

Chiral and Achiral Profiling of a Pesticide Formulation Using the ACQUITY UPC² System and the ACQUITY QDa Detector

Marian Twohig,¹ Michael O'Leary,¹ John P. McCauley,² and Rui Chen²

¹Waters Corporation, Milford, MA, USA

²Waters Corporation, New Castle, DE, USA

APPLICATION BENEFITS

- Chiral and achiral method development and sample analysis can be performed on the same system.
- Improved enantiomeric and diastereomeric resolution and shorter analysis time compared with normal phase separations resulting in higher sample throughput.
- Reliable and reproducible measurement of the enantiomer and/or diastereomer ratios.
- Detection and chiral resolution of structurally related formulation components.

WATERS SOLUTIONS

ACQUITY® UltraPerformance Convergence Chromatography™ System (UPC²®)

ACQUITY UPLC®

Photodiode Array (PDA) Detector

ACQUITY QDa™ Detector

ACQUITY UPC²™ BEH Column

Empower® 3 Chromatography Data Software

KEY WORDS

Chiral pesticides, UPC², enantiomer, diastereomer, chiral separation, chiral resolution, propiconazole, triazole fungicide, SFC, supercritical fluid chromatography, QDa

INTRODUCTION

Research associated with the development of new agricultural pesticide formulations centers around the design of products that provide highly effective and specific action towards the target organism with reduced application rates.¹ It is estimated that 30% of the pesticides on the market today have optical isomers.^{2,3} However, the desired activities often result from one single enantiomer in the optical isomer mixtures.⁴ It is therefore important to assess the enantiomeric purity of the chiral active ingredients in the formulation.^{1,5} In addition, the detection, characterization, and quantitation of the other components in the formulation are necessary to support product registration.

Liquid chromatography (LC) on chiral stationary phases (CSPs), such as polysaccharide stationary phases including amylose and cellulose, has been the most commonly used chiral separation technique.⁶⁻⁹ More recently, there has been an increasing adoption of supercritical fluid chromatography (SFC) on CSPs for chiral separation.^{10,11} The properties of a supercritical fluid, its high diffusivity and low viscosity in particular, enable high efficiency chiral separations with shorter run times. For example, triazole fungicides, such as propiconazole (structure shown in Figure 1), are a commonly used group of pesticides because of their potent activity against a broad spectrum of crop diseases. Using HPLC, the analysis times for the diastereomeric resolution of propiconazole range from 34 to 50 min.^{1,6-8} Similar resolutions were achieved for propiconazole using SFC, but the analysis times were reduced to 10 min.¹¹

UltraPerformance Convergence Chromatography (UPC²) applies the performance advantages of UPLC® to SFC, combining the use of supercritical CO₂ with sub-2-µm particle columns.^{12,13} UPC² is an orthogonal analytical technique to reversed-phase LC and can be used to solve complex separations challenges.

In this application note, we describe the analytical profiling of a triazole-containing pesticide formulation using UPC² and a combination of UV and mass detection. Waters® ACQUITY QDa Detector is a novel mass detector that can be integrated into existing liquid chromatography configurations in order to increase sensitivity and complement the results obtained when using only UV detectors.

EXPERIMENTAL

Instrumentation

All separations were performed on a Waters ACQUITY UPC² System equipped an ACQUITY UPC² Photodiode Array (PDA) and positive ion electrospray mass spectrometry (MS) using an ACQUITY QDa Detector. Empower 3 FR2 Software was used for data acquisition and processing.

Sample preparation

The authentic pesticide standards were made up in 50:50 acetonitrile/water. 2 grams (g) of the commercially available pesticide formulation was weighed, and 8 mL of 50:50 (v/v) acetonitrile/water was added. The resulting mixtures were sonicated for 10 minutes and the samples were syringe filtered into an autosampler vial using a 0.2- μ m PVDF filter in preparation for sample analysis.

