

# Improved OPA / FMOC Derivatized Amino Acid Methods using Many Column Configurations for a Range of Speed and Resolution Options

Henderson J.W. Jr., Brooks, A., Joseph, M.  
Agilent Technologies 2850 Centerville Rd. Wilmington, Del. USA 19808

FBA32-Tu  
HPLC2009  
Dresden, Germany

Agilent Technologies

## Introduction

A well established online automated OPA/FMOC derivitization method for amino acids has been improved and updated. The updated method can be used with a variety of ZORBAX Eclipse Plus C18 column configurations including 5, 3.5, and 1.8  $\mu$ m particle sizes, column lengths from 250 to 50 mm, and column inner diameters of 4.6, 3.0 and 2.1 mm in order to provide the sensitivity and flexibility for fast analyses and a variety of sample types and sizes. Scalability, lot-to-lot reproducibility, linearity, and data will be presented. Ten column options will be shown, ranging from rapid nine minute analyses of 23 amino acids including re-equilibration using short (50 mm) ZORBAX Rapid Resolution High Throughput columns (1.8  $\mu$ m), to 40 minute analyses using 250 mm traditional 5  $\mu$ m columns.

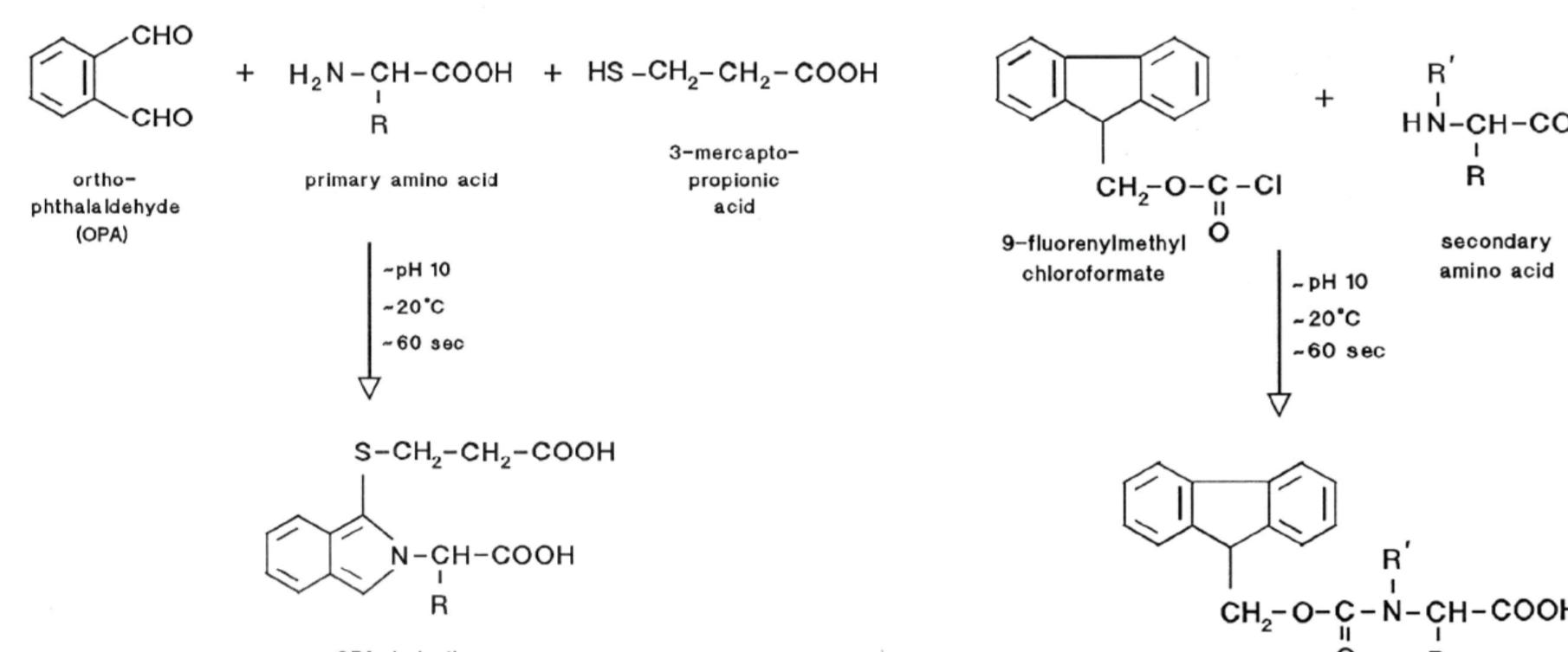
## The Ten ZORBAX Eclipse Plus C18 Options

