

AN1605: Identification of insulin oligomeric states using SEC-MALS

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Summary

Insulin, one of the most important mammalian hormones, regulates a multitude of metabolic functions including the control of the blood glucose level in the body. Under healthy conditions, insulin is produced and stored in the islet tissues of the pancreas and released depending on the metabolic situation. In patients suffering from diabetes, insulin cannot be sufficiently produced by the body. It has to be administered as a pharmakon via oral or injection pathways.

Under physiological conditions, insulin forms hexamer complexes in the presence of zinc ions. In the pancreas the hormone is also stored as zinc complexes. In pharmacological preparations therefore zinc is added to enable complex formation. To ensure safe and effective administration of insulin formulations, it is of vital interest to investigate, which oligomeric species are present in a specific preparation.

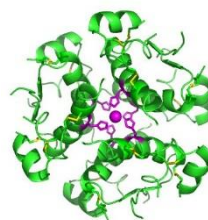
This application note shows how the combination of column chromatography (SEC/GPC) with Multi-Angle Light Scattering (MALS), Dynamic Light Scattering (DLS) and RI detection can be applied as a powerful tool to identify monomers, dimers, hexamers and higher aggregates of insulin. Using this approach, each preparation can be comprehensively characterized to determine optimal formulation, storage and administration conditions for the patient's benefit.

Introduction

In the analysis of proteins and peptides the question arises frequently whether the molecules under investigation are monomeric or form dimers or higher aggregates. The knowledge of this stoichiometric relation enables the

investigator to estimate if the protein is present in its biologically active form.

In such determinations the first step is usually the separation of the molecular species and their characterization.



To achieve this, size exclusion chromatography (SEC) is often used as the method of choice. Since this approach separates the molecules according to their hydrodynamic volumes, it does not automatically allow the exact mass determination

because molar mass is not necessarily a function of retention time but rather due to hydrodynamic properties.

A technology which overcomes this limitation is multi-angle light scattering (MALS). MALS measures molar mass in an absolute manner. This means no assumptions on the molecule's structure are made and no molar mass standards are needed. The determination relies on first principle measurements only and is therefore absolute. The MALS detection can be coupled to online DLS measurement to obtain hydrodynamic radii of the molecules. Moreover, CG-MALS measurements have been shown to be useful to detect the aggregation status of insulin^{1,2}.

The subject of the investigations presented here is the hormone insulin. Insulin plays a vital role in the regulation of mammalian blood glucose level and metabolism, cell growth and fat metabolism – only to name a few of its manifold functions. The discovery of insulin in 1921 represented a starting point for a revolution in the treatment of diabetes. Human insulin is a protein consisting of two peptide chains of 21 and 30 amino acids respectively, with total molar mass of 5.8 kDa. Human insulin self-associates and in the presence of zinc ions a hexameric complex is formed, 2 zinc ions bind per hexamer. Insulin is

stored in the β -cells of the pancreas as zinc complexes. In pharmaceutical preparations insulin is normally formulated at concentrations where the self-association is pronounced and hence zinc is added to stabilize the hexamer and thereby a more stable formulation. Upon dilution after delivery the hexamer dissociates and ends up as monomers in the blood stream. Hence it is of interest to investigate the oligomerization both from a formulation as well as pharmacological perspective.

Materials and Methods

An Agilent 1260 HPLC system was used which included an auto sampler, an isocratic pump and a degasser. The separation column was a Superose 12 300/10 from GE Healthcare.

For the detection we used a variable wavelength detector at 280 nm (Agilent), a [DAWN[®] 18-angle light scattering detector](#) equipped with a [WyattQELS[™]](#) for online DLS detection, and an [Optilab[®] refractive index detector](#). Molar masses were calculated from the MALS and dRI signals. The MALS, DLS and dRI detection instruments, as well as [ASTRA[®]](#) software for SEC-MALS analysis, were from Wyatt Technology.

The mobile phase consisted of 10 mM Tris, 140 mM NaCl, 2 mM Phenol and 200 ppm NaN₃ at pH = 7.7.