UPC² conditions

Achiral separation

Separation mode:	Gradient
Column:	ACQUITY UPC ² BEH, 3.0 x 100 mm, 1.7 μ m
Co-solvent (B):	Methanol
ABPR:	1990 psi/137 bar
Flow rate:	1.5 mL/min
UV detection:	220 nm
Column temp.:	35 °C
Injection volume:	0.5 μ L

Formulation A achiral gradient conditions:

0 min 3% B, 4 min 30% B, 6 min 30% B,
return to initial conditions.

Chiral separation

Separation mode:	Gradient
Column:	Amylose Chiral, 3.0 x 150 mm, 2.5 μ m
Co-solvent (B):	50:50 2-propanol/ethanol
ABPR:	1990 psi/137 bar
Flow rate:	2.0 mL/min
UV detection:	220 nm
Injection volume:	1 μ L

MS conditions

MS system:	ACQUITY QDa Detector
Ionization mode:	ESI +
Capillary voltage:	0.8 kV
Cone voltage:	10 V
Desolvation temp.:	600 °C
Source temp.:	150 °C
MS scan range:	100 to 600 <i>m/z</i>
Sampling rate:	5 Hz
Make up solvent:	98:2 MeOH/water with 0.1% ammonium hydroxide at 0.3 mL/min

PDA conditions

Detector:	ACQUITY UPC ² PDA
Wavelength range:	210 to 400 nm
Sampling rate:	20 Hz

When MS data is combined with the UV response, it allows the analyst to determine a wider range of analytes in one analytical run with an increased level of confidence.

The formulation sample first underwent achiral separations on a sub-2- μm stationary phase, followed by chiral separations on a 2.5- μm chiral CSP. A minor isomer of the active ingredient (AI) was identified. Further chiral analyses revealed similar chirality between the minor isomer and the AI.

RESULTS AND DISCUSSION

Figure 2 shows the ACQUITY UPC² chromatograms of the propiconazole standard (lower trace) and the formulation sample (top trace) obtained using an ACQUITY UPC² BEH Column. The retention times of peaks 1 and 2 in the formulation sample matched those of the propiconazole standard. These two peaks correspond to the propiconazole diastereomers. It is noted, however, that there were two minor peaks (peaks 3 and 4) observed in the formulation with retention times (t_R) of 2.22 min and t_R 2.26 min, respectively.

The UV spectra of peaks 3 and 4 resemble those for peaks 1 and 2 (Figure 2), indicating their structural similarity. In addition, the four peaks resulted in identical mass spectra with base peaks at m/z 342 and an isotopic pattern characteristic of dichlorinated compounds. The m/z matched the protonated propiconazole.