#	Method Category	Column (mm)	Analysis Time with Re-equilibration	Typical Minimum Resolution Factor	Approx. mL Solvent/Analysis*	Agilent
1	Traditional High Resolution	4.6 x 250, 5 $\mu$ m	40 min	2.4	64	1200 or 1200 SL
2		3.0 x 250, 5 $\mu$ m	40 min	2.4	28	
3	Rapid Resolution	4.6 x 150, 3.5 $\mu$ m	25 min	2	42	1200 or 1200 SL
4		3.0 x 150, 3.5 $\mu$ m	25 min	2	18	
5		2.1 x 150, 3.5 $\mu$ m	25 min	2	12	
6	Rapid Resolution	4.6 x 100, 1.8 $\mu$ m	16 min	2.4	28	1200SL
7	High Throughput	2.1 x 100, 1.8 $\mu$ m	16 min	2.4	8	
8		4.6 x 50, 1.8 $\mu$ m	9 min	1.5	23	
9		3.0 x 50, 1.8 $\mu$ m	9 min	1.5	10	
10		2.1 x 50, 1.8 $\mu$ m	9 min	1.5	5	

\* includes injector program and pre run DAD  
autobalancing (2.42 min), and re-equilibration time.

## The Online Pre-Column Derivatizations

The primary amino groups react with ortho-phthalaldehyde (OPA) in the presence of 3-mercaptopropionic acid (3-MPA) at about pH 10 to form an isouindole derivative. Secondary amino groups do not react. The OPA derivatized amino acid is then detected by UV at 338 nm.



The secondary amino groups react with 9-fluorenylmethyl chloroformate (FMOC) at about pH 10 to form a secondary amide. Secondary amino groups do not react. The FMOC derivatized amino acid is then detected by UV at 262 nm.

## Method Parameters

### Amino Acid Identification and Detection

- |                  |                |                    |
|------------------|----------------|--------------------|
| 1. Aspartic acid | 9. Arginine    | 17. Phenylalanine  |
| 2. Glutamic acid | 10. Alanine    | 18. Isoleucine     |
| 3. Asparagine    | 11. Tyrosine   | 19. Leucine        |
| 4. Serine        | 12. Cysteine   | 20. Lysine         |
| 5. Glutamine     | 13. Valine     | 21. Hydroxyproline |
| 6. Histidine     | 14. Methionine | 22. Sarcosine      |
| 7. Glycine       | 15. Norvaline  | 23. Proline        |
| 8. Threonine     | 16. Tryptophan |                    |

Primary Amino Acids 1-20 are detected at UV wavelength 338nm (DAD1, Sig=338,10 Ref=390,20 TT)

Secondary Amino Acids 21-23 are detected at UV wavelength 262nm (DAD1, Sig=262,4 Ref=390,20)

A programmed signal wavelength change from 338 nm to 262 nm is determined by choosing a switching time after lysine elutes and before hydroxyproline elutes

### The Automated Derivatization

G1376C well plate automatic liquid sampler (WPALS): G1329A automatic liquid sampler (ALS):

- 1) Draw 2.5  $\mu$ L from Borate vial (Agilent PN 5061-3339)
- 2) Draw 1.0  $\mu$ L from Sample vial
- 3) Mix 3.5  $\mu$ L in washport 5X
- 4) Wait 0.2 min
- 5) Draw 0.5  $\mu$ L from OPA vial (Agilent PN 5061-3335)
- 6) Mix 4.0  $\mu$ L in washport 10X max speed
- 7) Draw 0.4  $\mu$ L from FMOC vial (Agilent PN 5061-3337)
- 8) Mix 4.4  $\mu$ L in washport 10X max speed
- 9) Draw 32  $\mu$ L from Injection Diluent vial
- 10) Mix 20  $\mu$ L in washport 8X
- 11) Inject
- 12) Wait 0.1 min
- 13) Valve bypass

