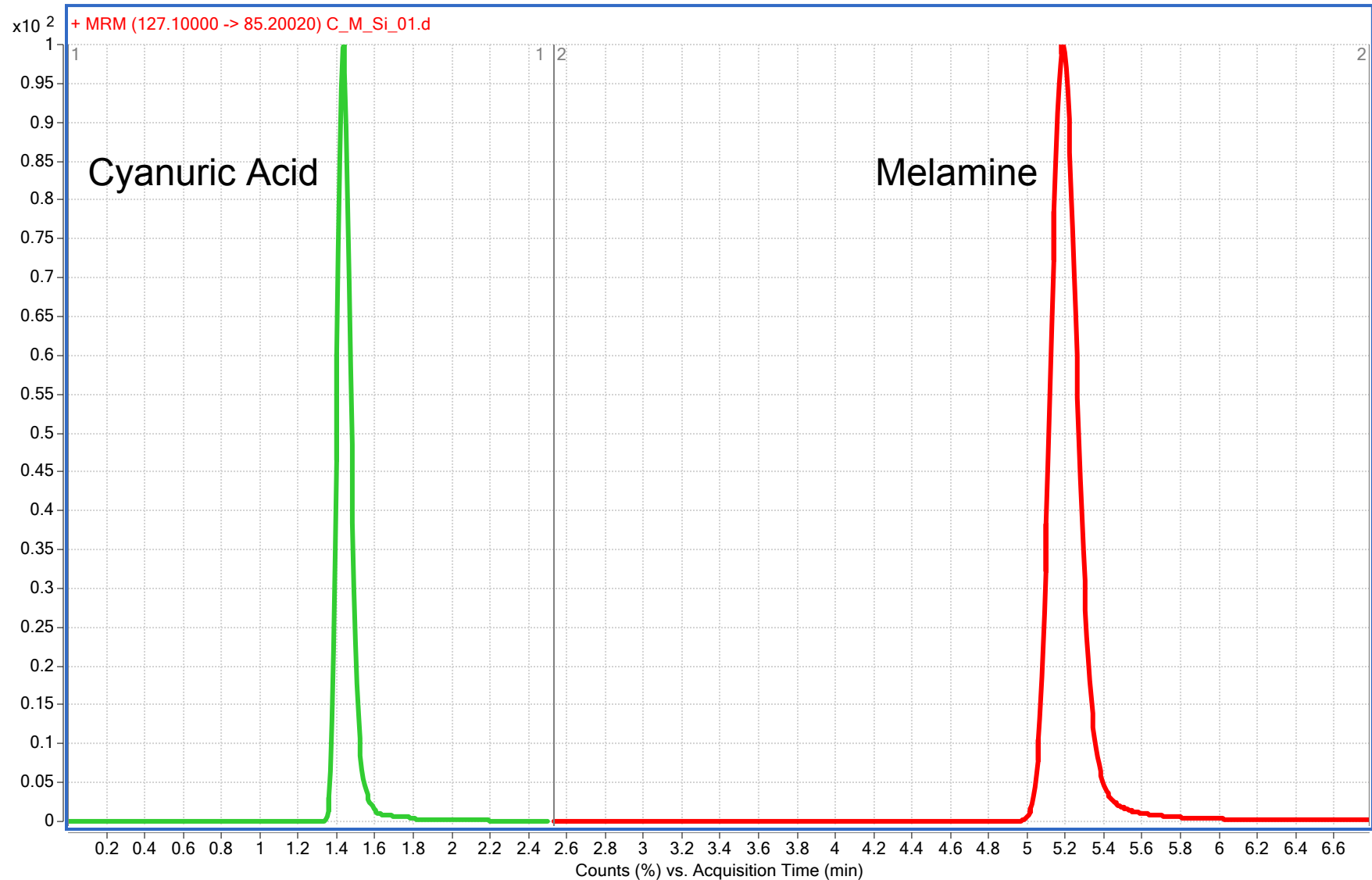
|                         |  |
|-------------------------|--|
| <b>HPLC system</b>      | <b>Agilent 1200 RRLC</b>                       |
| <b>Column 1</b>         | <b>Agilent Zorbax-NH2, 2.1×50 mm, 5 um</b>     |
| <b>Column 2</b>         | <b>Agilent Zorbax-Rx Sil, 2.1×150 mm, 5 um</b> |
| <b>Injection Volume</b> | <b>2 uL</b>                                    |
| <b>Temp</b>             | <b>25°C</b>                                    |
| <b>Flow rate</b>        | <b>0.4 mL/min</b>                              |
| <b>Mobile phase</b>     | <b>A - 5 mM Ammonium acetate in Water</b>      |
|                         | <b>B - 5 mM Ammonium acetate in ACN</b>        |

