GLYCOWORKS RapiFluor-MS QUICK START PROTOCOL

STREAMLINED PROTOCOL - 24 SAMPLE (3 x 8 FORMAT)

- Set heat blocks to at least 90 °C and 50 °C.
- Protocol is based on 1.5 mg/mL glycoprotein starting concentration.



STEP 1: Rapid Deglycosylation

- Reconstitute 1 vial of the Intact mAb Mass Check Standard (1 mg/vial) in 670 μL 18.2 MΩ water to create a 1.5 mg/mL solution.
 - Note: For glycoproteins with a formulation buffer containing nucleophiles or anionic reagents (e.g., His, Gly, Tris, PO $_s^*$), if a Glycan C_mAX Column is applied for LC separation, it is highly recommended to desalt the sample with water prior to Step 1.
- 2. Prepare 3% (w/v) RapiGest™ SF by dissolving 3 mg of RapiGest SF Surfactant in 60 µL of Rapid Buffer and 40 µL of water, vortex.
- 3. Dilute PNGase F enzyme (35 μ L) with 220 μ L water for a total of 255 μ L.
- Add 10 μL of 1.5 mg/mL glycoprotein into the provided tube.
- 5. Add 10 µL of buffered 3% (w/v) RapiGest SF solution to above tube, aspirate to mix.
- 6. Heat at least to 90 °C for 3 minutes.
- 7. Cool at room temperature for 3 minutes.
- 8. Add 10 μ L Rapid PNGase F and aspirate to mix.
- 9. Incubate at 50 °C for 5 minutes.
- 10. Cool at room temperature for 3 minutes.



STEP 2: Rapid Labeling of Glycosylamines

- 1. Add 110 µL of anhydrous DMF directly to one vial of 9 mg of *Rapi*Fluor-MS™ Reagent. Mix to solubilize.
- 2. Add 10 μL of the *Rapi*Fluor-MS solution to the deglycosylation mixture and aspirate to mix.
- 3. Allow the labeling to proceed at room temperature for 5 minutes.
- Dilute the reaction with 360 μL of acetonitrile (ACN) and aspirate to mix.





STEP 3: HILIC Cleanup of Labeled Glycosylamines

- Set up a GlycoWorks™ HILIC µElution Plate and add in shims or spacer and waste tray.
- 2. Condition wells by adding 200 μ L of water per well.
- 3. Equilibrate wells by adding 200 µL 85% ACN.
- 4. Load ACN-diluted samples (~400 μL).
- Wash wells with two (2) 600 µL volumes of 1% formic acid, 90% ACN.
- 6. Replace waste tray with sample collection tray loaded with 600 uL tubes.
- 7. Elute glycans with three (3) 30 µL volumes of SPE Elution Buffer into 600 µL tapered bottom inserts.
- Dilute SPE eluate with 310 µL of the GlycoWorks SPE Diluent (DMF/ACN). Aspirate to mix.
 - Note: For a Glycan $C_{_{18}}$ AX separation sample, either skip dilution Step 8 or dilute with 310 μ L of water.
- 9. Cap the tubes with pre-slit cap mats.
- ► For the complete Care and Use Manual, visit waters.com and search 715004903EN.
- ► For more details on this method, download Application Notes 720005506EN and 720007038EN.