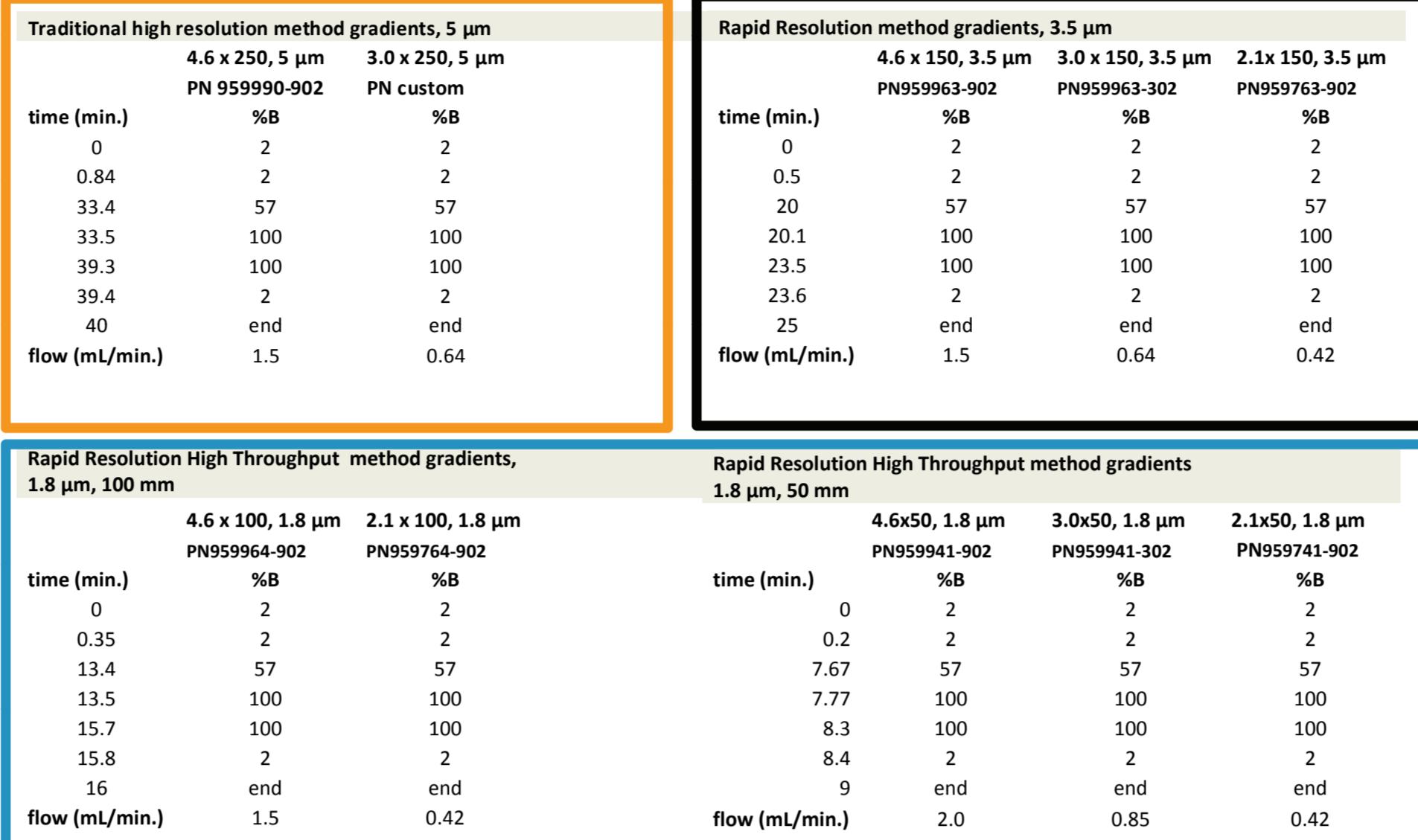
### The Agilent LC Flow Paths

The flow path is recorded carefully because it can have a significant effect on resolution, especially for gradients (delay volume) and 2.1 i.d. columns (extra column volume).