Three different insulin sample preparations all containing 0.6 mM insulin were analyzed:

- Sample 1: insulin analogue without added zinc
- Sample 2: human insulin with 0.1 mM zinc
- Sample 3: human insulin with 0.3 mM zinc

Results and Discussion

Reversible protein assembly that increases with protein concentration, or self-association, is a well-known effect for e.g. monoclonal antibodies and insulin³. In the case of insulin, the molecule forms dimers and other oligomers which can be separated by SEC and detected by light scattering techniques^{4,5}. Here we show the characterization of different insulin preparations by SEC-MALS.

Oligomers and self-association

Results of Sample 1 are shown with molar masses indicated throughout the chromatogram (Figure 1). The major fraction with molar mass corresponding to a

monomer is well separated from the very minor fraction corresponding to hexamer, and the molar mass is observed to be quite uniform across the main peak. Although not present as resolved peaks in UV, MALS analysis indicates a molar mass distribution that covers the range from monomer to hexamer.

It should be emphasized that light scattering identified the hexamer easily even though the UV signal suggests that only a very small amount of this fraction is present. This is due to the extremely high sensitivity of the LS detection method for large molecules.

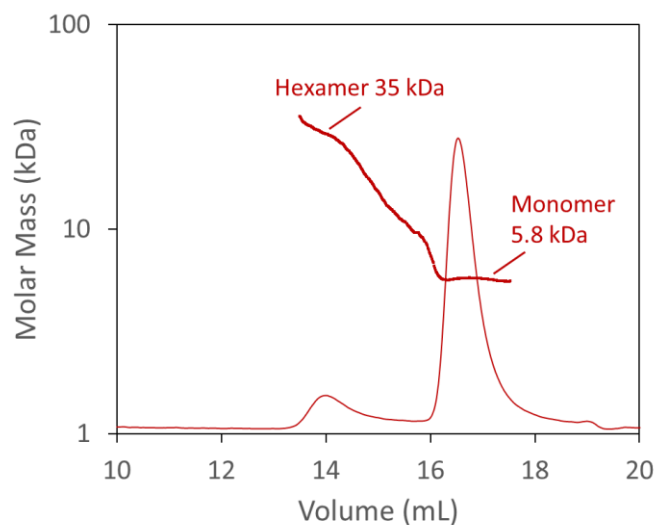


Figure 1. Plot of molar mass vs. time of Sample 1. The UV signal @ 280 nm is plotted as an overlay.

Formulation effects

Comparing sample 1 and 2 shows that the addition of 0.1 mM zinc lead to a shift in the equilibrium, going from mainly monomer in sample 1 to major fraction hexamer in Sample 2 (Figure 2). It is apparent from MALS analysis (not obvious from UV) that monomers and dimers exists in the minor peak of sample 2 and that the major peak is the hexamer.

Despite the long tail of the hexamer peak, MALS rules out the presence of oligomers with molar masses between dimer and hexamer. The tail is a result of non-ideal interactions between the hexamers and the column packing.

The presence of monomers and dimers, with the fraction of dimers increasing with concentration, might indicate dynamic equilibrium. This is further explored below.

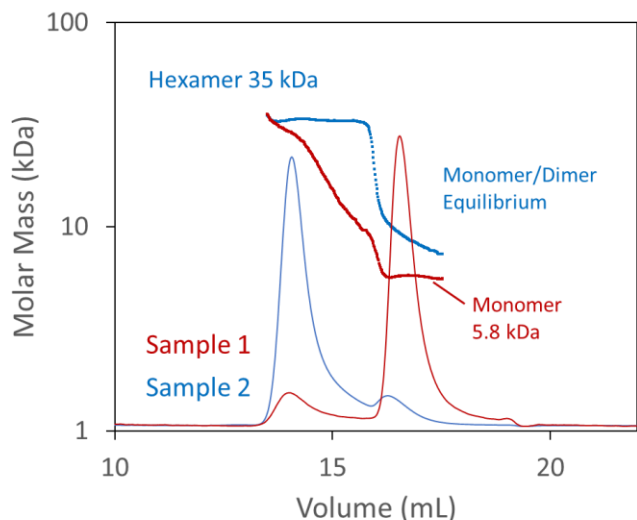


Figure 2: Plot of molar mass vs. time of Samples 1 and 2 (monomer, dimer, hexamer). The UV signal @ 280 nm is plotted as an overlay.