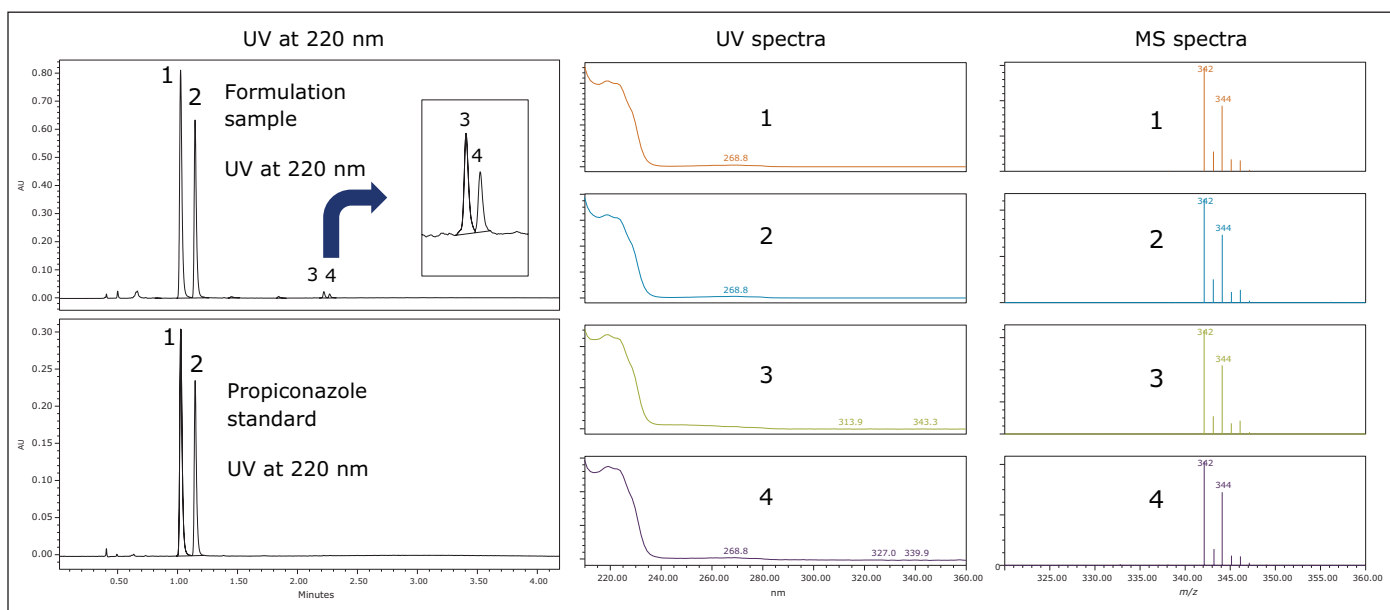


Figure 2. ACQUITY UPC² UV achiral separation of the propiconazole present in the formulation sample and propiconazole standard at 220 nm using the ACQUITY UPC² BEH Column. The UV and MS spectra for peaks 1-4 are also shown.

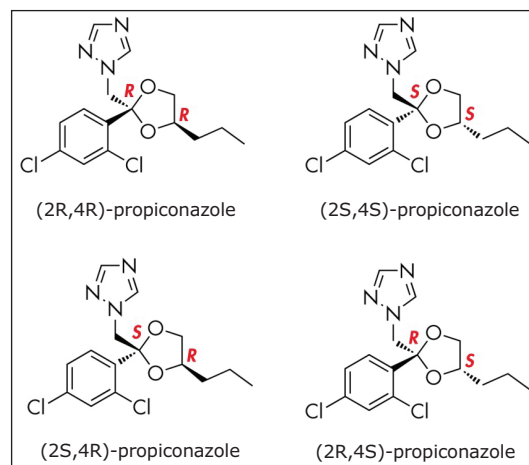


Figure 1. Structures of the propiconazole stereoisomers.

The ACQUITY UPC² System has multi-column switching capabilities and a choice of up to four co-solvents which conveniently allows both achiral and chiral method development and sample analysis to be performed on the same system. The method development process can be completed rapidly due to the shorter analysis times that are possible using this technique.

Figure 3 shows the ACQUITY UPC² chromatograms of propiconazole standard and the formulation sample using gradient separation on an Amylose Chiral Column. The two diastereomer peaks of propiconazole observed in Figure 2 were resolved into four individual peaks (1-4). Interestingly, the two minor isomer peaks in Figure 2 were also resolved into four peaks (5-8) in a comparable manner, indicating a similar chirality for propiconazole and the minor components.