Method name	Traditional (5 $\mu$ m High Resolution Methods)		Rapid Resolution (3.5 $\mu$ m Methods)		
	4.6 x 250, 5 $\mu$ m	3.0 x 250, 5 $\mu$ m	4.6 x 150, 3.5 $\mu$ m	3.0 x 150, 3.5 $\mu$ m	2.1 x 150, 3.5 $\mu$ m
LC Model	1100	1200	1200 SL	1200 SL	1200 SL
Pump	G1312A	G1311A quat	G1312B	G1312B	G1312B
Dampener/static mixer	yes	n/a	yes	yes	bypassed
Purge valve to ALS	G1328-87000 (green 500 mm)	G1328-87000 (green 500 mm)	G021-1823 (red 400 mm)	G021-1823 (red 400 mm)	G021-1823 (red 400 mm)
ALS	G1367A	G1329A	G1367C	G1367C	G1367C
Needle seat	G1367-87101 (green)	G1311-87201 (green)	G1367-87202 (red)	G1367-87202 (red)	G1367-87202 (red)
ALS to heat exchanger	G1313-87305 (green 180 mm)	G01090-87611 (red 105 mm)	G01090-87611 (red 105 mm)	G01090-87611 (red 105 mm)	G01090-87611 (red 105 mm)
Heat exch. to column or guard	G1316 3 $\mu$ L	G1316 3 $\mu$ L	G1316-80003 1.6 $\mu$ L	G1316-80003 1.6 $\mu$ L	G1316-80003 1.6 $\mu$ L
Optional guard cartridge	820950-936 4 $\mu$ L, 4.6 id	821125-936-4pk, 2.1 id	820950-936 4 $\mu$ L, 4.6 id	821125-936-4pk, 2.1 id	821125-936-4pk, 2.1 id
Column	959990-902	custom	959963-902	959963-302	959763-902
Post column to union	5065-9931 (200 mm green)	5065-9931 (200 mm green)	n/a	n/a	n/a
ZDV union to flow cell	5022-2184	5022-2185	n/a	n/a	n/a
Detector	G1315B	G1315D	G1315C	G1315C	G1315C
Flow cell	2 $\mu$ L G1315-60024	2 $\mu$ L G1315-60024	2 $\mu$ L G1315-60024	2 $\mu$ L G1315-60024	2 $\mu$ L G1315-60024
RRHT 1.8 $\mu$ m Methods (100 mm)	4.6 x 100, 1.8 $\mu$ m	2.1 x 100, 1.8 $\mu$ m	4.6 x 50, 1.8 $\mu$ m	3.0 x 50, 1.8 $\mu$ m	2.1 x 50, 1.8 $\mu$ m
Method name	4.6 x 100, 1.8 $\mu$ m	2.1 x 100, 1.8 $\mu$ m	4.6 x 50, 1.8 $\mu$ m	3.0 x 50, 1.8 $\mu$ m	2.1 x 50, 1.8 $\mu$ m
LC Model	1200	1200	1200 SL	1200 SL	1200 SL
Pump	G1312 B	G1312 B	G1312 B	G1312 B	G1312 B
Dampener/static mixer	yes	bypassed	yes	bypassed	bypassed
Purge valve to ALS	5021-1823 (red 400 mm)	5021-1823 (red 400 mm)	5021-1823 (red 400 mm)	5021-1823 (red 400 mm)	5021-1823 (red 400 mm)
ALS	G1367C	G1367C	G1367C	G1367C	G1367C
Needle seat	G1367-87201 (red)	G1367-87201 (red)	G1367-87201 (red)	G1367-87201 (red)	G1367-87201 (red)

## The Linear Gradients

The gradient profile (%) is identical for all columns. The different gradient delay times are mitigated by reducing delay volume and the isocratic hold in the beginning of the gradient program.



## The Mobile and Stationary Phase

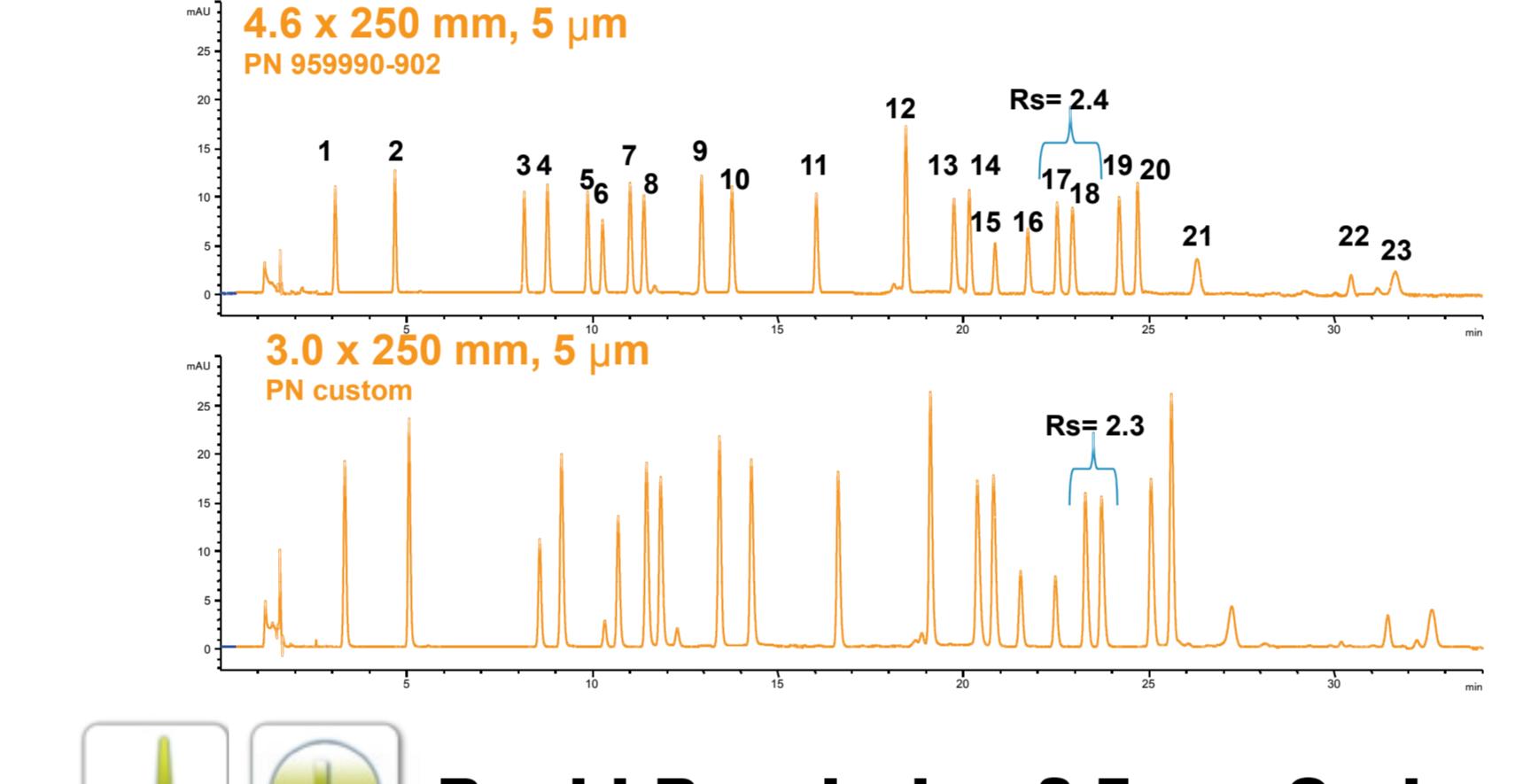
### Stationary Phase: ZORBAX Eclipse Plus C18