Further addition of zinc, in Sample 3, completely shifts the equilibrium towards the hexamer, as seen in Figure 3. In sample 3 another molecular species is detected which is identified as the dodecamer of the insulin molecule. It shows a molar mass of 70 kDa and, although present only in a small amount, generates a distinct signal clearly observed in by the LS detector.

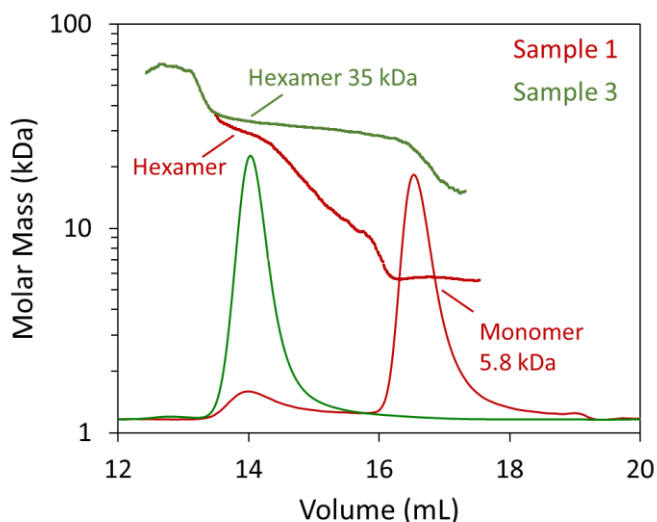


Figure 3: Plot of molar mass vs. time of Samples 1 and 3. The UV signal @ 280 nm is plotted as an overlay.

Concentration effects

SEC-MALS allows the determination of self-association tendencies. The effect of injection volumes on the sample was investigated, i.e. 50 and 200 μL of the same sample solution was injected to the SEC-MALS. Since the peak

width is approximately constant, increasing the injection volume results in higher peak concentrations, facilitating the identification of fast dynamic equilibria.

The increase of injected volume clearly affects the equilibrium of Sample 2 (Figure 4). As a higher insulin concentration is achieved, the equilibrium of the monomer-dimer is shifted towards dimer. The smallest species observed when 50 μL is injected is monomer while the corresponding smallest species is dimer when 200 μL is injected. The hexamer fraction is unaffected and shows no tendency to form larger oligomers.

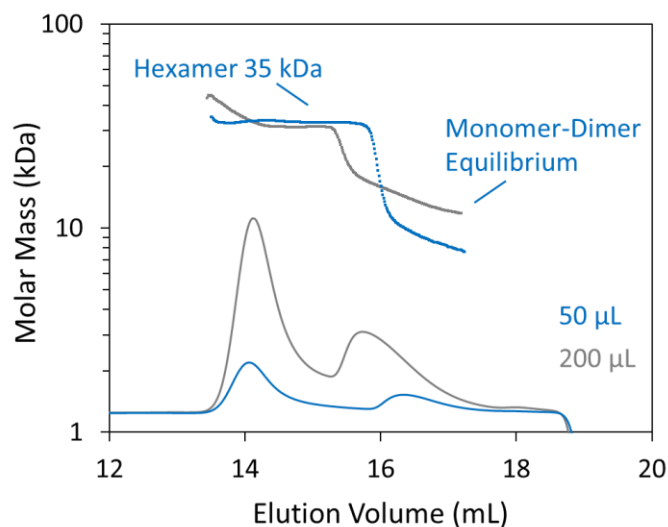


Figure 4: Plot of molar mass vs. time of Sample 2. The UV signal @ 280 nm is plotted as an overlay.

In the case of Sample 3 there are basically no effect of varying the injection volume (Figure 5). In presence of 0.3 mM zinc the system is “locked” on hexamer and identical molar masses are detected at both concentrations (50 and 200 μL injected). Obviously, the self-association behavior of the hexamer is significantly different from that of the monomer.

Conclusions

Multi-angle light scattering proves to be the method of choice for the examination of association phenomena not only for large proteins such as antibodies, but also for peptide molecules with lower molecular masses. Since the use of proteins and peptides for therapeutic purposes is permanently increasing, the application of light scattering as a powerful method of molecular characterization will also become more and more important.