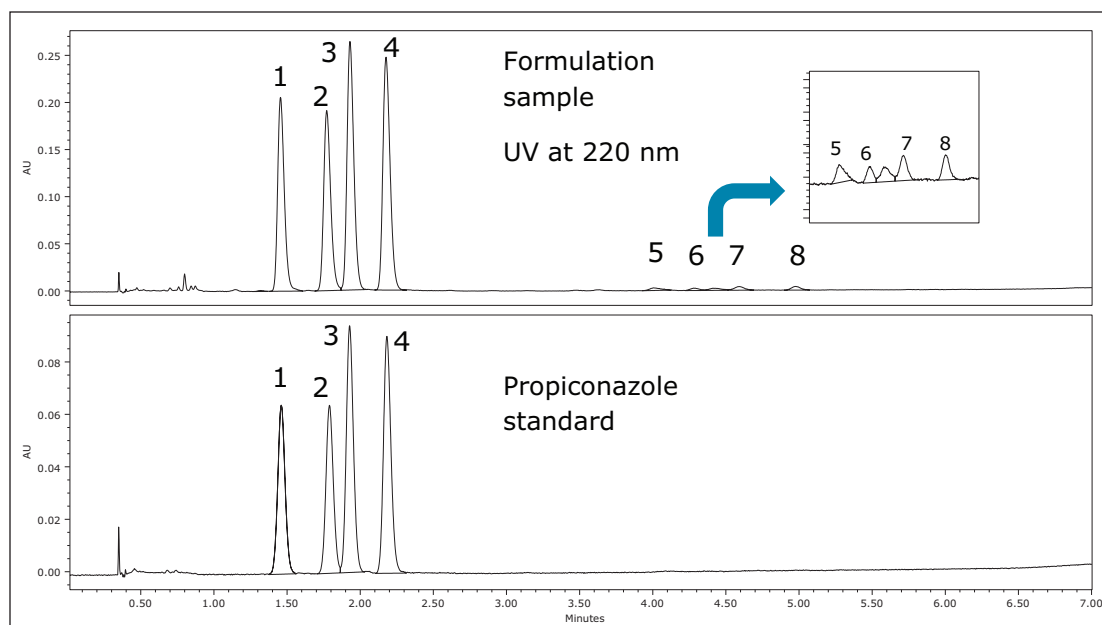


Figure 3. ACQUITY UPC² UV chromatogram at 220 nm showing the chiral resolution of the propiconazole stereoisomers and unknown chiral components in the formulation sample using an Amylose Chiral Column. The propiconazole standard is also shown.

Interrogation of the formulation sample data in Empower Software's mass analysis window showed similar UV and identical MS spectra for all eight peaks (Figures 4 and 5). Empower 3 Software's Mass Analysis window provides a single location to associate chromatographic peaks from all detectors used in the analysis with their corresponding spectra. The spectra from the detected peaks are time-aligned and displayed in a window above the chromatograms.