Column Temperature: 40 °C  
Mobile Phase A: 10 mM Na<sub>2</sub>HPO<sub>4</sub>; 10 mM Na<sub>2</sub>B<sub>4</sub>O<sub>7</sub>, pH 8.2; 5 mM NaN<sub>3</sub>  
Mobile Phase B: Acetonitrile: Methanol: Water (45:45:10, v: v: v)  
Injection Diluent: (0.25 mL H<sub>3</sub>PO<sub>4</sub> + 100 mL H<sub>2</sub>O)

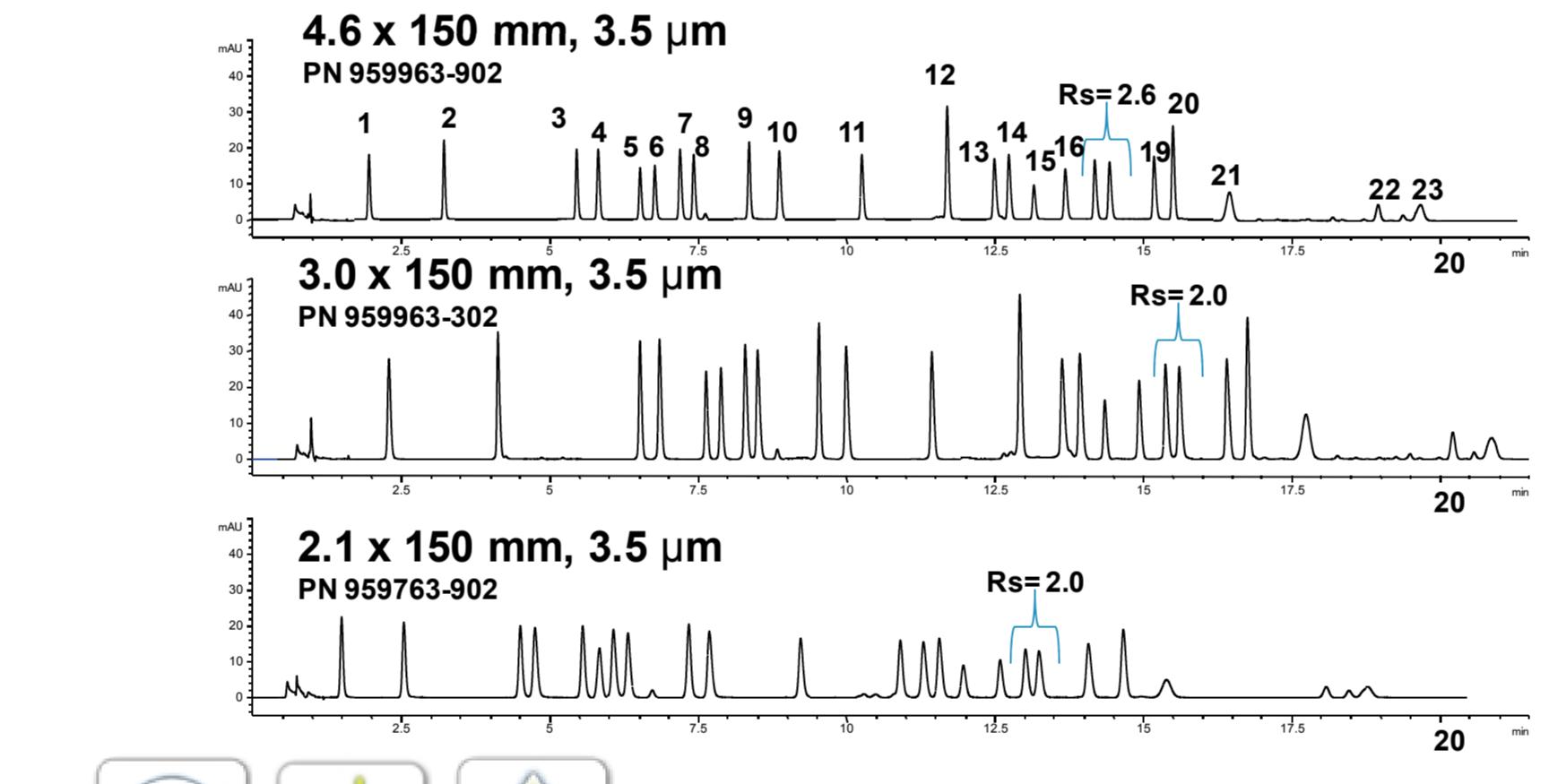
## Results



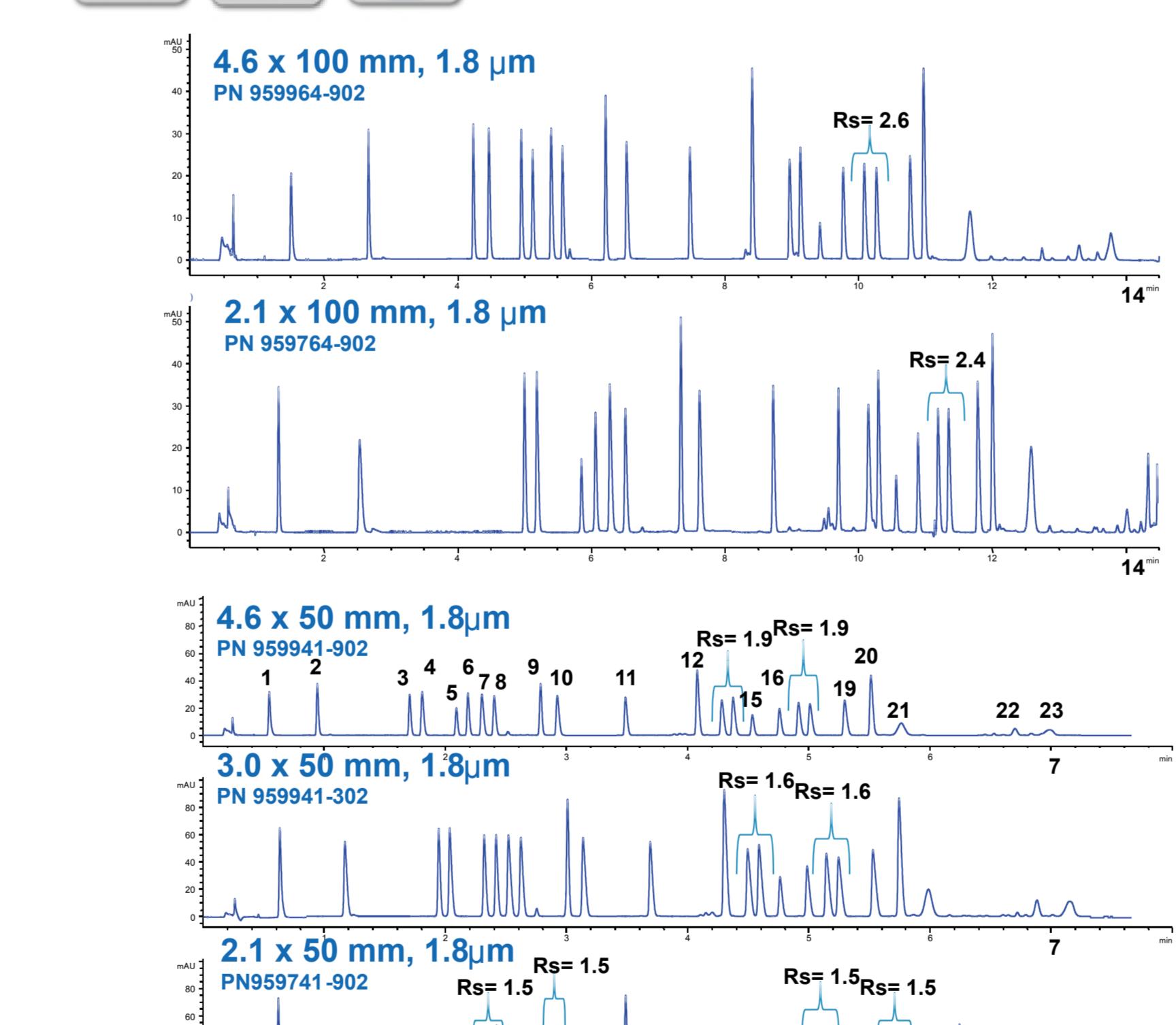
### The Eclipse Plus C18 5 $\mu$ m Options