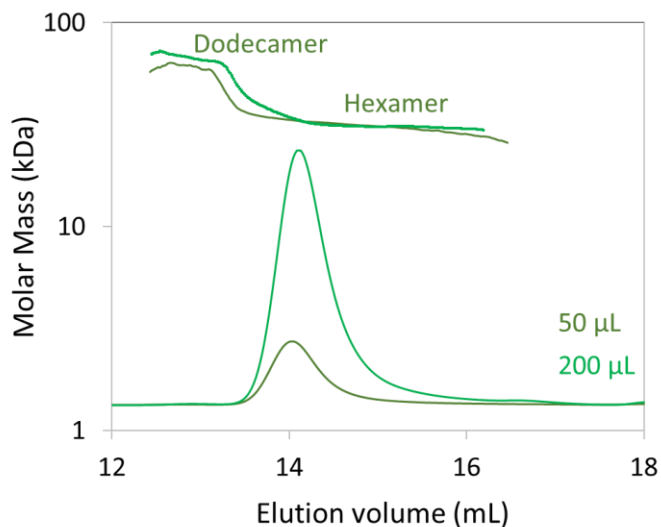
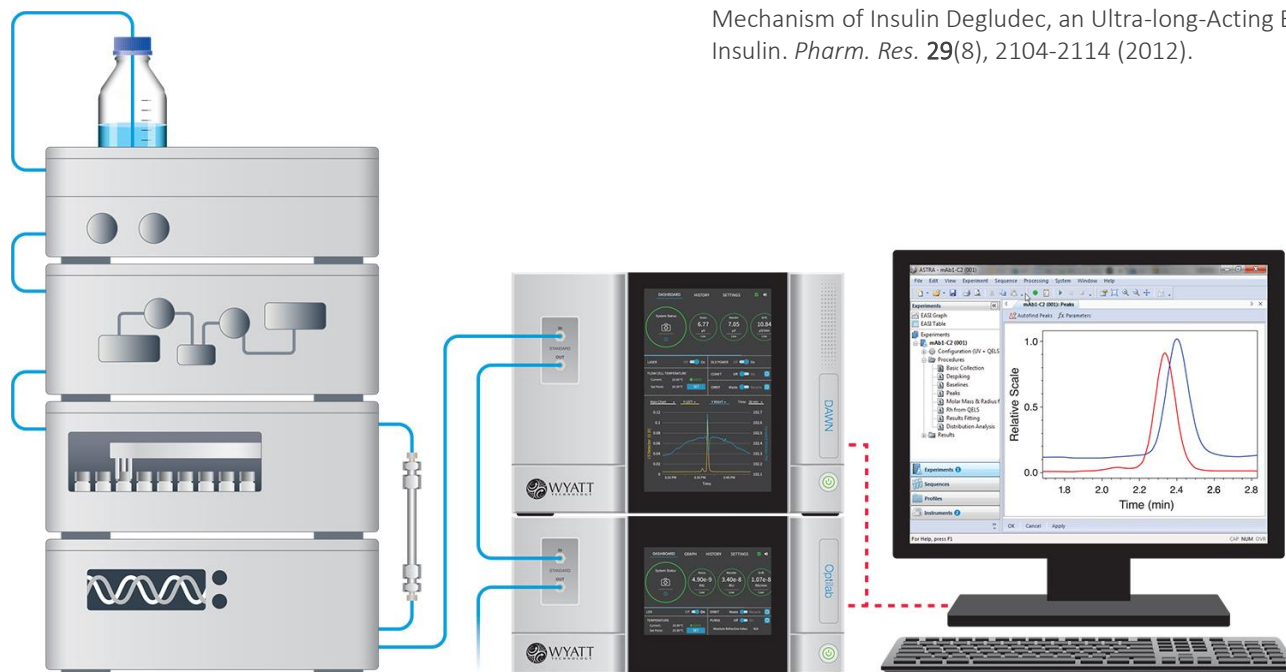


Figure 5: Plot of molar mass vs. time of Sample 3. The dRI signal is plotted as an overlay (the UV signal is saturated due the high sample load).



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