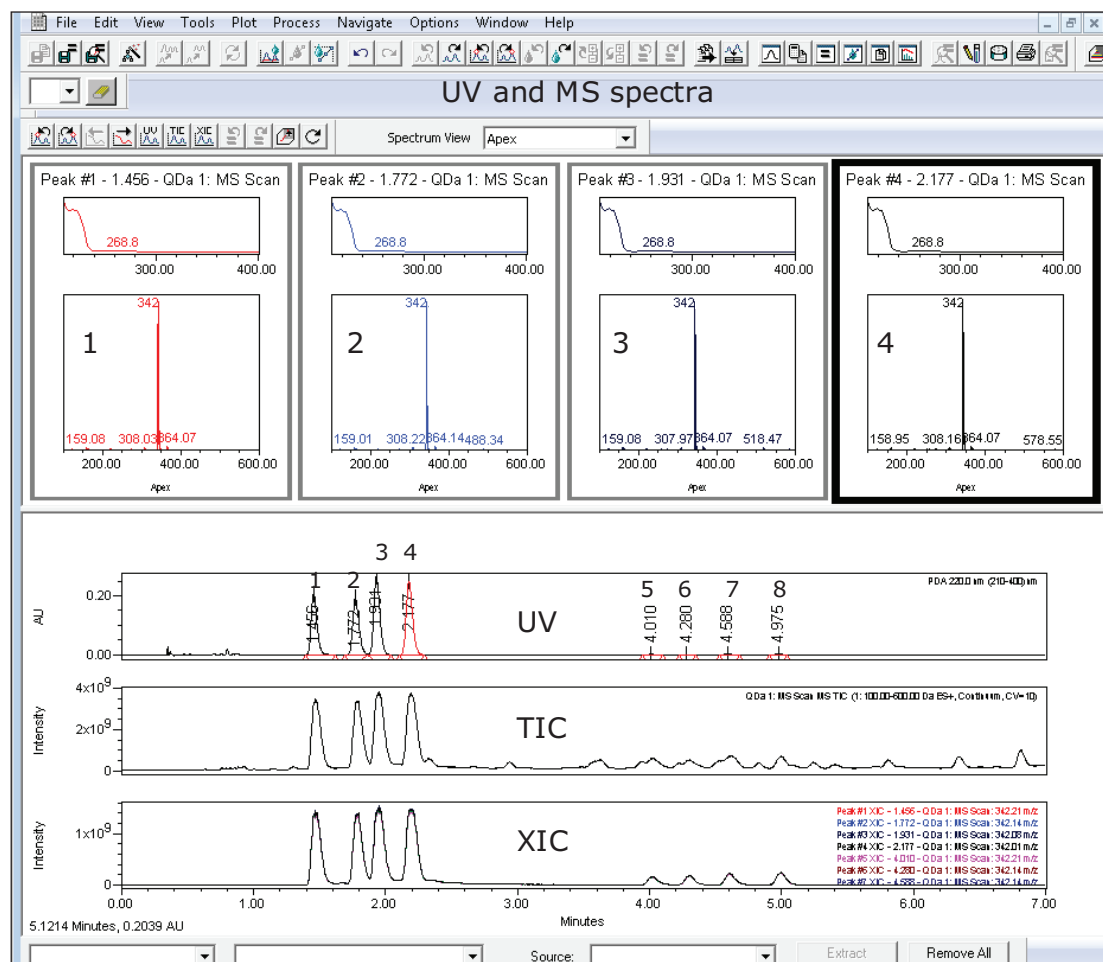


Figure 4. With Empower Software's Mass Analysis window, UV and MS spectra, along with UV and mass chromatograms and extracted ion chromatograms (XIC), can be viewed in a single window.

Matrix components visible in the UV chromatogram and the MS total ion chromatogram (TIC) of the formulation sample are clearly differentiated from the isomeric peaks of interest using an extracted ion chromatogram (Figure 5). The detection sensitivity and selectivity of the method are improved when using mass detection in combination with UV detection which is used in order to measure the enantiomeric purity of the chiral pesticide.