### Rapid Resolution 3.5 $\mu$ m Options



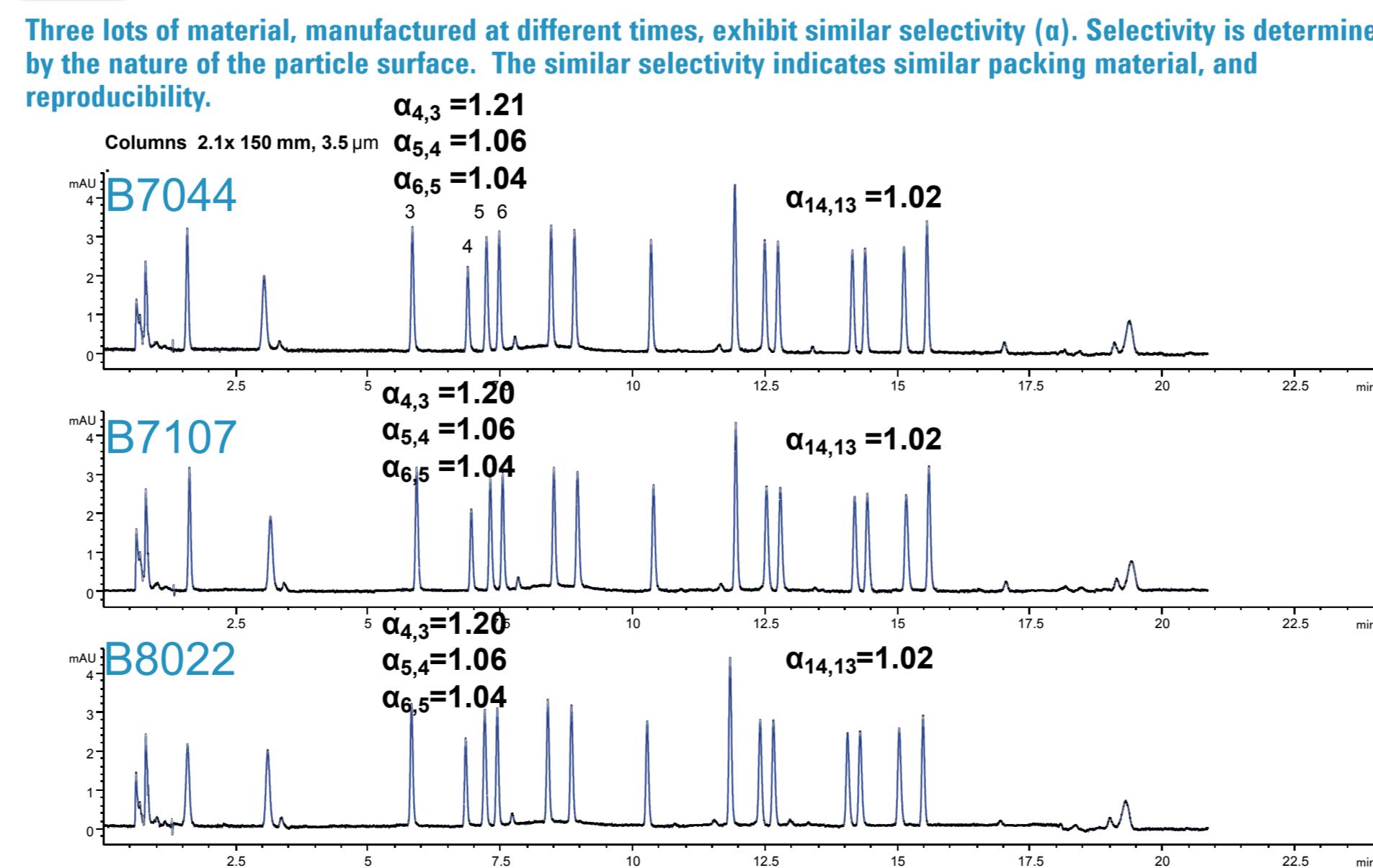
### RRHT 1.8 $\mu$ m Options



## Method Ruggedness

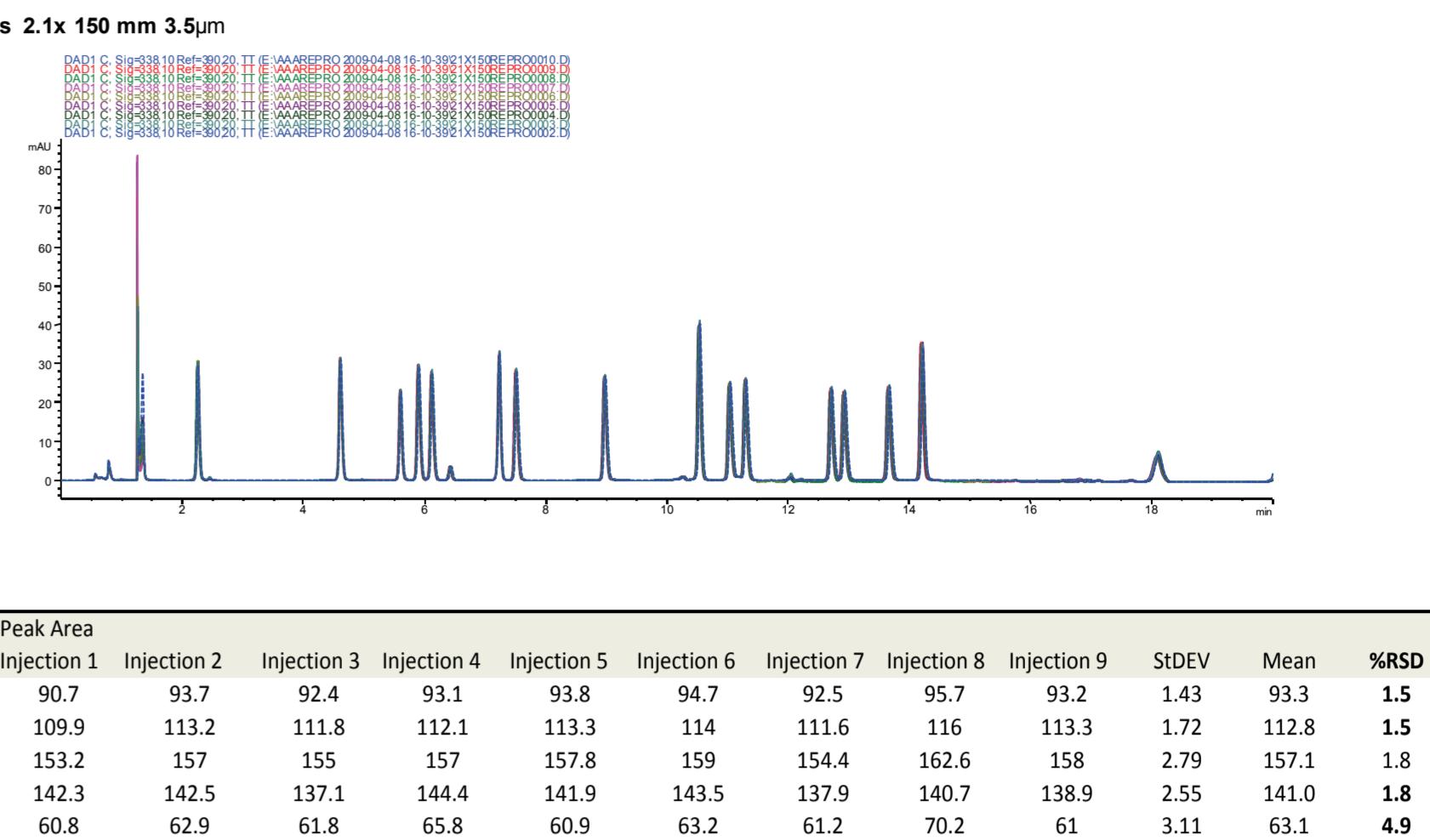
### Lot-to-Lot Reproducibility

Three lots of material, manufactured at different times, exhibit similar selectivity ( $\alpha$ ). Selectivity is determined by the nature of the particle surface. The similar selectivity indicates similar packing material, and reproducibility.



## Injection-to-Injection Reproducibility

Overlay of eight sequential injections showing reproducibility of the online derivatization and gradient programs. Peak area of early, middle and late eluting amino acids are statistically tabulated below. The other amino acids had similar statistics.



### Lifetime

Calibration curves of early, middle and late eluting amino acids show linearity over 1 to 1000 pmol/ $\mu$ L range using the Eclipse Plus C18, 2