# HILIC Separation Using Zorbax NH<sub>2</sub>, Column 1

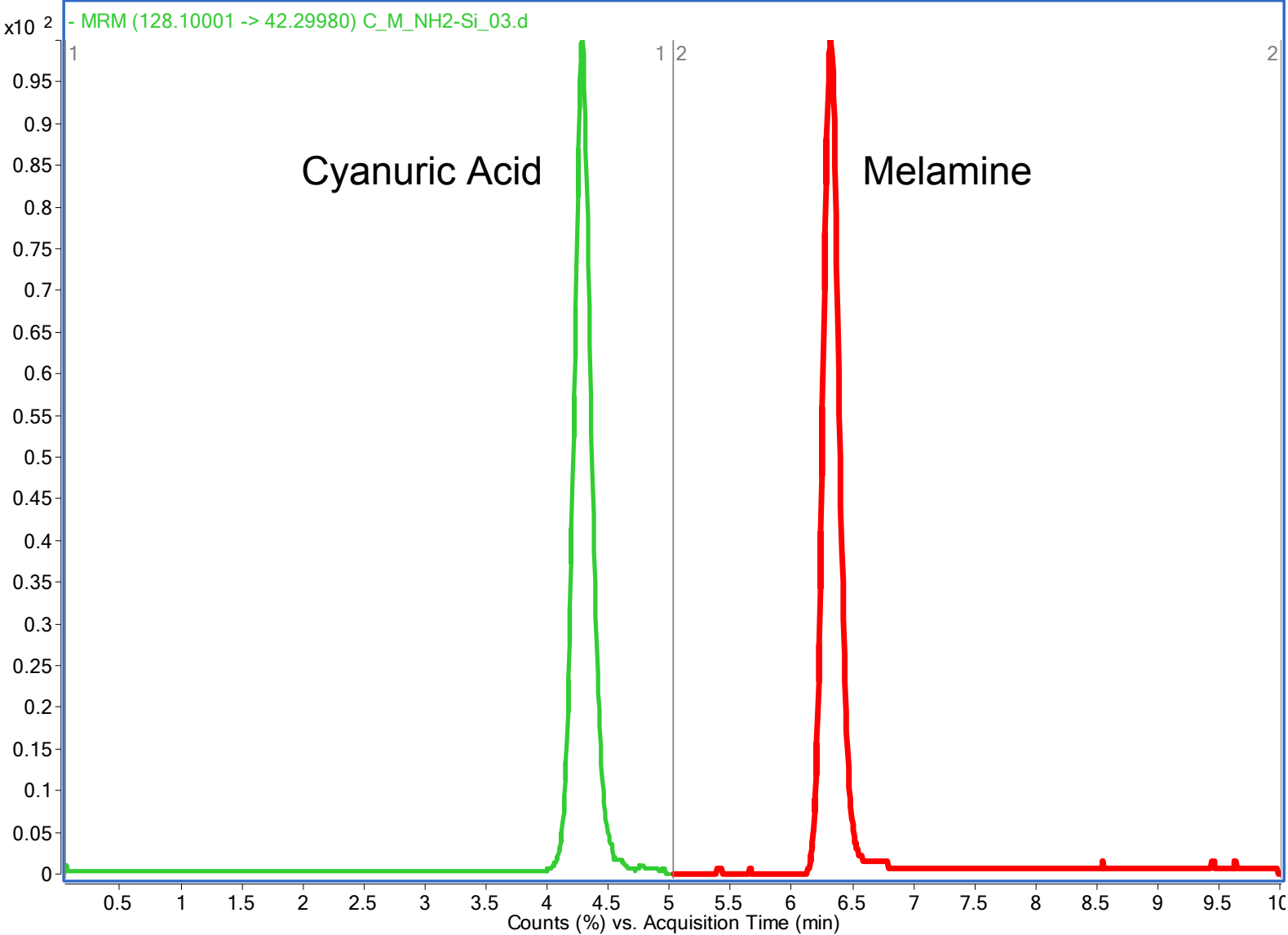




# HILIC Separation Using Zorbax Rx-Sil Column 2



# HILIC Separation with Both Columns in Series



# Conclusions

- HILIC has received renewed attention for the analysis of polar compounds that are unretained or poorly retained on RPC columns
- The technique is readily adaptable to MS and MS-MS detection
- 13 of the 23 new “specialty” columns introduced at Pittcon 2008 by 9 companies\*
- HILIC used by 4% of all users\*\*
- Publications are accelerating at exciting pace

\* Pittcon 2008 Report, LCGC No. America, March, 2008

\*\* R.E. Majors, LCGC No. Amer. Column Survey, 2007

# Acknowledgements

- **Wei Luan and Yanyan Fang, Agilent, China**
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