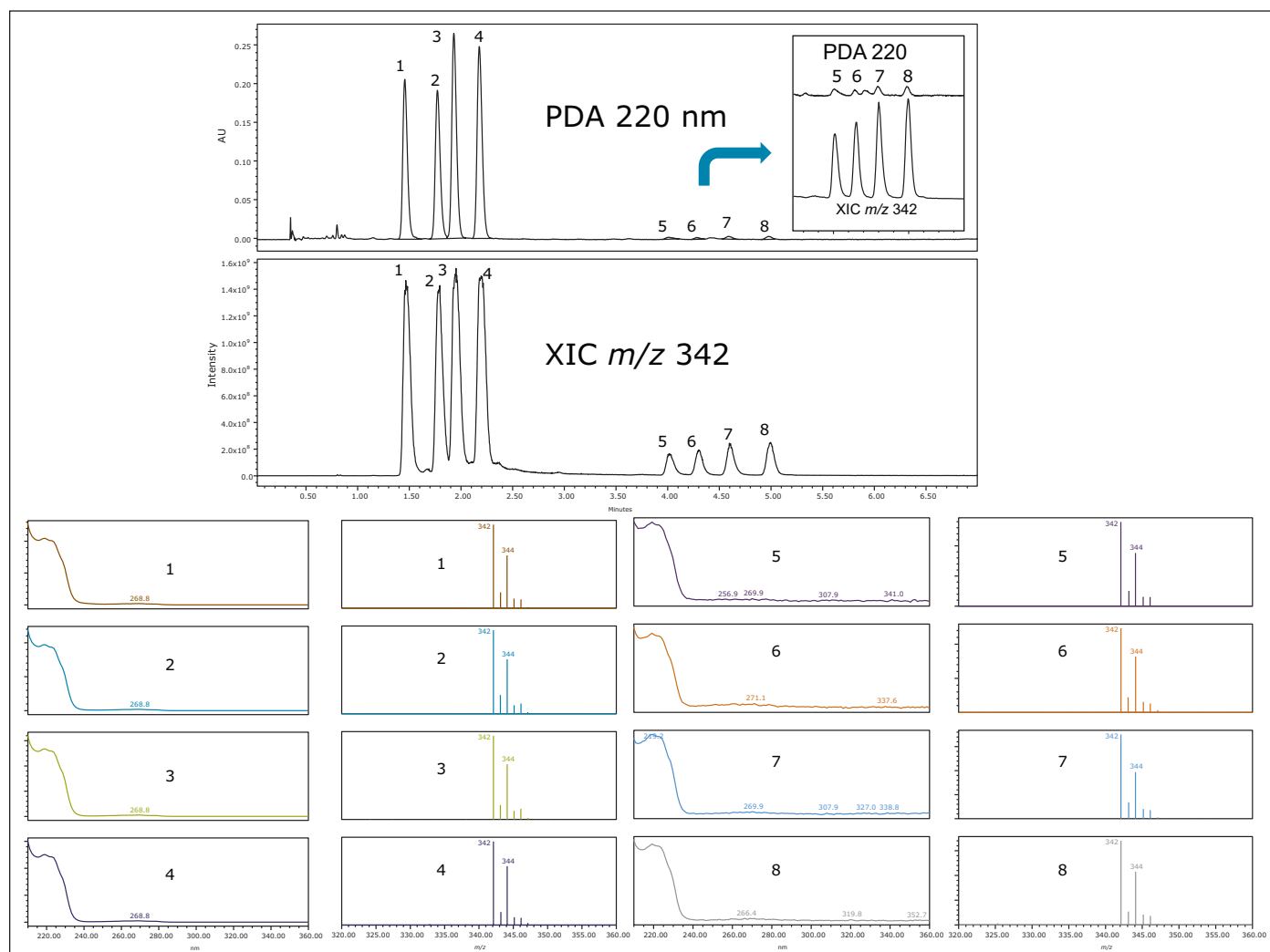


Figure 5. ACQUITY UPC² UV chromatogram at 220 nm showing the chiral resolution of the propiconazole stereoisomers and unknown chiral components in the formulation sample using an Amylose Chiral Column. The XIC of m/z 342 and the UV and MS spectra for each peak are also shown.

Based on the observations, it is postulated that the minor component is a regioisomer of propiconazole. A regioisomer of propiconazole originating from one of the nitrogens on the triazole ring was characterized by Glaser *et al.*¹⁴ Further experiments to isolate this compound for positive identification are currently underway.

CONCLUSIONS

In this study the achiral screen of the formulated pesticide products showed that the minor components detected using UV and mass detection had similar structural characteristics to the AI, propiconazole. The minor components had the same m/z and shared the same isotopic pattern as the triazole fungicide, AI. Subsequent chiral resolution of the propiconazole in the formulation in combination with simultaneous mass and UV detection provided valuable spectral information which allowed the minor components to be characterized as probable stereoisomers.

The addition of mass detection as a complementary analytical detection technique enhances confidence in compound detection and identification. The ACQUITY QDa Detector provides a cost-effective means to make mass detection part of the routine analysis in laboratories that have previously relied on less selective detectors.

The ACQUITY UPC² System has column switching capabilities so that both chiral and achiral columns can be used with a choice of four co-solvents that are compatible with MS analysis. The chiral and achiral method development and analysis can be performed on the same system. These methods use supercritical CO₂ as the primary mobile phase. The need to use large volumes of potentially hazardous solvents is reduced compared to normal phase separations. Consequently the cost associated with solvent waste disposal can also be reduced.

The ACQUITY UPC² System allows high efficiency separations that can increase sample throughput compared to traditional normal-phase separations.^{1,6-8} The diastomeric resolution of propiconazole using UPC² took place in less than 3 minutes, which is at least 10 times faster than normal phase methods reviewed in the literature.

The study of enantioselective properties has previously been a challenge due to the difficulty in resolving chiral compounds. The benefit of having faster analytical methods to resolve

chiral compounds is that critical information pertaining to their stereoselective behavior can be obtained more rapidly leading to increased laboratory productivity.

References

1. VPerez-Fernandez, M A Garcia, M L Marina. Chiral separation of agricultural fungicides. *J Chrom A*. 1218: 6561-6582, 2011.
2. B S Sekhon. Chiral pesticides. *J Pestic Sci*. 34,1: 1-12, 2009.
3. W P Liu. Pesticide Environmental Chemistry. Chemical Industry Press, Beijing, China. 341-343, 2006.
4. A Williams. Opportunities for chiral agrochemicals. *J Pestic Sci*. 46: 3-9, 1996.
5. EU Commission Regulation No 544/2011. (2011) [cited 2013 October 20]. Available from: <http://eurlex.europa.eu/LexUriServ/LexUriServ.do?uri=OJ:L:2011:155:0001:0066:EN:PDF>
6. H Zhang, M Quian, X Wang, X Wang, H Xu, Q Wang, M Wang. HPLC-MS/MS enantioseparation of triazole fungicides using polysaccharide based stationary phases. *J Sep Sci*. 35: 773-781, 2012.
7. Y Zhou, L Li, K Lin, X Zhu, W Liu. Enantiomer separation of triazole fungicides by high performance liquid chromatography. *Chirality*. 21: 421-427, 2009.
8. J Ye, J Wu, W Liu. Enantioselective separation and analysis of chiral pesticides by high-performance liquid chromatography. *Trends in Analytical Chemistry*. 28, 10: 1148-1163, 2009.
9. P Wang, S Jiang, D Liu, P Wang, Z Zhou. Direct enantiomeric resolutions of chiral triazole pesticides by high-performance liquid chromatography. *J Biochem Biophys Methods*. 62: 219-230, 2005.
10. L Jin, W Gao, H Yang, C Lin, W Liu. Enantiomeric resolution of five chiral pesticides on a chiralpak IB-H column by SFC. *J Chrom Sci*. 49: 739-743, 2011.
11. L Toribo, M J del Nozal, J L Bernal, J J Jimenez, C Alonso. Chiral separation of some triazole pesticides by supercritical fluid chromatography. *J Chrom. A*. 1046: 249-253, 2004.
12. A Grand-Guillaume, J L Perrenoud, D Guillarme. Comparison of ultra-high performance supercritical fluid chromatography and ultra-high performance liquid chromatography for the analysis of pharmaceutical compounds. *J Chromatogr A*. 1266: 158, 2012.
13. C Hamman, W Mengling, I Aliagas, D F Ortwine, J Pease, DE Schmidt Jr Victorino. The evaluation of 25 chiral stationary phases and the utilization of sub-2.0 μ m coated polysaccharide chiral stationary phases via super critical fluid chromatography. *J Chrom A*. 1305: 310-319, 2013.
14. R Glaser, I Adin, D Ovadia, E Mendler, M Drouin. Solid-state structure determination and solution-state NMR characterization of the (2R,4R)/(2S,4S)- and (2R,4S)/(2S,4R)-diastereomers of the agricultural fungicide propiconazole, the (2R,4S)/(2S,4R)-symmetrical triazole constitutional isomer, and a ditriazole analogue. *Structural Chem*. 6, 3: 145-156, 1995.

Waters

THE SCIENCE OF WHAT'S POSSIBLE.®

Waters, ACQUITY, ACQUITY UPLC, Empower, The Science of What's Possible, UPC² and UPLC are registered trademarks of Waters Corporation. ACQUITY QDa, ACQUITY UPC² and UltraPerformance Convergence Chromatography are trademarks of Waters Corporation. All other trademarks are the property of their respective owners.

©2014 Waters Corporation. Produced in the U.S.A. March 2014 720004977EN AG-PDF

Waters Corporation
34 Maple Street
Milford, MA 01757 U.S.A.
T: 1 508 478 2000
F: 1 508 872 1990
www.waters.com